



Temporal order of clinical and biomarker changes in familial frontotemporal dementia

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Unlike familial Alzheimer's disease, we have been unable to accurately predict symptom onset in presymptomatic familial frontotemporal dementia (f-FTD) mutation carriers, which is a major hurdle to designing disease prevention trials. We developed multimodal models for f-FTD disease progression and estimated clinical trial sample sizes in *C9orf72*, *GRN* and *MAPT* mutation carriers. Models included longitudinal clinical and neuropsychological scores, regional brain volumes and plasma neurofilament light chain (NfL) in 796 carriers and 412 noncarrier controls. We found that the temporal ordering of clinical and biomarker progression differed by genotype. In prevention-trial simulations using model-based patient selection, atrophy and NfL were the best endpoints, whereas clinical measures were potential endpoints in early symptomatic trials. f-FTD prevention trials are feasible but will likely require global recruitment efforts. These disease progression models will facilitate the planning of f-FTD clinical trials, including the selection of optimal endpoints and enrollment criteria to maximize power to detect treatment effects.

Frontotemporal dementia (FTD), marked by impairments in behavior, language and sometimes motor function, is a common form of early-onset dementia¹. Approximately 20–30% of FTD is caused by autosomal dominant mutations (familial, or f-FTD), usually in one of three genes: chromosome 9 open reading

frame 72 (*C9orf72*), progranulin (*GRN*) or microtubule-associated protein tau (*MAPT*)². FTD is uniformly fatal, and there are no approved therapies; however, a growing number of new treatments targeting *C9orf72*, *GRN* and *MAPT* are moving into clinical trials^{3,4}. Experience from Alzheimer's disease (AD), spinal muscular

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atrophy⁵ and amyotrophic lateral sclerosis (ALS)⁶ suggests treating FTD will be most successful if treatment is initiated early in the disease course, ideally before the onset of symptoms. Such a disease prevention approach has been implemented in the Dominantly Inherited Alzheimer's Network Trials Unit (DIAN-TU; <https://dian.wustl.edu/our-research/clinical-trial/>) platform clinical trial for dominantly inherited AD (DIAD) by including presymptomatic mutation carriers⁷. Prevention trials in DIAD have also been facilitated by fluid and molecular positron emission tomography imaging biomarkers that allow for the measurement of treatment-related changes in AD pathologies and neurodegeneration⁸.

There are many challenges to performing f-FTD clinical trials⁹. Although the clinical manifestations of the f-FTD mutations are similar, the biology and neuropathology associated with *C9orf72*, *GRN* and *MAPT* mutations are vastly different². Unlike AD¹⁰, little is known about the ontogeny of biomarker and clinical changes in f-FTD that could be used to determine enrollment criteria or identify the best clinical trial endpoints at different disease stages. Also, the age at which symptoms present is variable even within a family (for example, onset in the 30s versus the 70s in one family)¹¹, making it difficult to identify the individuals in late presymptomatic stages most likely to benefit from therapies. For instance, in *GRN*, familial age of onset only explains 14% of the variability in individual age at symptom onset¹².

f-FTD is rare, with only hundreds of mutation carriers for the rarest mutations, *GRN* and *MAPT*, known to exist worldwide¹². Therefore, to prepare for f-FTD trials, the Frontotemporal Dementia Prevention Initiative (FPI; www.thefpi.org), an international collaboration focused on organizing f-FTD prevention trials, combined data from the two largest f-FTD natural history studies worldwide: ALLFTD in North America (www.allftd.org) and GENFI in Europe and Canada (www.genfi.org)¹³. In rare neurogenetic diseases such as f-FTD, the US Food and Drug Administration has promoted the use of innovative approaches such as disease progression models (DPMs) for selecting clinical trial endpoints, determining enrollment criteria and analyzing the effects of interventions that might lead to deviations from expected disease progression¹⁴, and such models have been used successfully in DIAN-TU⁷. We developed Bayesian DPMs that jointly model the best known measures of f-FTD global clinical status, neuropsychological performance, brain volume and active neurodegeneration (plasma NfL) to model latent disease age (DA), which forecasts presymptomatic mutation carriers' proximity to symptom onset and enables comprehensive quantification of disease progression. We then conducted simulations of prevention and early symptomatic treatment trials, exploring the use of DA, plasma NfL and clinical measures as inclusion criteria to prioritize the recruitment of presymptomatic participants toward those most likely to exhibit measurable disease progression during a trial.

Results

Subject characteristics. Demographic and clinical data are presented in Table 1 and Supplementary Table 1. Of the 796 mutation carriers, *C9orf72* was the most common mutation (43.6%), followed by *GRN* (35.3%) and *MAPT* (21.1%). Across all three genetic groups, most participants were presymptomatic (Clinical Dementia Rating Scale plus National Alzheimer's Coordinating Center Frontotemporal Lobar Degeneration Module (CDR+NACC-FTLD) Global = 0, 54.4%). Most symptomatic participants presented with behavioral variant FTD (bvFTD; 68.6%), followed by primary progressive aphasia (PPA; 12.7%), which was driven largely by *GRN* (33.8% of symptomatic *GRN*). The average number of visits per mutation carrier was 2.1 (standard deviations (s.d.) = 1.1). The models incorporated 412 noncarrier family controls. A subset of participants had available NfL ($n = 981$, 1,948 observations) and magnetic resonance imaging (MRI) data ($n = 882$, 1,896 observations).

DPMs. Overview. When ALLFTD and GENFI participants were modeled separately, rates of progression were very similar between consortia on all measures (Fig. 1 and Supplementary Fig. 1); subsequent models combined all participants. To understand the temporal ordering of biomarker and clinical changes, disease progression curves were graphed in relation to predicted DA (Fig. 2).

MRI and plasma NfL. In *C9orf72*, MRI was the first biomarker to change (Figs. 2 and 3, Extended Data Fig. 1, Table 2 and Supplementary Tables 2–5), with visual inspection of the DPM curves, suggesting that brain volumes deviate from controls up to 40 years before expected onset. Thalamic volume in *C9orf72* was significantly lower than controls in the –40 to –10 epoch, with the largest effect size of all regions of interest (ROIs) (Extended Data Fig. 1 and Supplementary Table 5). Voxelwise quantification also underscored the early thalamic involvement (Extended Data Fig. 2 and Supplementary Fig. 2). In addition to the thalamus, most ROIs were smaller than controls (Extended Data Fig. 2 and Supplementary Table 5) and other mutation carriers (Supplementary Table 6) in the –40 to –10 epoch. In the –10 to 0 epoch, the temporal lobe showed the largest effect size (Extended Data Fig. 1), and it was the first ROI to deviate from controls (Supplementary Table 4; deviated at DA = –6.1; 95% CI, –9.4, –3.2) by 1 s.d., followed closely by parietal (DA = –6.1; 95% CI, –9.2, –3.2) and frontal (DA = –4.9; 95% CI, –7.5, –2.7) lobes. The largely overlapping credible intervals indicate these differences in temporal ordering are not statistically significant. The longitudinal rate of volume loss was relatively stable across the across epochs in *C9orf72* compared to the other genetic groups (Supplementary Table 4). Visual inspection of the DPM curves suggested mean NfL values in *C9orf72* begin to deviate from controls approximately 30 years before estimated onset. NfL levels in *C9orf72* were significantly higher than controls in all DA epochs and became elevated 1 s.d. above controls 3 years before estimated onset (95% CI, –0.7, –5.8).

In *GRN*, visual inspection suggested NfL begins to deviate from controls about 15 years before symptom onset, followed by MRI 5–10 years before onset (Fig. 2 and Supplementary Fig. 1). Baseline plasma NfL concentrations in *GRN* were significantly elevated relative to controls in all DA epochs (Fig. 3 and Supplementary Table 5) and elevated compared to all other genetic groups in the symptomatic phase (Supplementary Table 6). NfL concentrations became elevated by 1 s.d. compared to controls 4.9 years before onset (95% CI, –3.4, –7). *GRN* also displayed the most rapid rates of NfL increase in the symptomatic epoch (Figs. 1 and 2 and Supplementary Table 4). The frontal and temporal lobes were the first brain regions to differ from controls by 1 s.d. in the DPM (–1.1 and –1.2 years before estimated onset, respectively). The insula was significantly atrophied compared to controls in the –40 to –10 epoch (Extended Data Fig. 1 and Supplementary Table 5), and all ROIs had smaller mean volumes than controls in the –10 to 0 epoch, except the striatum ($P = 0.057$). In the symptomatic stage, volume loss in all ROIs was more rapid than the other genetic groups, with the frontal, temporal, medial temporal, insular and striatal ROIs losing volume most rapidly (Supplementary Fig. 3 and Supplementary Table 4).

Medial temporal lobe (MTL) atrophy was the first observed biomarker change in *MAPT*, diverging from controls ~10 years before symptoms based on visual inspection (Fig. 2), and reaching 1 s.d. below controls 1.8 years before onset (95% CI, –3.2, –0.5). The MTL was the only region with significant volume loss compared to controls in the presymptomatic phase (Extended Data Fig. 1 and Supplementary Table 5). The remaining temporal regions and insula were the next regions to become atrophied by 1 s.d. compared to controls (Supplementary Table 4), with overlapping credible intervals. In the symptomatic stage, frontal, temporal and medial temporal, insular and striatal regions showed the greatest degree

Table 1 | Characteristics of the study participants

Characteristic	All carriers	<i>C9orf72</i> ⁺	<i>GRN</i> ⁺	<i>MAPT</i> ⁺	Noncarriers	<i>P</i>	Pairwise comparisons
Sample size	796	347	281	168	412		
ALLFTD sample size	275	127	68	80	161		
GENFI sample size	521	220	213	88	251		
Age (yr), mean (s.d.)	50.2 (13.9)	51.2 (13.7)	52.2 (13.7)	44.9 (13.3)	45.9 (13.0)	<0.001	(NC = <i>MAPT</i>) < (<i>C9</i> = <i>GRN</i>)
Female, <i>n</i> (%)	447 (56.1)	188 (54.2)	167 (59.4)	92 (54.8)	239 (58.0)	0.51	
Education (yr)	14.4 (3.2)	14.5 (3.0)	14.2 (3.4)	14.7 (3.0)	14.8 (2.9)	0.07	
Visits (total number), mean (s.d.)	2.1 (1.1)	2.0 (1.0)	2.1 (1.1)	2.5 (1.2)	2.2 (1.1)	<0.001	(<i>C9</i> = <i>GRN</i> , NC = <i>GRN</i> , <i>C9</i> < NC) < <i>MAPT</i>
1	292	135	114	43	137		
2	233	120	68	45	106		
3	158	53	57	48	118		
≥4	113	39	42	32	51		
Total number of observations	1,695	690	592	413	910		
Follow-up length (if >1 visit) (yr), mean (s.d.)	2.0 (0.9)	1.9 (0.9)	2.1 (0.9)	2.2 (0.9)	2.2 (0.8)	<0.001	<i>C9</i> < (<i>GRN</i> = <i>MAPT</i> = NC)
Race, <i>n</i> (%)							
White	776 (97.5)	342 (98.6)	274 (97.5)	160 (95.2)	404 (98.0)	0.11	
People of color ^a	19 (2.4)	4 (1.2)	7 (2.5)	8 (4.8)	6 (1.5)		
Unknown	1 (0.1)	1 (0.3)	0	0	2 (0.5)		
CDR+NACC-FTLD Global, <i>n</i> (%)							
0	433 (54.4)	171 (49.3)	168 (59.8)	94 (56.0)	412 (100%)	0.03 ^b	<i>C9</i> < <i>GRN</i> , <i>C9</i> = <i>MAPT</i> , <i>GRN</i> = <i>MAPT</i>
0.5	127 (16.0)	61 (17.6)	39 (13.9)	27 (16.1)	NA	0.45	
≥1	236 (29.7)	115 (33.1)	74 (26.3)	47 (28.0)	NA	0.16	
Estimated years since onset cMedian (IQR)	4.4 (4.7)	5 (4.7)	2.7 (2.4)	6 (7.8)	NA	<0.001	<i>GRN</i> < <i>C9</i> , <i>GRN</i> < <i>MAPT</i> , <i>C9</i> = <i>MAPT</i>
Symptomatic diagnoses, <i>n</i> (%)							
bvFTD	162 (68.6)	85 (73.9)	38 (51.4)	39 (83.0)	NA	<0.001	<i>GRN</i> < (<i>C9</i> = <i>MAPT</i>)
PPA	30 (12.7)	4 (3.5)	25 (33.8)	1 (2.1)	NA	<0.001	(<i>C9</i> = <i>MAPT</i>) < <i>GRN</i>
CBS	2 (0.9)	-	2 (2.7)	-	NA	0.13	
PSP	3 (1.3)	1 (0.9)	1 (1.4)	1 (2.1)	NA	0.78	
ALS	4 (1.7)	4 (3.5)	-	-	NA	0.14	
FTD-MND	11 (4.7)	11 (9.6)	-	-	NA	0.002	(<i>GRN</i> = <i>MAPT</i>) < <i>C9</i>
MCI	4 (1.7)	2 (1.7)	1 (1.4)	1 (2.1)	NA	1.0	
AD dementia	5 (2.1)	1 (0.9)	3 (4.1)	1 (2.1)	NA	0.35	
Other ^d	4 (1.7)	3 (2.6)	1 (1.4)	1 (2.1)	NA	NA	
Missing	9 (3.8)	4 (3.5)	2 (2.7)	3 (6.4)	NA	NA	

Demographics were calculated using baseline values. Demographic variables and other participant characteristics were compared across genetic groups and controls using regression with pairwise group contrasts for most variables. Sex, race, CDR+NACC-FTLD and diagnostic categories were compared using chi-squared with Bonferroni-adjusted pairwise comparisons when the omnibus test was significant. For chi-squared tests in which any bins were <10, the Fisher's exact test was used. All tests were two sided. Symptomatic clinical diagnoses are displayed in those with a CDR+NACC-FTLD Global ≥1. ^aDue to the small number of participants of color in this sample, a single bin was used to protect participants' identities. ^bControls were not included in pairwise comparisons for CDR+NACC-FTLD. ^cMedian (interquartile range) of baseline values for symptomatic cases based on clinician-reported age of onset. ^dOther diagnoses include dementia not otherwise specified (*n* = 2) or the clinician marked 'other' without entering additional information. CBS, corticobasal syndrome; PSP, progressive supranuclear palsy syndrome; ALS, amyotrophic lateral sclerosis; MND, motor neuron disease; MCI, mild cognitive impairment; NA = not applicable; NC = noncarrier.

of cross-sectional atrophy (Extended Data Fig. 1, Supplementary Fig. 4 and Supplementary Table 5). Longitudinally, the MTL, followed by the remainder of the temporal lobe, striatum and insular regions, were the regions to lose volume most rapidly in the symptomatic phase (Supplementary Table 4). NfL levels began to diverge from controls closer to symptom onset in *MAPT* than *C9orf72* or *GRN*, with mean values showing significant elevations during the symptomatic but not presymptomatic epochs (Extended Data

Fig. 1 and Supplementary Tables 5 and 6), and average values did not reach 1 s.d. above controls until 4.6 years after estimated symptom onset (95% CI, 7.1, 2.4).

We conducted a voxelwise sensitivity analysis in each DA epoch to complement the coarse-grained ROIs used in the DPMs and to illustrate the findings were not dependent on the DPMs. Results of this sensitivity analysis (Extended Data Fig. 2 and Supplementary Figs. 2–4) supported the patterns observed using ROIs.

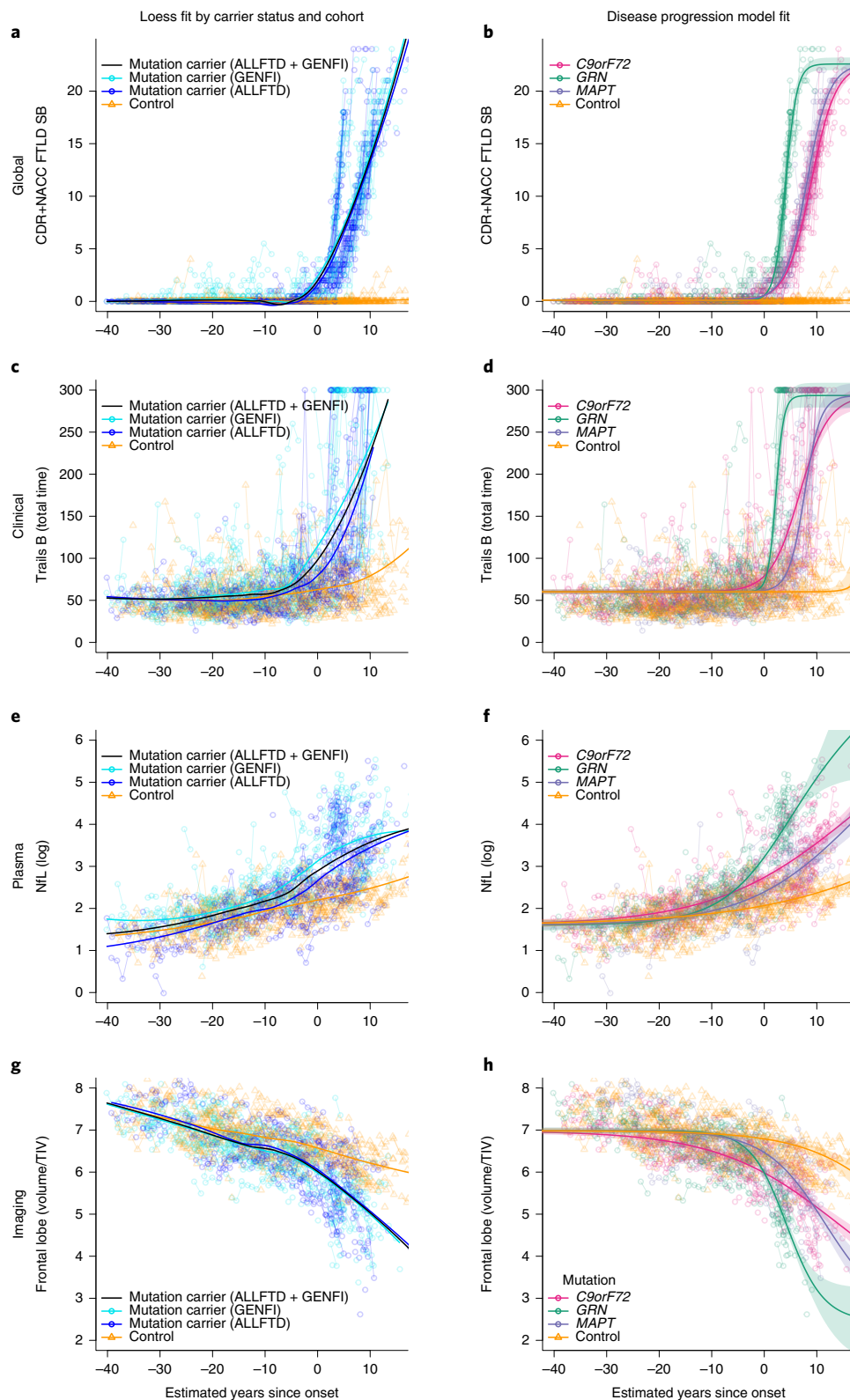


Fig. 1 | Raw data points overlaid on model estimated fit. a,c,e,g, Raw data points for mutation carriers (blue) and noncarrier controls (gold) are presented for several representative measures as a function of model estimated DA, with a loess (locally estimated scatterplot smoothing) fit to each group displayed using thick solid lines. In these panels, mutation carriers are color coded based on whether they were enrolled through ALLFTD or GENFI. These panels highlight the consistency in progression regardless of cohort. **b,d,f,h**, Raw data points are colored by mutation as a function of DA. In these panels, the overall fit for each group was derived from the Bayesian DPM and is displayed using thick solid lines. Shaded areas indicate the 95% credible interval of the estimate. Age-related changes in controls are observed in panels **c-h**. Figures for all modeled measures are included in Supplementary Fig. 1. CDR+NACC-FTLD SB, Clinical Dementia Rating Scale plus National Alzheimer's Coordinating Center's Frontotemporal Lobar Degeneration Module Sum of Boxes; Trails B, Trail Making Test, Part B (total time displayed in seconds); NFL (log), log-transformed plasma NFL; TIV, total intracranial volume.

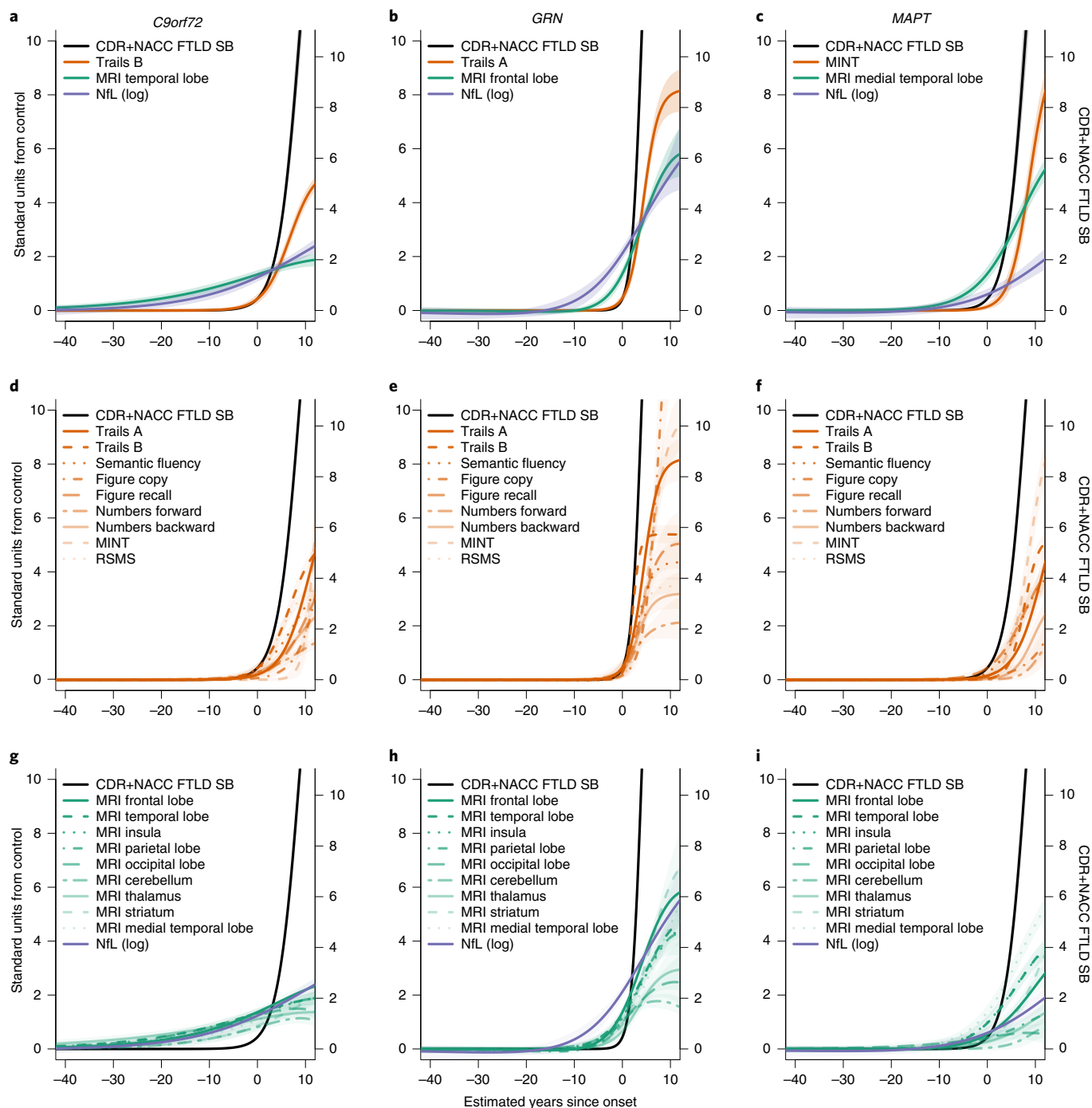


Fig. 2 | Temporal ordering of clinical and biomarker changes in f-FTD. These figures display the empirically derived model-estimated curves in each genetic group. In all figures, model estimated time from onset (years) is on the x axis. The left y axis indicates the number of standard deviations of abnormality compared to controls. The right y axis indicates CDR+NACC-FTLD Box Score units to provide a context for understanding the degree of clinical impairment associated with changes in the other biomarkers and to provide a raw estimate corresponding to the standardized CDR+NACC-FTLD Box Score (black line). **a–c**, Mean curves for the CDR+NACC-FTLD Box Score, NfL and a selected imaging and clinical measure for each genetic group, based on which measure is first elevated by one standard deviation from controls and the measure's rate of longitudinal progression. **d–i**, All clinical, imaging and fluid biomarkers. The shaded areas indicate the 95% credible interval of the estimate. These figures suggest brain atrophy and elevations in NfL levels are detectable before symptom onset and that each mutation shows a unique cascade of biomarker changes. MINT, Multilingual Naming Test; RSMS, Revised Self-Monitoring Scale; Stand, standard.

Global ratings and clinical measures. Visual inspection of the curves revealed a rapid CDR+NACC-FTLD-SB increase after symptom onset, and all genetic groups had cross-sectional elevations in CDR+NACC-FTLD-SB before symptom onset (Fig. 3 and

Supplementary Table 5); note that statistical comparisons of this measure should be interpreted with caution given that controls were defined as having a baseline CDR+NACC-FTLD = 0 (as is typical in most clinical dementia research studies) and thus have no variance

Domain	Measure	Mutation	DA epoch		
			-40 to -10 YSO	-10 to 0 YSO	0+ YSO
Global	CDR+NACC FTLD SB	<i>C9orf72</i>	0.07	0.10	0.41
		<i>GRN</i>	0.06	0.09	0.52
		<i>MAPT</i>	0.07	0.15	0.47
Clinical	Trails B	<i>C9orf72</i>	0.01	0.07	0.32
	Trails A	<i>GRN</i>	0.02	0.04	0.29
	MINT	<i>MAPT</i>	0.00	0.02	0.40
Plasma	NfL (log)	<i>C9orf72</i>	0.05	0.22	0.28
		<i>GRN</i>	0.04	0.15	0.69
		<i>MAPT</i>	0.00	0.00	0.28
Imaging	Temporal lobe	<i>C9orf72</i>	0.08	0.34	0.27
	Frontal lobe	<i>GRN</i>	0.00	0.08	0.41
	Medial temporal lobe	<i>MAPT</i>	0.02	0.07	0.57

Fig. 3 | Comparison of mutation carriers with controls at three epochs of DA.

Cross-sectional baseline differences between mutation carriers and controls are presented as effect sizes (omega squared). Bolded cells indicate statistical significance ($P < 0.05$). Comparisons in which mutation carriers are more impaired than controls at an omega squared > 0.00 are colored, with darker shades illustrating larger effect sizes. CDR+NACC-FTLD SB scores and log-transformed NfL levels are presented for all genetic groups. Clinical and imaging measures were selected for each genetic group based on how early they deviated from controls in the DPM and rate of longitudinal progression. Note that statistical comparisons for the CDR+NACC-FTLD SB should be interpreted with caution given that controls were defined as having a baseline CDR+NACC-FTLD = 0 and thus have no variance due to this selection process. A similar figure including all modeled measures can be found in the extended data figures (Extended Data Fig. 1). YSO, years since onset.

due to this selection process. Similar to the MRI results, *GRN* exhibited the most rapid CDR+NACC-FTLD-SB changes following symptom onset (Fig. 2 and Supplementary Table 4). Visual inspections of the curves indicated that neuropsychological and Revised Self-Monitoring Scale impairments relative to controls were generally observed only after symptom onset for all genetic groups (Fig. 2 and Supplementary Table 5), and no measure reached 1 s.d. worse than controls until after symptom onset (Supplementary Table 4). In direct statistical comparison, *C9orf72* expansion carriers performed worse than controls on Trails A and B at all DA epochs (Supplementary Table 5). *GRN* performed worse than controls on Trails A at all epochs, and worse than controls on Trails B in the -10 to 0 epoch. *MAPT* mutation carriers exhibited impairments in the Figure Copy in the -10 to 0 epoch, with a trend toward impairment on the MINT in this epoch. Longitudinally, the most rapid change in the symptomatic stage relative to controls was observed for Trails A and B in *C9orf72*; Trails A, MINT and Benson Copy in *GRN*; and the MINT and Trails B in *MAPT* (Supplementary Table 4).

Raw values were modeled. The same pattern of findings was observed (Supplementary Fig. 5) in a sensitivity analysis adjusting for nuisance covariates (details in Methods).

Patient-level estimates. DA estimates at baseline ranged from -40 to 21. The precision of individual DA estimates depends on the proximity to symptom onset and follow-up duration. In mutation carriers with at least one postbaseline visit who were >10 years from expected onset, the average uncertainty of the DA estimate (95% CI) was ± 14.6 years. For those -10 to 0 years from onset, this uncertainty decreased to ± 5.5 years, and after onset, the average uncertainty of the estimate was ± 0.9 years. To better understand the impact of level of impairment, rate of progression, and model priors (that is, years since onset) on estimated DA, individual patient-level

data were examined (Extended Data Fig. 3). With increasing DA, performance is increasingly impaired across multiple measures, and there is a greater tendency for progressive impairment from baseline to final observations. In those furthest from onset, when most scores tend to be within normal limits, prior information about their age has a large influence on estimated DA. Examining cases that the model estimated to be presymptomatic (DA < 0) despite a CDR+NACC-FTLD-SB > 0 , these participants tend to perform in the average range across other measures and stay stable or show improvements over time.

Application to clinical trials. The DPM curves suggest that clinical trial endpoint selection might differ by genetic group and disease stage (Fig. 2). To explore this further, simulation studies based on the natural history data were conducted to estimate the sample sizes required to measure a 50% reduction in various potential endpoints for 2- and 4-year presymptomatic prevention trials and 1.5- and 2-year early symptomatic treatment trials (Table 3; 1:1 randomized parallel design; details in Methods). Prevention trial designs included only participants with a CDR+NACC-FTLD-Global = 0 at baseline. Simulations explored the use of baseline NfL and DA as additional inclusion criteria to define a high-risk population most likely to show clinical change over the course of the trial, thereby increasing power. Sample size estimates for prevention trials were generally lowest when using biomarkers (NfL or MRI) as the outcome. For example, a 2-year prevention trial requiring a DA within 5 years of onset would require sample sizes of 52 total participants for *GRN* (MRI frontal), 108 for *MAPT* (MRI MTL) and 424 for *C9orf72* (MRI temporal) if MRI is used as an endpoint. Based on the estimated number of eligible participants from the FPI dataset (assuming no additional recruitment efforts), 2-year trials appear to be feasible for *GRN* if MRI is used as the outcome, whereas a 4-year trial would be required for *MAPT*. Additional recruitment would be required for a *C9orf72* prevention trial to be sufficiently powered to detect a 50% treatment effect.

Symptomatic trial simulations included all participants with a CDR+NACC-FTLD-Global = 1 and subsets of high-risk participants with a CDR+NACC-FTLD-Global of 0 or 0.5 defined based on elevated NfL ($\log(\text{NfL}) > 3.0$) or an estimated DA within 2.5 years of onset (Table 3). Based on these simulations, it would be feasible to power trials for all three genetic groups using the CDR+NACC-FTLD-SB and neuropsychological tests, measures most likely to be approvable by regulatory bodies as clinically meaningful endpoints¹⁵. For example, within a population having a CDR+NACC-FTLD-Global = 1 or a DA within 2.5 years of onset in those with a CDR+NACC-FTLD-Global < 1 , the estimated sample sizes using CDR+NACC-FTLD-SB as the primary endpoint for a 2-year trial were 68 total participants for *GRN*, 120 for *MAPT* and 124 for *C9orf72*.

Discussion

We present the efforts of the international FPI to establish the largest known cohort of f-FTD cases worldwide, gathered from North American (ALLFTD) and European/Canadian (GENFI) natural history studies. We harmonized clinical, neuropsychological, biofluid, and neuroimaging measurements to build DPMs that allow direct comparisons of effect sizes for mean values and rates of change between the best available measures for characterizing FTD. The DPMs revealed important insights about the earliest manifestations of f-FTD and the temporal ordering of biomarker and clinical changes. Across all three FTD mutation carrier groups, regional brain atrophy and plasma NfL elevations were the first measurable manifestations of disease, potentially developing 10 to 40 years before the earliest clinical features. Neuropsychological changes typically occurred later, contemporaneous with the emergence of informant-reported symptoms (CDR+NACC-FTLD-SB).

Table 2 | Baseline descriptive statistics of measures for each genetic group at three epochs

Mutation status	Outcome measure	DA epoch			
		–40 to –10 YSO	–10 to 0 YSO	0+ YSO	
Age-matched controls	<i>n</i> (prop) at baseline	229 (0.56)	85 (0.21)	98 (0.24)	
	Mean age (s.d.) at baseline	36.8 (7.7)	52.6 (6.7)	61.6 (7.7)	
	Mean raw score (s.d.; range)	CDR+NACC-FTLD SB	0 (0; 0-0)	0 (0; 0-0)	0 (0; 0-0)
		Trails A (total time in seconds)	22.76 (8.03; 8-78)	26.36 (9.39; 12-61)	31.07 (14.67; 12-89)
		Trails B (total time in seconds)	53.81 (21.93; 19-187)	62.06 (29.48; 27-202)	73.63 (30.43; 31-167)
		MINT (total correct)	29.92 (1.75; 24-32)	29.94 (1.62; 26-32)	29.95 (1.92; 25-32)
		MRI frontal/TIV	7.07 (0.48; 5.39-8.21)	6.68 (0.41; 5.83-7.55)	6.33 (0.45; 5.27-7.28)
		MRI temporal/TIV	4.76 (0.29; 3.76-5.62)	4.54 (0.22; 4.07-5.03)	4.24 (0.28; 3.46-4.79)
		MRI MTL/TIV	1.03 (0.06; 0.81-1.22)	1.02 (0.06; 0.89-1.19)	0.97 (0.07; 0.8-1.13)
		NfL (log)	1.67 (0.43; 0.38-3.27)	2.05 (0.38; 1.06-2.94)	2.42 (0.43; 1.71-3.76)
C9orf72	<i>n</i> (prop) at baseline	135 (0.39)	63 (0.18)	149 (0.43)	
	Mean age (s.d.) at baseline	38.3 (8.8)	54.6 (8.2)	61.5 (9)	
	Mean raw score (s.d.; range)	CDR+NACC-FTLD SB	0.19 (0.57; 0-3)	0.31 (0.69; 0-3.5)	8.32 (6.23; 0-22)
		Trails B (total time in seconds)	58.92 (21.85; 28-151)	84 (45.61; 23-300)	168.25 (88.4; 35-300)
		MRI temporal/TIV	4.58 (0.29; 3.95-5.22)	4.16 (0.32; 3.43-4.71)	3.76 (0.46; 2.29-4.78)
		NfL (log)	1.89 (0.48; 0.94-3.89)	2.58 (0.6; 1.72-4.76)	3.31 (0.85; 1.54-5.54)
	Mean standardized units from control (s.d.; range)	CDR+NACC-FTLD SB	-	-	-
		Trails B	0.23 (1; -1.18-4.43)	0.74 (1.55; -1.32-8.07)	3.11 (2.91; -1.27-7.44)
		MRI temporal/TIV	-0.62 (1; -2.79-1.56)	-1.75 (1.43; -5.04-0.76)	-1.71 (1.65; -6.92-1.94)
		NfL (log)	0.51 (1.11; -1.68-5.1)	1.37 (1.57; -0.85-7.06)	2.07 (1.96; -2.01-7.18)
GRN	<i>n</i> (prop) at baseline	125 (0.44)	72 (0.26)	84 (0.3)	
	Mean age (s.d.) at baseline	41 (10.3)	58.2 (7.5)	63.7 (8.8)	
	Mean raw score (s.d.; range)	CDR+NACC-FTLD SB	0.08 (0.26; 0-2)	0.31 (0.71; 0-3)	9.19 (6.53; 0-24)
		Trails A (total time in seconds)	25.37 (9.2; 9-63)	30.57 (10.73; 16-81)	72.12 (46.48; 23-150)
		MRI frontal/TIV	7.03 (0.52; 5.39-8.93)	6.4 (0.52; 5.25-7.48)	5.15 (0.92; 2.62-7.77)
		NfL (log)	1.87 (0.43; 0.82-3.34)	2.45 (0.56; 1.57-4.27)	4.04 (0.65; 2.14-5.35)
	Mean standardized units from control (s.d.; range)	CDR+NACC-FTLD SB	-	-	-
		Trails A	0.33 (1.15; -1.71-5.01)	0.45 (1.14; -1.1-5.82)	2.8 (3.17; -0.55-8.11)
		MRI frontal/TIV	-0.08 (1.09; -3.49-3.89)	-0.68 (1.26; -3.46-1.92)	-2.59 (2.02; -8.18-3.18)
		NfL (log)	0.46 (1; -1.95-3.84)	1.04 (1.45; -1.25-5.79)	3.74 (1.49; -0.62-6.75)
MAPT	<i>n</i> (prop) at baseline	69 (0.41)	37 (0.22)	62 (0.37)	
	Mean age (s.d.) at baseline	34.1 (9.2)	46.3 (9.5)	56.1 (8.6)	
	Mean raw score (s.d.; range)	CDR+NACC-FTLD SB	0.15 (0.48; 0-2.5)	0.39 (0.76; 0-3)	7.9 (6.51; 0-24)
		MINT (total correct)	29.88 (1.8; 25-32)	29.16 (3; 17-32)	21.22 (8.04; 1-32)
		MRI MTL/TIV	1.05 (0.06; 0.87-1.16)	0.98 (0.07; 0.77-1.08)	0.72 (0.14; 0.46-1.04)
		NfL (log)	1.69 (0.45; 0.39-2.53)	1.98 (0.55; 0.93-3.44)	3.04 (0.55; 1.93-5.1)
	Mean standardized units from control (s.d.; range)	CDR+NACC-FTLD SB	-	-	-
		MINT	-0.02 (1.03; -2.82-1.19)	-0.48 (1.85; -7.98-1.27)	-4.56 (4.2; -15.12-1.07)
		MRI MTL/TIV	0.41 (1.04; -2.7-2.15)	-0.69 (1.29; -4.46-1.15)	-3.33 (1.86; -6.87-1.04)
		NfL (log)	0.04 (1.03; -2.95-1.98)	-0.19 (1.45; -2.91-3.63)	1.45 (1.26; -1.11-6.17)

Baseline raw and standardized values for several measures are displayed for controls and mutation carriers at three DA epochs. Each participant was assigned to a DA epoch based on the estimated DA at their first visit. Clinical and imaging measures were selected by choosing the 'best' measure for each genetic group based on when they became elevated compared to controls and the rate of longitudinal change (descriptive statistics for all modeled measures are displayed in Supplemental Tables 2 and 3). Raw imaging measures are presented as percentage of total intracranial volume to account for head size. Mean standardized units from controls indicates the number of standard deviations from the control group. Prop, proportion. MTL, medial temporal lobe

The genetic groups displayed differences in patterns of disease progression that are relevant for clinical care and clinical trial planning. The striking concordance in disease progression between the two independent North American and European cohorts supports the validity of the models, suggesting that the natural history of the disease is strongly determined by pathogenic mutations and that global

clinical trials of f-FTD therapies are feasible. Finally, we leveraged the DPMs and natural history data to simulate prevention and early symptomatic treatment trial scenarios, including candidate participant selection criteria and primary endpoints, to provide evidence for the feasibility of running presymptomatic prevention trials and symptomatic treatment trials in f-FTD.

Table 3 | Clinical trial sample size estimates**Presymptomatic prevention trial (CDR+NACC-FTLD Global = 0)**

Genetic group	Estimated number of eligible participants	Inclusion criteria	Primary endpoint: sample size estimates (50% treatment effect)							
			CDR+NACC-FTLD-SB		Neuropsychological tests		NfL (log)		MRI volume	
			2 yr	4 yr	2 yr	4 yr	2 yr	4 yr	2 yr	4 yr
<i>C9orf72</i> MRI = temporal NP = Trails B	171	All CDR 0	>10,000	4,994	>10,000	6,784	3,397	699	1,639	394
	13	CDR 0 and NfL (log) > 3	582	334	1,113	386	>10,000	638	537	173
	38	CDR 0 and DA > -5	508	224	657	184	527	153	424	119
	20	CDR 0 and DA > -2.5	266	111	364	96	439	123	402	102
<i>GRN</i> MRI = frontal NP = Trails A	168	All CDR 0	3,144	1,526	3,844	1,576	684	271	826	459
	7	CDR 0 and NfL (log) > 3	250	179	250	140	158	51	71	46
	26	CDR 0 and DA -5	297	182	267	130	99	30	52	27
	10	CDR 0 and DA -2.5	182	104	159	79	84	26	37	24
<i>MAPT</i> MRI = MTL NP = MINT	94	All CDR 0	7,073	2,733	>10,000	3,741	3,059	802	1,492	526
	4	CDR 0 and NfL (log) > 3	283	188	373	220	>10,000	501	147	72
	19	CDR 0 and DA -5	362	190	641	265	595	149	108	39
	14	CDR 0 and DA -2.5	191	97	311	134	438	117	72	24

Early symptomatic treatment trial (all CDR+NACC-FTLD Global = 1 enriched with 0 and 0.5 participants)

Genetic Group	Estimated number of eligible participants	Inclusion criteria	Primary endpoint: sample size estimates (50% treatment effect)							
			CDR+NACC-FTLD-SB		Neuropsychological tests		NfL (log)		MRI volume	
			1.5 yr	2 yr	1.5 yr	2 yr	1.5 yr	2 yr	1.5 yr	2 yr
<i>C9orf72</i> MRI =temporal NP = Trails B	94	ALL CDR 0.5 and 1	188	129	340	203	811	483	639	367
	37	All CDR 1 and (CDR 0 and 0.5 if NfL > 3)	161	115	370	222	1,806	782	645	358
	83	All CDR 1 and (CDR 0 and 0.5 if DA > -2.5)	176	124	400	207	740	423	678	360
	67	All CDR 1 and (CDR 0 and 0.5 if DA > 0)	117	79	275	161	628	384	669	359
<i>GRN</i> MRI = frontal NP =Trails A	67	ALL CDR 0.5 and 1	76	66	115	79	133	76	44	30
	33	All CDR 1 and (CDR 0 and 0.5 if NfL > 3)	97	84	124	92	182	110	49	36
	48	All CDR 1 and (CDR 0 and 0.5 if DA > -2.5)	79	68	105	74	127	75	36	26
	38	All CDR 1 and (CDR 0 and 0.5 if DA > 0)	39	32	62	41	124	72	32	22
<i>MAPT</i> MRI = MTL NP = MINT	43	ALL CDR 0.5 and 1	175	136	300	196	845	437	124	74
	11	All CDR 1 and (CDR 0 and 0.5 if NfL > 3)	89	66	138	91	1,719	769	95	59
	43	All CDR 1 and (CDR 0 and 0.5 if DA > -2.5)	164	120	244	163	779	419	109	63
	31	All CDR 1 and (CDR 0 and 0.5 if DA > 0)	96	66	150	104	627	359	83	48

In the top half of the table, sample size estimates (total n for a trial) are presented for presymptomatic prevention trials in which all enrolled participants are presymptomatic based on CDR+NACC-FTLD Global score = 0. Within each genetic group, sample sizes are estimated for trials enrolling all presymptomatic participants as well as three additional scenarios in which NfL or DA are used to enroll high-risk participants likely to be proximal to symptom onset. In the bottom half of the table, estimates are presented for an early symptomatic trial in which all participants with a CDR+NACC-FTLD Global = 1 are eligible, and those with CDR+NACC-FTLD Global < 1 are included based on different inclusion criteria. The estimated number of eligible participants refers to the number of participants in the current dataset that meet the specified inclusion criteria. For each genetic group, we select a representative MRI and neuropsychological measures (displayed in the first column). Bolded cells indicate that the sample size estimates are less than or within 15 participants of the number eligible. All trial designs assume 1:1 randomization treatment versus control, 10% attrition per year, and have a primary analysis of a change from baseline in the primary endpoint. Additional details of the assumptions underlying these simulations can be found in Supplementary Table 9. NP, group-specific neuropsychological measure.

The validity of our DPM models is supported by the results of previous studies focusing on individual biomarkers or clinical measures in f-FTD. Because the models incorporate both new and some previously analyzed historical data, we were able to replicate and extend the results of previous studies. We also directly compared the relative utility of different assessments at different stages of disease. Consistent with previous MRI studies demonstrating brain atrophy can be detected in presymptomatic f-FTD^{16–19}, MRI-measured brain atrophy was the first biomarker to change in *C9orf72* and *MAPT*, but our models revealed that NfL elevations preceded atrophy by a few years in *GRN*. In *C9orf72*, the thalamus and most other brain regions were smaller than controls 10 to 40 years before onset, supporting the hypothesis that *C9orf72* repeat expansions may affect early brain development^{19,20}. Also consistent with prior work, the

most rapid rates of atrophy occurred in *GRN* with widespread brain involvement within 10 years of onset^{21,22}. Despite differences in analytic methods, and the inclusion of a much larger dataset, the DPMs developed in this study allowed us to replicate the findings of earlier, smaller analyses. In an earlier MRI study, Rohrer and colleagues¹⁷ defined expected disease onset based on each genetic group's mean age of onset rather than using model derived DA used here. Similar to the previous study, we detected medial temporal atrophy in *MAPT* 15 years before onset followed by atrophy of the insula. Temporal lobe atrophy in presymptomatic *MAPT* has been consistently reported^{16,18,23}, and the insula may be a common region of early atrophy in *MAPT*²⁴.

We and others have previously shown that NfL concentrations are elevated in the plasma^{25–27} and cerebrospinal fluid^{28,29} of

symptomatic FTD patients compared to other neurological conditions. In the current study, we verified that the genotype-related patterns of plasma NfL elevation that were measured in two different laboratories, in two independent f-FTD cohorts, were very similar and for the purposes of DPM, could be combined. In *C9orf72*, NfL levels began to deviate from controls approximately 30 years before onset and remained significantly elevated compared to controls in all presymptomatic epochs. In *GRN*, NfL levels begin to increase 15 years before onset and were elevated compared to controls in the late presymptomatic stages. In contrast, NfL levels begin to increase just proximal to symptom onset in *MAPT*, and presymptomatic *MAPT* mutation carriers did not show increased levels compared to controls. In the symptomatic stage, NfL levels rose more than twice as fast in *GRN* than the other genetic groups. These results extend previous fluid biomarker studies showing NfL concentrations become elevated early in f-FTD, are harbingers of symptom onset and rise most rapidly in *GRN*^{25,27,30–32}.

Paralleling the biomarker findings, global disease severity (CDR+NACC-FTLD-SB) and neuropsychological measures declined more rapidly in *GRN* than *C9orf72* or *MAPT* mutation carriers. Although *GRN* was previously shown to have the longest disease course in an international f-FTD cohort¹², disease duration in that study was determined based on clinical interview rather than the data-driven approach taken in the current study; moreover, the *C9orf72* sample in the prior study had a higher proportion of participants with ALS or FTD with motor neuron disease than the current study (30.3% versus 13.1%), and these diagnoses were associated with more rapid disease progression^{12,33}. Neuropsychological impairments relative to age-matched controls were typically observed after symptom onset in all groups, although abnormalities on a few measures were detected in the presymptomatic stages. These findings add to prior studies suggesting that cognitive changes can be detected in the presymptomatic phases of f-FTD and that there are genotype-specific cognitive profiles^{34–37}. Future work should continue to explore the development and validation of novel neuropsychological measures for early detection and monitoring, including digital cognitive tests and cognitive composite scores (for example, GENFI-COG) that may improve early detection and reduce sample size estimates³⁷.

An overarching aim of this study was to develop models that inform the design of f-FTD clinical trials. Simulation studies were conducted to estimate the sample sizes necessary to power prevention and early symptomatic treatment trials. These studies also explored the use of NfL and DA estimates as inclusion criteria to enroll presymptomatic mutation carriers at heightened risk for clinical progression during a trial. The simulations revealed important information that will be directly applicable to clinical trial design. First, using NfL and MRI biomarkers as surrogate endpoints for prevention trials would allow trials to be conducted with many fewer participants than clinical measures. Second, prevention trials appear most feasible for *MAPT* and *GRN* relative to the estimated number of eligible participants based on our dataset, however, given that *C9orf72* is the most common mutation causing FTD and ALS, recruiting the estimated sample sizes may be feasible. Third, using estimated DA to select high-risk presymptomatic participants for trial enrollment leads to a sizeable reduction in sample sizes. This reduction in sample size must be balanced against the reduction in number of eligible participants (of that DA), but these simulations show that *GRN* and *MAPT* trials enrolling presymptomatic participants within 5 years of estimated onset would be feasible based on the estimated number of eligible participants from our current dataset. Fourth, clinical measures perform very well in the early symptomatic trial simulations, and sample sizes for trials using the CDR+NACC-FTLD-SB as a primary outcome are feasible for all three genetic groups. Not only was this measure statistically powerful for measuring change, but given that it reflects informant-reported clinical status, it could also be considered a

clinically meaningful outcome and approvable endpoint from a regulatory perspective¹⁵.

The clinical trial simulations included in this study used standard, two-arm, parallel-group clinical trial designs. Future work to explore innovative trial designs and analysis methods may enable trials with smaller samples sizes and/or increased power for smaller (but clinically meaningful) treatment effects. With the incorporation of a treatment effect parameter, the DPM-predicted versus posttreatment progression could potentially be used as a primary endpoint in clinical trials to estimate the slowing in disease progression across multiple endpoints^{7,38}. In rare diseases such as f-FTD, analytic methods may also simulate data from natural history participants to generate ‘synthetic’ participants to decrease sample sizes and reduce allocation to placebo as has been encouraged in recent US Food and Drug Administration guidance¹⁴. Additionally, platform trials based on DPMs allow multiple therapies to be tested simultaneously with comparisons made to a shared placebo group further improving trial efficiency in rare populations³⁹.

There are important limitations to this work. Known genetic modifiers of f-FTD disease progression were not included, such as specific mutations (for *MAPT*) and *TMEM106B*, a modifier of penetrance in *GRN*^{7,40}. We were also limited in the clinical measures that we could include in the analysis to those that were readily harmonizable between ALLFTD and GENFI, excluding a variety of promising novel measures that were not available in both cohorts^{34,35}. Future models will likely be improved by including a more exhaustive collection of measures and biomarkers⁴¹ and approaches accounting for heterogeneity in f-FTD features⁴². Because disease onset was defined as CDR+NACC-FTLD-SB=0.5, noncarrier controls by definition had CDR+NACC-FTLD-SB = 0 at baseline, which reduced the variance in this measure, thereby potentially overestimating the effect size relative to other measures where there was more variance in the controls. Because abnormal global status may reflect other brain pathologies in the controls that could potentially obscure important findings, we believe that the requirement for CDR+NACC-FTLD-SB=0 in controls was appropriate.

The DPMs produced for the current study have additional limitations related to less informative clinical data at early stages of disease and missing data at late stages of disease. In subjects estimated to be within 10 years of symptom onset, the accuracy is ± 5.5 years, which approaches the accuracy of familial age of onset-based estimates which are useful in DIAD⁴³, but not possible in most f-FTD syndromes¹². However, individuals furthest from onset are typically within normal limits on all contributing measures forcing the model to rely heavily on prior information about participants’ chronological age to estimate DA. This results in considerable uncertainty around exact DA in those furthest from expected onset (for example, ± 14 years in the -40 to -10 epoch). To visually assess how the weight of evidence (number of measures that changed over the range of visits) related to each subject’s DA, we color coded measurements in each individual mutation carrier in Supplementary Fig. 6. This revealed that in more severely impaired mutation carriers at later DA, there was more missing data, particularly MRI. This suggests an important limitation to the use of MRI as an outcome in symptomatic mutation carriers: data may be missing because scans are harder to acquire in advanced patients, possibly because they either cannot travel to research centers or they cannot lie still in a MRI scanner. Such informative missing data also impact the DPMs, potentially biasing the models toward a smaller standard deviation from normal; this is a limitation and a direction for future research. Finally, the current study is limited by the lack of racial and ethnic diversity of the sample. Improving the diversity of participants in FTD research is an urgent priority⁴⁴, however, it should be noted that in genetic f-FTD there are known founder effects for *C9orf72*⁴⁵ and *GRN* mutations⁴⁶ with European ancestry, leading to strong associations with particular racial and ethnic groups.

In conclusion, these DPMs will facilitate the planning of f-FTD clinical trials, including selection of optimal endpoints and enrollment criteria to maximize power to detect treatment effects¹⁴. Brain atrophy and plasma NfL elevations are measurable years before symptom onset, paving the way for using these biomarkers in clinical trials of agents that could prevent or delay the clinical manifestations of f-FTD. The models also highlight the challenges of conducting adequately powered trials in rare f-FTD populations and provide a roadmap for development of new biomarkers and clinical endpoints that may improve power to detect effects in presymptomatic stages of disease and create a renewed sense of urgency to identify eligible trial participants outside of Europe and North America.

Online content

Any methods, additional references, Nature Research reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41591-022-01942-9>.

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Methods

Participants. Participants included 796 carriers of pathogenic mutations in the *C9orf72*, *GRN* or *MAPT* genes and 412 noncarrier controls from families with a known mutation in one of these genes. Participants were enrolled through Advancing Research and Treatment for Frontotemporal Lobar Degeneration (ARTFL; NCT02365922) and Longitudinal Evaluation of Familial Frontotemporal Dementia Subjects (LEFFTDS; NCT02372773)⁴⁷, which recently combined into the ARTFL/LEFFTDS Longitudinal Frontotemporal Lobar Degeneration (ALLFTD; NCT04363684) study. These studies enrolled participants through a consortium of 18 centers across the US and Canada between 2015 and 2020. Participants were also enrolled through the Genetic Frontotemporal Initiative (GENFI), which involves 25 research centers across Europe and Canada. GENFI 2 participants from the 5th Data Freeze (2015–2019) were included. All participants were required to have completed the Clinical Dementia Rating Scale (CDR) plus Behavioral and Language Domains from the National Alzheimer's Coordinating Center (NACC) FTLD module (CDR+NACC-FTLD). GENFI 1 (2012–2015) participants were excluded because the CDR+NACC-FTLD was not collected during that study period. Some participants in GENFI 2 and ALLFTD cohorts underwent longitudinal evaluations, and all available data were included. ALLFTD participants received travel compensation and remuneration up to \$100 based on the study they participated in. For GENFI, Travel, accommodations, or other reasonable expenses are offered to the participants to cover any costs they incur in order to attend the research visits. The ALLFTD study was approved through the Trial Innovation Network at Johns Hopkins University. Local ethics committees at each of the sites approved the study, and all participants provided written informed consent or assent with proxy consent.

ALLFTD inclusion/exclusion criteria relevant to this study. Inclusion criteria. Participants must be a member of a family with a known pathogenic mutation in the *GRN* or *MAPT* genes or with a pathogenic expansion in the *C9orf72* gene. The participant does not have to know their own genetic status but must be at least 18 years of age. The predominant phenotype in most families is cognitive or behavioral. However, families may present with motor-dominant syndromes without exclusion. Participants must have a reliable informant who personally speaks with or sees that subject at least weekly. Participants are sufficiently fluent in English to complete all measures. Participants must be willing and able to consent to the protocol and undergo yearly evaluations over 3 years. Participants must be willing and able to undergo neuropsychological testing (at least at baseline visit) and have no contraindication to MRI imaging. Noncarrier family controls were included in the current study if they were clinically normal at baseline, defined by a CDR+NACC-FTLD Global = 0.

Exclusion criteria. Known presence of a structural brain lesion (for example, tumor, cortical infarct). Presence of another neurologic disorder that could impact findings (for example, multiple sclerosis).

GENFI inclusion/exclusion criteria relevant to this study. Inclusion criteria. Participants are at least 18 years old. Participants must be a member of a family with a known pathogenic mutation in the *GRN* or *MAPT* genes or with a pathogenic expansion in the *C9orf72* gene. If the participant is cognitively impaired, there must be an available caregiver that can escort them. The participant must have an identified informant. The participant must be fluent in the language of their country of assessment. Noncarrier family controls were included in the current study if they were clinically normal at baseline, defined by a CDR+NACC-FTLD Global score = 0.

Exclusion criteria. Participant has another medical or psychiatric illness that would interfere in completing assessments. Participant is pregnant. Local MRI and lumbar puncture contraindications. The predominant phenotype in most families is cognitive or behavioral. However, families may present with motor-dominant syndromes without exclusion.

Genetic testing. ALLFTD participants had genetic testing at the University of California, Los Angeles using published methods⁴⁸. GENFI participants were genotyped at their local sites according to previous methods⁴⁷. Briefly, in ALLFTD and GENFI, DNA samples were screened using targeted sequencing of a custom panel of genes previously implicated in neurodegenerative diseases, including *GRN* and *MAPT*. The presence of hexanucleotide repeat expansions in *C9orf72* was detected in ALLFTD using both fluorescent and repeat-primed PCR and in GENFI using repeat-primed PCR.

Clinical assessment. The ALLFTD and GENFI multidisciplinary assessments includes neurological history and examination and collateral interview⁴⁷. Documented years since onset, which was entered as prior in the model, was based on clinical interview.

The CDR+NACC-FTLD module is an eight-domain rating scale based on informant report^{49–51}. A Global score was calculated to categorize disease severity as presymptomatic (0), questionable or mild symptoms of neurodegenerative disease

(0.5), or clear symptoms of dementia (1, 2, or 3)⁴⁹. A sum of the eight box scores (CDR+NACC-FTLD-SB) was also calculated; this score ranges from 0 to 24, with higher scores indicating greater functional impairment⁴⁹.

A subset of neuropsychological tests from the Uniform Data Set (UDS) Neuropsychological Battery, version 3.0⁵² was available for both consortia: Trail Making Test Parts A and B, the Multilingual Naming Test (Boston Naming Test in GENFI), Number Span Forward and Backward (Digit Span in GENFI), Benson Figure Copy and Delayed Recall and Animal Fluency. Conversion tables from the UDS Crosswalk study were used to harmonize Number Span/Digit Span and the MINT/BNT⁵³. Upon review of neuropsychological test scores in the controls, one outlier score was removed. As a sensitivity analysis to consider the impact of additional demographic covariates (that is, sex, education and language), statistical harmonization of the neuropsychological data was conducted using a *W*-score approach^{42,54}, which is a standardized score controlled for nuisance covariates. Regression models were built using baseline neuropsychological test scores in the noncarrier controls, with separate models in each consortium. All regressions included sex and education. In the GENFI cohort, primary language was included as an additional categorical covariate. Next, in all participants at every time point, the difference between their actual score and predicted score (based on regression conducted in controls) was divided by the standard deviation of the control group to derive a standardized estimate compared to controls with the same demographic background.

Neuroimaging. Image acquisition. Details of image acquisition, processing, and harmonization can be found below and have been published elsewhere⁵⁵. ALLFTD participants were scanned at 3T on MRI scanners (scanner types are displayed in Supplementary Table 7). T1-weighted images from ALLFTD were acquired as Magnetization Prepared Rapid Gradient Echo images using the following parameters: 240 × 256 × 256 matrix; about 170 slices; voxel size = 1.05 × 1.05 × 1.25 mm³; flip angle, echo time and repetition time varied by vendor. A standard imaging protocol was used across all centers, managed, and reviewed for quality by a core group at the Mayo Clinic, Rochester.

GENFI participants underwent volumetric T1-weighted MRI using the standard GENFI protocol^{17,56}. A variety of 1.5T and 3T scanners were used across the sites: Siemens Trio, Siemens Skyra, Siemens Prisma, Philips and General Electric. The scan protocols were designed at the start of the GENFI study to ensure that there was adequate matching between the scanners and the quality of the images. T1-weighted images from GENFI were acquired using the following parameters: 256 × 256 × 208 matrix; 208 slices; voxel size = 1.1 mm isotropic, flip angle = 8°, echo time and repetition time varied by vendor. All scans were quality checked and those with movements or artifacts were removed. Furthermore, if any participants displayed moderate to severe vascular disease or any other brain lesions, they were also excluded from the analysis.

Image processing. The same image processing steps were performed on ALLFTD and GENFI data. Before any preprocessing of the images, all T1-weighted images were visually inspected for quality control. Images with excessive motion or image artifact were excluded. T1-weighted images underwent bias field correction using N3 algorithm⁵⁷. The segmentation was performed using SPM12 (Wellcome Trust Center for Neuroimaging, London, UK, <http://www.fil.ion.ucl.ac.uk/spm>) unified segmentation⁵⁸. A customized group template was generated from the segmented gray and white matter tissues and cerebrospinal fluid by non-linear registration template generation using the Large Deformation Diffeomorphic Metric Mapping framework⁵⁹. Subjects' native space gray and white matter were geometrically normalized to the group template, modulated, and then smoothed in the group template. The applied smoothing used a Gaussian kernel with 8-mm full width half maximum. Every step of the transformation was carefully inspected from the native space to the group template.

Regional volume estimates were calculated from individual subjects' smoothed, modulated gray matter in template space, by taking the mean of all voxels in several a priori ROIs⁶⁰ by taking the mean of all voxels within the following regions: Frontal, Temporal, Medial Temporal (consisting of amygdala, hippocampus, entorhinal cortex, and parahippocampal gyrus ROIs), Parietal, and Occipital Lobes, Striatum, Insula, Thalamus, and Cerebellum. Volume estimates were then represented as percentage of total intracranial volume. To understand the effects of scanner and to present voxelwise maps, a *W*-score was created at each voxel to represent volume relative to controls after adjusting for covariates. First, a multivariable linear model was fit for each voxel in a reference group that consisted of the first available scan for noncarrier family controls. Predictors in this model were total intracranial volume (TIV) and scanner platform^{42,54}. Next, for each voxel of every available MRI in the study, the same model was fit, entering TIV and scanner, using the coefficients from the reference group to extract a residual. This residual was then divided by the standard deviation of the residuals in the reference group. Therefore, the *W*-score represents the gray matter content at that voxel as the number of standard deviations away from the expected mean for a reference group, accounting for TIV and scanner platform. We then created a mean *W*-score value for each ROI and entered it into the model as a sensitivity analysis. Mean *W*-scores at each voxel in mutation carriers are also presented in supplemental figures.

Plasma NfL. ALLFTD methods. Plasma NfL light concentrations were measured at the Mayo Clinic in Jacksonville using the Quanterix single-molecule array technology (Simoa) @ NF-Light Advantage Kit (103186, lot 501992) and the HD-X instrument according to the instructions provided. Samples were tested in duplicate using kits from the same lot. In addition to the two quality control samples provided with the kit, all assays included five interassay controls. Before each assay, plasma samples were thawed, mixed thoroughly by low-speed vortexing, centrifuged at 10,000 g for 5 min, and transferred to 96-well plates that were then sealed to minimize sample evaporation. Samples were diluted four times by the instrument. If levels of NfL in a sample exceeded the upper limit of the calibration curve, the sample was retested at a higher dilution. Across all assays, the percent coefficient of variations of the mean NfL concentration for the interassay controls were below 10%.

GENFI methods. Plasma NfL concentrations were measured at baseline with Simoa, using the commercially available Simoa Neurology 4-Plex A kit (Quanterix, 102153, Lot#: 50216). Plasma samples were thawed at room temperature (one cycle), mixed thoroughly and centrifuged at 14,000 g for 3 min. The supernatant was loaded onto a Quanterix HD-1 Analyzer with a 1:4 specified dilution. Measures were completed in duplicate over a total of six batches, each with an eight-point calibration curve tested in triplicate and two controls tested in duplicate. Plasma concentrations were interpolated from the calibration curve within the same batch and corrected for the dilution. All samples were quantifiable within the dynamic range of 0.69 to 2,000 pg ml⁻¹ and with an average coefficient of variation below 10%. Instrument operators were blinded to clinical and genetic information.

Prior publications. Prior publications have included some of the data included in these models, including publications describing MRI^{16,18,23,42,55,61–63}, NfL^{25,27,30} and clinical data^{12,31,35,37,41,51,55}. For full lists of publications from these consortia see <https://www.allftd.org/publications> and <https://www.genfi.org/publications/>. This study is the first comprehensive effort to combine clinical, imaging, and plasma biomarker data across consortia.

Statistical analyses. All available data were included in the statistical analyses. Complete cases were not required, and no imputation was conducted. Statistical tests were two sided.

Participant characteristics. Demographic variables and other participant characteristics (Table 1) were compared across genetic groups and controls using regression with pairwise group contrasts for most variables. Sex, race, CDR+NACC-FTLD and diagnostic categories were compared using chi-squared with Bonferroni-adjusted pairwise comparisons when the omnibus test was significant. For chi-squared tests in which the sample size of any bin was <10, the Fisher's exact test was used.

DPMs. DPMs were built using a Bayesian mixed effects framework, with the goal of estimating a single latent disease stage parameter for each person, which we refer to as DA. The DPM is a joint model of all 20 measures listed in Supplementary Table 8. DA is the estimated difference between an individual's chronological age and the age of symptom onset (defined for this study as a CDR+NACC-FTLD-SB = 0.5). This estimate is positive for symptomatic cases and negative for presymptomatic cases. The model included priors based on an individual's time from expected symptom onset. In symptomatic cases, we used the clinician's estimate of time from symptom onset, sampled from a normal distribution with a small amount of error (s.d. = 4) to acknowledge the imperfection of this estimate. For presymptomatic cases and noncarrier controls, we used the mean age of the mutation group as a prior, sampled from a normal distribution with more noise (s.d. = 10). The prior standard deviations of 4 and 10 were chosen to be relatively noninformative. For a subject with an observed clinician's estimate of time since symptom onset, there is a 95% prior probability that the true age of onset was within ±8 years of the clinician's estimate. For a subject whose onset has not yet been observed, there is a 95% prior probability that the true age of onset was within ±20 years of the mean estimated age of onset from the same mutation group.

DA was then estimated from a joint analysis of all available clinical, neuropsychological, imaging, and NfL data. Simultaneously, overall disease progression of each endpoint was modeled as a function of latent DA with several parameters, including expected value at 'normal', total decline, endpoint and mutation-specific rate and timing of progression. To account for variability in values of each endpoint at healthy across subjects, we included subject-specific random effects that were correlated across similar endpoints (see Grouping variable in Supplementary Table 8).

First, models were built separately in each cohort. Visual inspection suggested sufficient alignment between disease progression of all endpoints across the two consortia and subsequent models combined both cohorts within a single analysis. A detailed description of the approach follows:

Latent disease stage DPM:

- Model each endpoint, $k = 1:K$ for each subject, $i = 1:N$, for each visit, $j = 1:J_i$, as a function of latent disease stage, where $Y_{i,j,k}$ is the value of the endpoint k for subject i at visit j , and $X_{i,j}$ is the age for subject i at visit j .

- DA was defined as years since onset: age at visit minus age at onset, $D_{ij} = X_{i,j} - \alpha_i$. Age at onset is a latent variable that is estimated for each subject.
- The observed value $Y_{i,j,k}$ was assumed to be distributed normally with a subject and endpoint-specific mean and endpoint-specific variance that is a function of the mean.

$$Y_{i,j,k} \sim N(\mu_{i,j,k}, \sigma_k^2)$$

$$\mu_{i,j,k} = f_{i,k}(D_{ij})$$

- The subject and endpoint-specific mean decay function, $f_{i,k}(x)$, followed an exponential decay as a function of DA with location and scale parameters that are mutation specific. Mutations are denoted $m = 1:4$ for *C9orf72*, *GRN*, *MAPT*, and noncarriers respectively, m_i is an indicator of the mutation ($m = 1:4$) for subject i .

$$f_{i,k}(D_{ij}) = (\delta_{0,k} + \delta_{0,k,i}) + \frac{\delta_{1,k} - \delta_{0,k}}{1 + \exp(\theta_{k,m_i} + \beta_{k,m_i} * D_{ij})}$$

Model components and prior distributions:

- $\delta_{0,k}$: Value of the endpoint at normal/healthy state. Normal prior distribution with mean fixed based on expected value of endpoint at a normal state (see Supplementary Table 8) and s.d. = 10.
- $\delta_{1,k}$: Worst value for the endpoint (floor). Normal prior distribution with mean fixed based on expected worst value of the endpoint (Supplementary Table 8) and s.d. = 10.
- $\delta_{0,k,i}$: Subject and endpoint-specific random effects in value of the endpoint at normal state that are correlated across similar endpoints (Supplementary Table 8 shows groupings). Random effects are standardized based on the estimated endpoint-specific variability across subjects at normal, $\sigma_{\delta_{0,k}}^2$, and have a hierarchical prior distribution with subject-specific standardized mean for each group, g , of endpoints, $\mu_{\delta_{0,k},i}$, and group-specific variability across endpoints within a subject, $\sigma_{\mu_{\delta_{0,k},i}}^2$.

$$\frac{\delta_{0,k,i}}{\sigma_{\delta_{0,k}}} \sim N\left(\mu_{\delta_{0,k},i}, \sigma_{\mu_{\delta_{0,k},i}}^2\right); i = 1 : N; k = 1 : K$$

$$\mu_{\delta_{0,k},i} \sim N(0, 1); g = 1 : G;$$

$$1/\sigma_{\delta_{0,k}}^2 \sim \text{Gamma}(0.1, 0.1); k = 1 : K;$$

$$1/\sigma_{\mu_{\delta_{0,k},i}}^2 \sim \text{Gamma}(0.1, 0.1); g = 1 : G$$

Hyper-prior distributions for the endpoint-specific variability across subjects at normal and the group-specific variability across endpoints in that group within a subject have a mean value of 1 on the precision and s.d. = 10.

- $\theta_{k,m}$: Endpoint- and mutation-specific overall location of mean decay function. Location parameter was set for endpoint, $k = 1$, that corresponds to CDR+NACC-FTLD-SB so that the model is anchored to assume that a DA of 0 corresponds to a value on CDR+NACC-FTLD-SB of 0.5. For all other endpoints, we placed a noninformative prior distribution on the location parameter.

$$\theta_{k,m} \sim N(0, 10^2); k = 2 : K; m = 1 : 4.$$

In particular, $1/(1 + \exp(\theta_{k,m}))$, provides the percentage of the total decline of the endpoint at 'onset' (DA = 0). A value of 1 implies the endpoint is fully declined, a value of 0.5 implies 50% of the total decline. Under the above noninformative prior, 95% of the distribution of $1/(1 + \exp(\theta_{k,m}))$ is between 0 (<0.00001) and 1 (>0.99999) with a median value of 0.50.

- $\beta_{k,m}$: Endpoint and mutation-specific overall slope of mean decay function. For all endpoints and mutations, we placed a noninformative prior distribution on the scale parameter.

$$\beta_{k,m} \sim N(0, 10^2); k = 1 : K; m = 1 : 4.$$

- α : Age at onset per subject.

If value was observed within the dataset, we assumed that the prior distribution of a subjects age of onset was normal with a mean of the observed value and an s.d. = 4.

If value was not observed within the dataset, we assumed that the prior distribution of a subjects age of onset was normal with a mean of the imputed value (imputed as the mutation and study-specific mean from all observed ages of onset) and an s.d. = 10.

$$\alpha_i \sim N(\mu_{\alpha,i}, \sigma_{\alpha,i}^2);$$

$$Z_{i,j,k} \sim N(\hat{\mu}_{i,j,k}, \hat{\sigma}_k^2);$$

$\mu_{\alpha,i}$: Imputed or observed age of onset per subject.
 $\sigma_{\alpha,i}$: 4 if observed, 10 if imputed.
 σ_k^2 : Endpoint-specific measurement error.

$$1/\sigma_k^2 \sim \text{Gamma}(0.1, 0.1); k = 1 : K.$$

Computation. The Bayesian model was computed in R version 4.1.2, using the rjags package. This package uses Markov chain Monte Carlo to generate a sequence of dependent samples from the posterior distribution of the parameters. The Markov chain Monte Carlo had a burnin of 10,000 samples, followed by 100,000 samples.

Secondary analyses using estimated DA. After building the DPMs, we extracted estimates of DA for each observation. We then further explored the data in two different ways. For each endpoint, we first plotted raw values for mutation carriers and noncarriers as a function of DA. For each measure, we provide mutation-specific estimates for the age at which that measure deviates from controls by 1 s.d. Second, we binned mutation carriers and controls based on their DA at baseline (that is, epoch 1: DA = -40 to -10; epoch 2: DA = -10 to 0; epoch 3: >0). Epochs were chosen for illustrative purposes and to allow for a frequentist statistical analysis. For the cross-sectional data, we first compared the three genetic groups within an epoch by fitting a linear regression with the clinical measure or biomarker as the outcome, and genetic group as a three-level categorical variable. Multiple comparisons were controlled for using the Tukey method. Within each epoch, we also compared carriers to controls. Using the first available MRI scan for each participant, voxelwise mean W-scores for each bin were displayed for illustrative purposes. We also provide estimates of rates of change within each epoch based on the Bayesian DPM. Each DA estimate is associated with a 95% credible interval. The mean of these credible intervals is presented for each epoch to provide an estimate of how the model accuracy varies as a function of DA; we hypothesized greater uncertainty further away from onset, as most measures will be in the normal range at this stage, and thus, the model is more reliant on prior knowledge (that is, baseline age for presymptomatic cases).

Clinical trial simulation. Virtual clinical trial simulations are used to understand operating characteristics of proposed clinical trial designs. We simulated virtual patient outcomes under different assumptions for key design parameters to create simulated example trials. Within clinical trial simulation, generally, thousands of example trials are simulated under different sets of assumptions (scenarios) including trial sample size, randomization ratio, length of follow-up, targeted population, control progression rates and variability, and treatment effects. Overall average operating characteristics may then be summarized to quantify important characteristics of the proposed design (for example, type I error, power, treatment effect estimates).

Clinical trial simulation requires assumptions to be made about the underlying data. Results from the DPM can be used to create evidence-based assumptions about rates of progression and variability of progression of each endpoint for a target population.

To create a single simulated clinical trial dataset of participant-level endpoint values over time we used the following approach for subject i at visits $j = 1:N_j$ for endpoints $k = 1:K$:

- Simulate CDR+NACC-FTLD Global score at baseline given the mutation of the subject and distribution specified in Supplementary Table 9 (informed by natural history data).
- Simulate the DA at baseline given the CDR+NACC-FTLD Global score and the mutation type from the distribution specified in Supplementary Table 9 (informed by natural history data).
- Simulate a subject-level random effect at normal for each endpoint k by first simulating the overall subject-level standard units from normal for each group of endpoints, $g = 1:G$

$$\mu_{\delta_{0,g,i}}^* \sim N(0, 1); g = 1 : G$$

and then simulating the subject and endpoint-specific effect using sampled subject-level standard units from above for each group, g , and posterior estimates from the DPM

$$\delta_{0,k,i}^* \sim N\left(\mu_{\delta_{0,g,i}}^* * \hat{\sigma}_{\delta_{0,k}} + \hat{\sigma}_{\delta_{0,k}}^2 * \hat{\sigma}_{\mu_{\delta_{0,g}(k)}}^2\right).$$

- Simulate observed value of endpoint k , at visit j , $Z_{i,j,k}$, from a normal distribution with a subject and endpoint-specific mean and endpoint-specific variance based on the posterior mean results DPM, the subject-level DA at each visit, $DA_{i,j}$, and the subject-level random effect at normal, $\delta_{0,k,i}^*$ simulated above:

$$\hat{\mu}_{i,j,k} = (\hat{\delta}_{0,k} + \delta_{0,k,i}^*) + \frac{\hat{\delta}_{1,k} - \hat{\delta}_{0,k}}{1 + \exp(\hat{\theta}_{k,m_1} + \hat{\beta}_{k,m_1} * DA_{i,j})}.$$

- Subject may additionally be accepted / rejected on enrollment into the simulated clinical trial based on inclusion/exclusion criteria for CDR+NACC-FTLD Global score, DA at baseline, and/or NFL at baseline.

The expected change from baseline (mean and s.d.) over different timepoints for each endpoint for a placebo participant given a set of enrollment criteria are calculated using the above simulation strategy across 10,000 simulated datasets. The expected mean and s.d. of the change from baseline for a placebo participant is then used to calculate the sample size needed (N) to achieve 80% power for a 50% slowing in progression assuming 10% attrition rate per year and 1:1 randomization.

Enrollment criteria was defined based on baseline values of CDR+NACC-FTLD Global, log(NfL), and estimated DA. Presymptomatic trial designs consider only participants with a baseline CDR+NACC-FTLD Global = 0 and explored inclusion criteria to define a subpopulation at heightened risk for symptom onset based on elevated NfL ($\log(\text{NfL}) > 3.0$) or an estimated DA within 5 years or 2.5 years of onset. The hypothesis was that enrolling those presymptomatic cases close to onset would reduce the sample size needed to detect an effect by increasing the likelihood that the participants change on the endpoints during the trial period. Early symptomatic trial designs (CDR+NACC-FTLD = 0, 0.5 and 1) included all participants with a baseline Global score = 1. These simulations explored additional inclusion criteria for presymptomatic participants (Global score of 0 or 0.5) to define a high-risk subpopulation based on NfL or an estimated DA cutoff (-2.5 or 0).

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability

The datasets analyzed for the current study reflect collaborative efforts of two research consortia: ALLFTD and GENFI. Each consortium provides clinical data access based on established policies for data use: processes for request are available for review at allftd.org/data for ALLFTD data and by emailing genfi@ucl.ac.uk. Certain data elements from both consortia (for example raw MRI images) may be restricted due to the potential for identifiability in the context of the sensitive nature of the genetic data. The deidentified combined dataset will be available for request through the FTD Prevention Initiative in 2023 (<https://www.thefpi.org/>).

Code availability

Custom R code is available at <https://doi.org/10.5281/zenodo.6687486>.

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Author contributions

A.M.S., M.Q. and B. Wendelberger had full access to the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. A.M.S., M.Q., B. Wendelberger, H.W.H., L.R., H.J.R., J.D.R. and A.L.B. were responsible for concept development and design. A.M.S., M.Q. and B. Wendelberger conducted statistical analyses. M.Q. and B. Wendelberger developed the custom code for the DPMS. L.P., T.F.G. and C.H. processed the NfL data. Y.C., A. Wolf and S.Y.M.G. processed the neuroimaging data. A.M.S., M.Q. and B. Wendelberger drafted the manuscript. A.M.S., M.Q., B. Wendelberger, H.W.H., L.R., H.J.R., J.D.R. and A.L.B. critically revised the manuscript. A.L.B. supervised the research. B.F.B., H.J.R., J.D.R. and A.L.B. obtained funding. All authors contributed to acquisition, analysis or interpretation of data or revision of the manuscript.

Competing interests

B.A. receives research support from the Centers for Disease Control and Prevention, the National Institutes of Health (NIH), Ionis, Alector and the CJD Foundation. He has provided consultation to Acadia, Ionis and Sangamo. B.F.B. has served as an investigator for clinical trials sponsored by EIP Pharma, Alector and Biogen. He receives royalties from the publication of a book entitled *Behavioral Neurology of Dementia* (Cambridge Medicine, 2009, 2017). He serves on the Scientific Advisory Board of the Tau Consortium. He receives research support from the NIH, the Mayo Clinic Dorothy and Harry T. Mangurian Jr. Lewy Body Dementia Program and the Little Family Foundation. H.B. receives research support from the NIH. A.L.B. receives research support from the NIH, the Tau Research Consortium, the Association for Frontotemporal Degeneration, Bluefield Project to Cure Frontotemporal Dementia, Corticobasal Degeneration Solutions, the Alzheimer's Drug Discovery Foundation and the Alzheimer's Association. He has served as a consultant for Acovian, AGTC, Alector, Arkuda, Arvinas, Boehringer Ingelheim, Denali, GSK, Life Edit, Humana, Oligomerix, Oscotec, Roche, TrueBinding and Wave and received research support from Biogen, Eisai and Regeneron. B.C.D. is a consultant for Acadia, Alector, Arkuda, Biogen, Denali, Eisai, Genentech, Lilly, Merck, Novartis, Takeda and Wave Lifesciences; receives royalties from Cambridge University Press, Elsevier and Oxford University Press; and receives grant funding from the NIA, the National Institute of Neurological Disorders and Stroke, the National Institute of Mental Health and the Bluefield Foundation. K.D.-R. receives research support from the NIH and serves as an investigator for a clinical trial sponsored by Lawson Health Research Institute. S.D. has participated or is currently participating in clinical trials of anti-dementia drugs sponsored by Biogen, Ionis Pharmaceuticals, Wave Life Sciences and Janssen. He has received speaking honorarium/advisory fees from Eisai, Biogen, Innoderm Neurosciences and NeuroCatch and receives salary support from the Fond de Recherche du Québec – Santé. K.F. receives research support from the NIH. J.A.F. receives research support from the NIH. T.F. receives research support from the NIH. L.F. receives research support from the NIH. R.G. receives research support from the NIH. T.F.G. receives research support from the NIH. N.G. has participated or is currently participating in clinical trials of anti-dementia drugs sponsored by Bristol Myers Squibb, Eli Lilly/Avid Radiopharmaceuticals, Janssen Immunotherapy, Novartis, Pfizer, Wyeth, SNIFF (The Study of Nasal Insulin to Fight Forgetfulness) and the A4 (The Anti-Amyloid Treatment in Asymptomatic Alzheimer's Disease) trial. She receives research support from Tau Consortium and the Association for Frontotemporal Dementia and is funded by the NIH. J.G. is serving as a consultant to the Novartis Alzheimer's Prevention Advisory Board. She receives research support from the NIH, Huntington's Disease Society of America and the New York State Department of Health (RFA 1510130358). J.G.-R. receives research support from the NIH and is on the editorial board of *Neurology*. N.G.-R. receives royalties from UpToDate and has participated in multicenter therapy studies by sponsored by Biogen, TauRx, AbbVie, Novartis and Lilly. He receives research support from the NIH. M.G. receives grant support from the NIH, Avid and Piramal; participates in clinical trials sponsored by Biogen, TauRx and Alector; serves as a consultant to Bracco and UCB; and serves on the editorial board of *Neurology*. G.-Y.H. has served as an investigator for clinical trials sponsored by AstraZeneca, Eli Lilly and Roche/Genentech. He receives research support from the Canadian Institutes of Health Research and the Alzheimer Society of British Columbia. E.D.H. receives research support from the NIH. D.I. receives support from the NIH, BrightFocus Foundation and Penn Institute on Aging. D.T.J. receives research support from the NIH and the Minnesota Partnership for Biotechnology and Medical Genomics. K.K. served on the Data Safety Monitoring Board for Takeda Global Research & Development Center and data monitoring boards of Pfizer and Janssen Alzheimer Immunotherapy and received research support from Avid Radiopharmaceuticals, Eli Lilly, the Alzheimer's Drug Discovery Foundation and the NIH. D. Kerwin has served on an advisory board for AbbVie and as site principal investigator for studies funded by Roche/Genentech, AbbVie, Avid, Novartis, Eisai, Eli Lilly and UCSF. D. Knopman serves on the data and safety monitoring board of the DIAN-TU study; is a site principal investigator for clinical trials sponsored by Biogen, Lilly and the University of Southern California; and is funded by the NIH. J. Kornak has provided expert witness testimony for Teva Pharmaceuticals in

Forest Laboratories Inc. et al. v. Teva Pharmaceuticals USA, Inc., case numbers 1:14-cv-00121 and 1:14-cv-00686 (D. Del. filed 31 January 2014 and 30 May 2014 regarding the drug Memantine) and for Apotex/HEC/Ezra in *Novartis AG et al. v. Apotex Inc.*, case number 1:15-cv-975 (D. Del. filed 26 October 2015 regarding the drug Fingolimod). He has also given testimony on behalf of Puma Biotechnology in *Hsingching Hsu et al. vs. Puma Biotechnology, Inc., et al.* 2018 regarding the drug Neratinib. He receives research support from the NIH. J. Kramer receives royalties from Pearson. W.K. receives research funding from AstraZeneca, Biogen, Roche, the Department of Defense and the NIH. M.I.L. receives research support from the NIH. J.L. reports speaker fees from Bayer Vital, Biogen and Roche; consulting fees from Axon Neuroscience and Biogen; author fees from Thieme medical publishers and W. Kohlhammer GmbH medical publishers; non-financial support from AbbVie; and compensation for duty as part-time chief medical officer from MODAG outside the submitted work. I.L. receives research support from the NIH (grants: 2R01AG038791-06A, U01NS100610, U01NS80818, R25NS098999, U19AG063911 -1 and 1R21NS114764-01A1), the Michael J Fox Foundation, the Parkinson Foundation, the Lewy Body Association, CurePSP, Roche, AbbVie, Biogen, Centogene. EIP Pharma, Biohaven Pharmaceuticals, Novartis, Brain Neurotherapy Bio and United Biopharma SRL - UCB. She was a member of the scientific advisory board of Lundbeck and is a scientific advisor for Amydis and Rossy Center for Progressive Supranuclear Palsy University of Toronto. She receives her salary from the University of California, San Diego and as Chief Editor of *Frontiers in Neurology*. P.L. is a site primary investigator for clinical trials by Alector, AbbVie and Woolsey. He serves as an advisor for Retrotrope. He receives research and salary support from the NIH-NIA and the Alzheimer's Association-Part the Cloud partnership. D.L. receives research support from the NIH. I.R.M. receives research funding from the Canadian Institutes of Health Research, the Alzheimer's Association US, the NIH and the Weston Brain Institute. M.M. reports grant funding from the Canadian Institutes of Health Research relating to this work and grants from the Canadian Institutes of Health Research, Woman's Brain Health Initiative, Brain Canada, Ontario Brain Institute, Weston Brain Institute and Washington University outside of this submitted work. M.M. has received personal fees for serving on a scientific advisory committee for Ionis Pharmaceuticals, Alector Pharmaceuticals, Wave Life Sciences and Biogen Canada outside of this submitted work. M.M. has received royalties from Henry Stewart Talks outside of this submitted work. M.M. is a clinical trial site investigator for Roche and Alector Pharmaceuticals outside of this submitted work. S.M. has served as an investigator for clinical trials sponsored by AbbVie, Allon Therapeutics, Biogen, Bristol Myers Squibb, C2N Diagnostics, Eisai, Eli Lilly and Co., Genentech, Janssen Pharmaceuticals, Medivation, Merck, Navidea Biopharmaceuticals, Novartis, Pfizer and TauRx Therapeutics. He receives research support from the NIH. M.F.M. receives research support from the NIH. B.L.M. is Director and Internal Advisor of The Bluefield Project to Cure FTD, Co-Director and Scientific Advisor of the Tau Consortium, Co-Director of the Global Brain Health Institute, Co-Director and medical advisor for The John Douglas French Foundation, scientific advisor for The Larry L. Hillblom Foundation, Scientific Advisor for Association for Frontotemporal Degeneration, Scientific Advisor for National Institute for Health Research Cambridge Biomedical Research Centre and its subunit, the Biomedical Research Unit in Dementia, external advisor to University of Washington ADRC, Stanford University ADRC, Arizona Alzheimer's Disease Center, Massachusetts Alzheimer Disease Research Center, and scientific advisor to The Buck Institute for Research on Aging. B.L.M. serves as Editor-in-Chief for *Neurocase*, on the editorial board of *ALS/FTD Journal* (Taylor & Francis), Section Editor for *Frontiers* and editor for *PLOS Medicine*. He receives royalties from Cambridge University Press, Guilford Publications, Johns Hopkins Press, Oxford University Press, Taylor & Francis Group, Elsevier and Up-to-Date. C.U.O. receives research funding from the NIH, Lawton Health Research Institute, National Ataxia Foundation, Alector and Transposon. He is

also supported by the Robert and Nancy Hall Brain Research Fund, the Jane Tanger Black Fund for Young-Onset Dementias and a gift from Joseph Trovato. He is a consultant with Alector and Acadia Pharmaceuticals. L.P. receives research support from the NIH. M.Q. is an employee of Berry Consultants, where she serves as a consultant to numerous pharmaceutical and device companies. R.R. receives research funding from the NIH and the Bluefield Project to Cure Frontotemporal Dementia. R.R. is on the scientific advisory board of Arkuda Therapeutics and receives royalties from progranulin-related patent. She is also on the scientific advisory board of the Fondation Alzheimer. E.M.R. receives research support from the NIH. K.P.R. receives research support from the NIH and the National Science Foundation and serves on a medical advisory board for Eli Lilly. K.R. receives research support from the NIH. E.D.R. receives research support from the NIH, the Bluefield Project to Cure Frontotemporal Dementia, the Alzheimer's Association, the BrightFocus Foundation, Biogen and Alector; has served as a consultant for AGTC and on a data and safety monitoring board for Lilly; and owns intellectual property related to tau. J.D.R. has served on a medical advisory board and had a consultancy agreement with Alector, Arkuda Therapeutics, Wave Life Sciences, Prevail Therapeutics, UCB, AC Immune, Astex Pharmaceuticals, Biogen, Takeda and Eisai. J.C.R. receives research support from the NIH and is a site principal investigator for clinical trials sponsored by Eli Lilly and Eisai. H.J.R. has received research support from Biogen Pharmaceuticals, has consulting agreements with Wave Neuroscience and Ionis Pharmaceuticals and receives research support from the NIH. J.B.R. has research grants unrelated to the current work from AstraZeneca, Janssen, Lilly and GSK via the Dementias Platform UK and has provided consultancy unrelated to the current work to Asceneuron, Astex, Curasen, UCB, SV Health, WAVE and Alzheimer Research UK. R.S.-V. receives personal fees from Wave Pharmaceuticals for attending advisory board meetings; personal fees from Roche Diagnostics, Janssen and Neuraxpharm for educational activities; and research grants to her institution from Biogen and Sage Therapeutics outside the submitted work. R.S. receives support from the NIA, the National Institute of Neurological Disorders and Stroke, the Parkinson's Disease Foundation and Acadia Pharmaceuticals. A.M.S. received research support from the NIA/NIH, the Bluefield Project to Cure FTD and the Larry L. Hillblom Foundation. He has provided consultation to Passage Bio and Takeda. M.C.T. has served as an investigator for clinical trials sponsored by Biogen, Avanex, Green Valley, Roche/Genentech, Bristol Myers Squibb, Eli Lilly/Avid Radiopharmaceuticals and Janssen. She receives research support from the Canadian Institutes of Health Research. N.T. was employed by the Association for Frontotemporal Degeneration and is now employed by Alector. L.V.V. receives research support from the Alzheimer's Association, the American Academy of Neurology, the American Brain Foundation and the NIH and has provided consultation for Retrotrope. S.W. receives research support from the NIH. B. Wendelberger is an employee of Berry Consultants, where she serves as a consultant to numerous pharmaceutical and device companies. B. Wong receives research support from the NIH. All other authors have no competing interests.

Additional information

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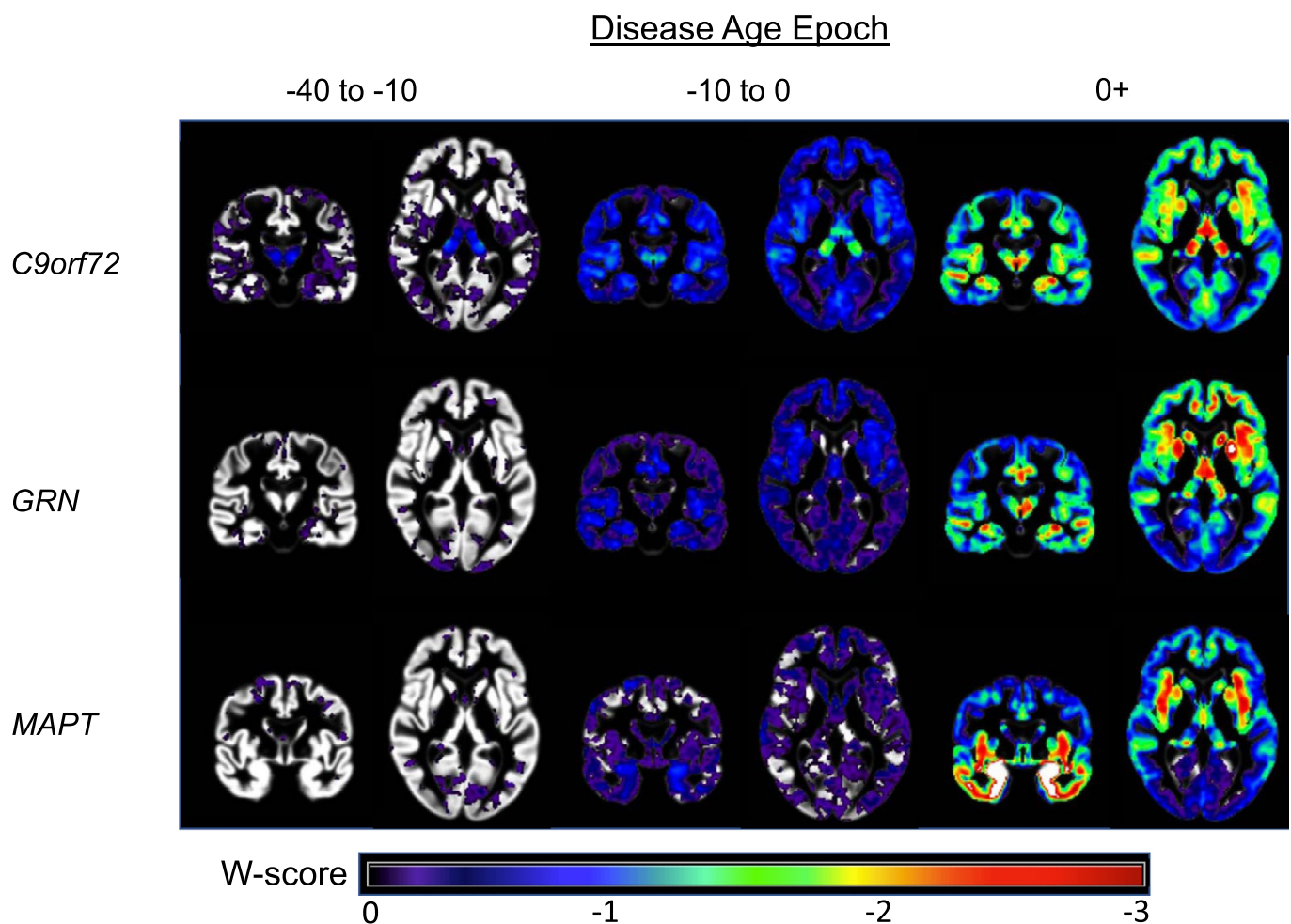
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Domain/Sub-Domain	Measure	Mutation	Disease Age Epoch			
			-40 to -10 YSO	-10 to 0 YSO	0+ YSO	
Global	CDR®+NACC FTLD SB	<i>C9orf72</i>	0.07	0.10	0.41	
		<i>GRN</i>	0.06	0.09	0.52	
		<i>MAPT</i>	0.07	0.15	0.47	
Clinical	Numbers Forward	<i>C9orf72</i>	0.00	0.01	0.10	
		<i>GRN</i>	0.00	0.01	0.22	
		<i>MAPT</i>	0.00	0.00	0.01	
		Numbers Backward	<i>C9orf72</i>	0.00	0.01	0.24
			<i>GRN</i>	0.00	0.01	0.31
			<i>MAPT</i>	0.01	0.00	0.06
		Trails A	<i>C9orf72</i>	0.03	0.04	0.20
			<i>GRN</i>	0.02	0.04	0.29
			<i>MAPT</i>	0.00	0.01	0.11
	Trails B	<i>C9orf72</i>	0.01	0.07	0.32	
		<i>GRN</i>	0.00	0.02	0.50	
		<i>MAPT</i>	0.00	0.01	0.21	
	Language	Semantic Fluency	<i>C9orf72</i>	0.00	0.00	0.34
			<i>GRN</i>	0.00	0.01	0.40
			<i>MAPT</i>	0.00	0.01	0.33
		MINT	<i>C9orf72</i>	0.00	0.00	0.16
			<i>GRN</i>	0.00	0.00	0.30
			<i>MAPT</i>	0.00	0.02	0.40
	Memory	Figure Recall	<i>C9orf72</i>	0.02	0.00	0.18
			<i>GRN</i>	0.00	0.01	0.34
			<i>MAPT</i>	0.00	0.01	0.28
	Visuospatial	Figure Copy	<i>C9orf72</i>	0.00	0.01	0.10
			<i>GRN</i>	0.01	0.01	0.12
			<i>MAPT</i>	0.00	0.04	0.03
	Behavior	RSMS	<i>C9orf72</i>	0.00	0.01	0.52
			<i>GRN</i>	0.00	0.00	0.39
			<i>MAPT</i>	0.01	0.02	0.40
Plasma	NfL (log)	<i>C9orf72</i>	0.05	0.22	0.28	
		<i>GRN</i>	0.04	0.15	0.69	
		<i>MAPT</i>	0.00	0.00	0.28	
Imaging	Frontal Lobe	<i>C9orf72</i>	0.04	0.28	0.39	
		<i>GRN</i>	0.00	0.08	0.41	
		<i>MAPT</i>	0.00	0.01	0.20	
	Temporal Lobe	<i>C9orf72</i>	0.08	0.34	0.27	
		<i>GRN</i>	0.00	0.14	0.25	
		<i>MAPT</i>	0.01	0.00	0.45	
	Medial Temporal Lobe	<i>C9orf72</i>	0.02	0.26	0.21	
		<i>GRN</i>	0.00	0.15	0.21	
		<i>MAPT</i>	0.02	0.07	0.57	
	Parietal Lobe	<i>C9orf72</i>	0.06	0.30	0.29	
		<i>GRN</i>	0.00	0.08	0.21	
		<i>MAPT</i>	0.00	0.01	0.01	
	Occipital Lobe	<i>C9orf72</i>	0.06	0.25	0.22	
		<i>GRN</i>	0.01	0.08	0.08	
		<i>MAPT</i>	0.00	0.01	0.01	
	Insula	<i>C9orf72</i>	0.05	0.23	0.29	
		<i>GRN</i>	0.01	0.12	0.26	
		<i>MAPT</i>	0.01	0.01	0.49	
	Striatum	<i>C9orf72</i>	0.01	0.12	0.21	
		<i>GRN</i>	0.00	0.02	0.37	
		<i>MAPT</i>	0.01	0.00	0.25	
	Thalamus	<i>C9orf72</i>	0.10	0.28	0.23	
		<i>GRN</i>	0.00	0.06	0.15	
		<i>MAPT</i>	0.00	0.01	0.06	
	Cerebellum	<i>C9orf72</i>	0.01	0.12	0.09	
		<i>GRN</i>	0.00	0.05	0.10	
		<i>MAPT</i>	0.03	0.05	0.01	

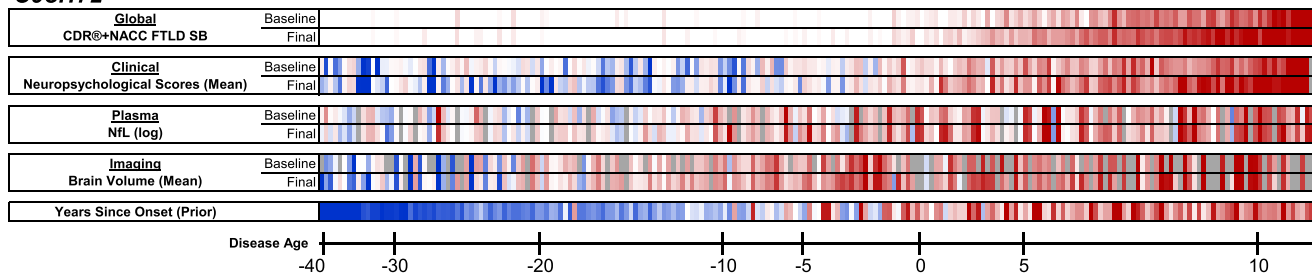
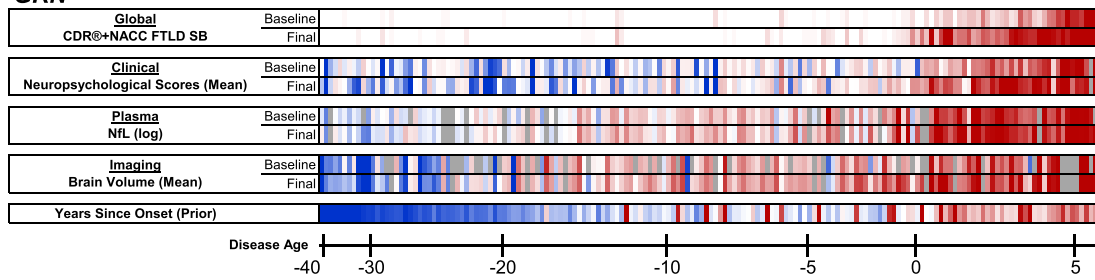
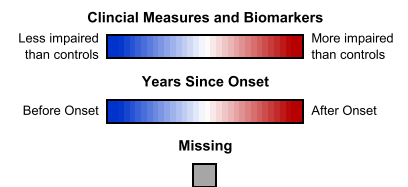
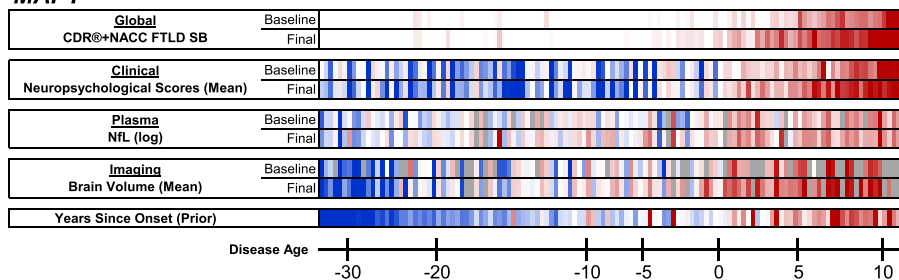


Extended Data Fig. 1 | See next page for caption.

Extended Data Fig. 1 | Baseline comparisons between mutation carriers and controls by DA epoch. Cross-sectional baseline differences between mutation carriers and controls are presented as effect sizes (omega squared). Bolded cells indicate statistical significance ($p < 0.05$) using two-sided tests without multiple comparison correction. Comparisons in which mutation carriers are more impaired than controls are colored, with darker shades illustrating larger effect sizes. Note that statistical comparisons for the CDR+NACC-FTLD SB should be interpreted with caution given that controls were defined as having a baseline CDR+NACC-FTLD=0 and thus have no variance due to this selection process. *Abbreviations: EF: Executive Functioning; NfL: Plasma neurofilament light chain levels; RSMS: Revised Self-Monitoring Scale. CDR+NACC FTLD SB: Clinical Dementia Rating Scale plus National Alzheimer's Coordinating Center's Frontotemporal Lobar Degeneration Module Sum of Boxes.*



Extended Data Fig. 2 | Voxelwise atrophy by estimated disease stage in familial frontotemporal dementia. Voxelwise maps display brain atrophy as the number of standardized units from controls (W -scores), controlling for head size and scanner. Images are shown in radiological orientation (that is, right is left). Voxelwise results are presented with a greater number of axial slices in Supplementary Figs. 2–4. Results were generally consistent with the region of interest findings, supporting the validity of the DPM approach. In *C9orf72*, thalamic atrophy, particularly in the pulvinar, was the primary region of atrophy in the –40 to –10 epoch and continued to be a region of prominent atrophy throughout the disease course. Medial temporal lobe volume loss became prominent in the –10 to –0 epoch. Frontoinsular, medial parietal, and medial temporal atrophy became prominent in the symptomatic phase (see also Supplementary Fig. 2). In *GRN*, subtle early cerebellar atrophy was observed (–40 to –10), along with atrophy in frontotemporal, subcortical, and insular structures in the –10 to 0 epoch. In the symptomatic stage, atrophy extended into the temporal lobe, frontoparietal regions, and striatum (see also Supplementary Fig. 3). Atrophy in *MAPT* appeared to begin in the medial temporal lobe and temporal pole (–10 to 0), and symptomatic mutation carriers showed temporal, insular, ventral and medial frontal, and striatal atrophy (see also Supplementary Fig. 4).

C9orf72**GRN****MAPT**

Extended Data Fig. 3 | Patient-level data contributing to the disease progression models. For each genetic group, each mutation carrier with longitudinal data is displayed in a single column, organized on the x-axis by their model estimated Disease Age at baseline. Participants' baseline and last available (Final) observation for each outcome are presented. For the CDR+NACC-FTLD-SB, white cells indicate a score of 0, and increasingly dark red tones denote higher scores (that is, more severe impairments or greater atrophy). Log-transformed plasma NfL concentrations and the mean of all available neuropsychological scores and regional gray matter volume estimates are also presented, with the color scale indicating their scores relative to controls of the same Disease Age. Lastly, the model's prior estimate of Years Since Onset is displayed. For participants with documented onset, we display the difference between their chronological age and the clinician estimated age of onset. For those participants in whom clinical onset has not yet occurred (or this data was unavailable), we display the difference between their chronological age and the mean age of onset for their mutation.

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- A description of all covariates tested
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Give P values as exact values whenever suitable.
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Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection	For ALLFTD, clinical, neuropsychological, and biomarker data are recorded in a customized iMedidata RAVE database. MRI images and metadata are uploaded and entered into a restricted access archive at the Laboratory of NeuroImaging (LONI). For GENFI, data entry is carried out via a customised XNAT database, specifically designed to accept all data collected as part of the GENFI study including MRI, clinical and neuropsychological data.
Data analysis	Data analyses were performed using the R (1.4.2) JAGS package with custom code available through Zenodo (10.5281/zenodo.6687486), Stata (17.0), nonparametric nonuniform intensity normalization (N3) algorithm, and SPM12.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

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The datasets analyzed for the current study reflect collaborative efforts of two research consortia: ALLFTD and GENFI. Each consortium provides clinical data access based on established policies for data use: processes for request are available for review at allftd.org/data for ALLFTD data and by emailing genfi@ucl.ac.uk for

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Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	The sample for this study drew from all available participants with this condition from the two major United States and European consortia. The sample size for this study is over 1000 participants, which gives us broad representation across the disease spectrum. Between group comparisons included a minimum of 138 participants, which provides sufficient power to detect clinically relevant effect sizes for between-group comparisons.
Data exclusions	The parent studies from which these data were drawn have exclusion criteria that are described in the manuscript. All available data were included in the study.
Replication	To verify the reproducibility of our findings, we fit models in two consortia separately. These separate models, including raw data points, are presented in the manuscript and illustrated the high degree of replicability in progression across cohorts.
Randomization	This study built models using observational data to model the natural history of familial frontotemporal dementia. Randomization was not relevant to our study design.
Blinding	This study built models using observational data. The presence or absence of a mutation was relevant to model parameters and therefore was required to build the models. Randomization was not performed and therefore no blinding occurred related to randomization.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

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<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology	<input type="checkbox"/>	<input checked="" type="checkbox"/> MRI-based neuroimaging
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms		
<input type="checkbox"/>	<input checked="" type="checkbox"/> Human research participants		
<input type="checkbox"/>	<input checked="" type="checkbox"/> Clinical data		
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern		

Antibodies

Antibodies used

ALLFTD plasma NfL Measurement: Plasma NfL light concentrations were measured at the Mayo Clinic in Jacksonville using the Quanterix (Lexington, MA) single-molecule array technology (Simoa) @ NF-Light Advantage Kit (Cat#103186, Lot 501992) and the HD-X instrument according to the instructions provided

GENFI plasma NfL measurement: Plasma NfL concentrations were measured at baseline with single-molecule array technology (Simoa), using the commercially available Simoa Neurology 4-Plex A kit (Quanterix, Lexington, MA, Cat# 102153).

Validation

ALLFTD: Samples were tested in duplicate using kits from the same lot. In addition to the two quality control samples provided with the kit, all assays included five inter-assay controls. Prior to each assay, plasma samples were thawed, mixed thoroughly by low-speed vortexing, centrifuged at 10,000 g for five minutes, and transferred to 96-well plates that were then sealed to minimize sample evaporation. Samples were diluted four times by the instrument. If levels of NfL in a sample exceeded the upper limit of the calibration curve, the sample was retested at a higher dilution. Across all assays, the percent coefficient of variations of the mean NfL concentration for the inter-assay controls were below 10%.

GENFI: Plasma samples were thawed at room temperature (one cycle), mixed thoroughly, and centrifuged at 14,000g for 3 minutes. The supernatant was loaded onto a Quanterix HD-1 Analyzer with a 1:4 specified dilution. Measures were completed in duplicate

over a total of six batches, each with an eight-point calibration curve tested in triplicate and two controls tested in duplicate. Plasma concentrations were interpolated from the calibration curve within the same batch and corrected for the dilution. All samples were quantifiable within the dynamic range of 0.69 to 2,000 pg/mL and with an average coefficient of variation below 10%. Instrument operators were blinded to clinical and genetic information.

Human research participants

Policy information about [studies involving human research participants](#)

Population characteristics	This study enrolled participants from families with known familial FTD mutations. Participants above the age of 18 were included. Participants were included across a range of clinical phenotypes, with most participants presenting with behavioral variant FTD or primary progressive aphasia. We consider the impacts of known covariates such as age (mean=50.2, SD=1.9), sex (56.1% female), education (mean=14.4, SD=3.2), and language of testing.
Recruitment	Participants were recruited from the major familial FTD research consortia in the United States and Europe. Participants were recruited from subjects/kindreds already identified at the collaborating centers. Referrals were also solicited from other centers interested in familial FTD, non-profit organizations that support FTD research and patient advocacy, and consortia websites. There is currently a lack of ethnic and sociocultural diversity in FTD research populations which may limit the generalizability of these results to underrepresented populations. Patients were recruited from academic research centers that are specialized in these diseases. This may not reflect true population.
Ethics oversight	The ALLFTD study was approved through the Trial Innovation Network (TIN) at Johns Hopkins University. Local ethics committees (Institutional Review Boards) at each of the sites approved the study, and all participants provided written informed consent or assent with proxy consent.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Clinical data

Policy information about [clinical studies](#)

All manuscripts should comply with the ICMJE [guidelines for publication of clinical research](#) and a completed [CONSORT checklist](#) must be included with all submissions.

Clinical trial registration	NCT02365922; NCT04363684; NCT02372773
Study protocol	https://clinicaltrials.gov/ct2/show/NCT02365922 ; https://clinicaltrials.gov/ct2/show/NCT04363684 ; https://clinicaltrials.gov/ct2/show/NCT02372773
Data collection	Participants were enrolled through Advancing Research and Treatment for Frontotemporal Lobar Degeneration (ARTFL) and Longitudinal Evaluation of Familial Frontotemporal Dementia Subjects (LEFFTDS), which recently combined into the ARTFL/LEFFTDS Longitudinal Frontotemporal Lobar Degeneration (ALLFTD) Study. ALLFTD enrolled participants through a consortium of 18 centers across the US and Canada between 2015 and 2020. Participants were also enrolled through the Genetic Frontotemporal Initiative (GENFI), which involves 25 research centers across Europe and Canada. GENFI participants from the 5th Data Freeze (2015-2019) were included
Outcomes	Data was acquired from ongoing observational studies. Outcomes were clinical measures and biomarkers that were readily available across the GENFI and ALLFTD consortia and are related to the symptoms of FTD. The Clinical Dementia Rating Scale (CDR [®]) plus Behavioral and Language Domains from the National Alzheimer's Coordinating Center (NACC) FTLD module, neuropsychological tests from the Uniform Data Set and the Revised Self Monitoring Scale were scored according to standardized procedures. Volumetric brain MRI was analyzed using predefined regions of interests and voxelwise analyses. Plasma neurofilament light chain levels were assessed using Quanterix Simoa technology as described below.

Magnetic resonance imaging

Experimental design

Design type	Region of interest comparison and voxelwise display of effect sizes
Design specifications	Gray matter volume in each of several preselected ROIs was entered into the model as a percentage of total intracranial volume. In addition, we conducted sensitivity analyses in which gray matter volume at each voxel was converted to a W-scores based on a reference group, covarying for head size and scanner. The mean W-score for each region of interest was extracted and modeled in the disease progression model. Voxelwise W-scores were also presented as a sensitivity analysis.
Behavioral performance measures	No behavioral data was gathered during this volumetric sequence.

Acquisition

Imaging type(s)	Structural
Field strength	1.5 and 3 Tesla.
Sequence & imaging parameters	T1-weighted images from ALLFTD were acquired as Magnetization Prepared Rapid Gradient Echo (MP-RAGE) images

Sequence & imaging parameters	using the following parameters: 240x256x256 matrix; about 170 slices; voxel size = 1.05x1.05x1.25 mm ³ ; flip angle, TE and TR varied by vendor. T1-weighted images from GENFI were acquired using the following parameters: 256x256x208 matrix; 208 slices; voxel size = 1.1 mm isotropic, flip angle = 8 degrees, TE and TR varied by vendor.
Area of acquisition	Whole brain scans.
Diffusion MRI	<input type="checkbox"/> Used <input checked="" type="checkbox"/> Not used

Preprocessing

Preprocessing software	T1-weighted images underwent bias field correction using N3 algorithm. The segmentation was performed using SPM12 (Wellcome Trust Center for Neuroimaging, London, UK, http://www.fil.ion.ucl.ac.uk/spm) unified segmentation.
Normalization	A customized group template was generated from the segmented gray and white matter tissues and cerebrospinal fluid by non-linear registration template generation using the Large Deformation Diffeomorphic Metric Mapping framework. ¹² Subjects' native space gray and white matter were geometrically normalized to the group template, modulated, and then smoothed in the group template. The applied smoothing used a Gaussian kernel with 8~mm full width half maximum. Every step of the transformation was carefully inspected from the native space to the group template.
Normalization template	ICBM
Noise and artifact removal	The applied smoothing used a Gaussian kernel with 8~mm full width half maximum.
Volume censoring	Volume censoring was not performed.

Statistical modeling & inference

Model type and settings	Modeling was not performed on the raw images; regions of interest were extracted and included in the disease progression models along with other clinical variables and plasma NfL levels, and thus were treated like other clinical measures. Voxelwise images were used for display purposes and were not used to conduct statistical tests.
Effect(s) tested	Statistical tests were not performed on the raw images; regions of interest were extracted and included in the disease progression models along with other clinical variables and plasma NfL levels.
Specify type of analysis:	<input type="checkbox"/> Whole brain <input type="checkbox"/> ROI-based <input checked="" type="checkbox"/> Both
Anatomical location(s)	Regions of interest were extracted using the Desikan atlas. Parcellation was conducted based on historical precedence in this disease.
Statistic type for inference (See Eklund et al. 2016)	Statistical inference was not performed at the voxel or cluster level.
Correction	For between-group ROI comparisons, multiple comparisons were controlled for using the Tukey method.

Models & analysis

n/a	Involvement in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> Functional and/or effective connectivity
<input checked="" type="checkbox"/>	<input type="checkbox"/> Graph analysis
<input checked="" type="checkbox"/>	<input type="checkbox"/> Multivariate modeling or predictive analysis