




Selective carbonic anhydrase IX and XII inhibitors based around a functionalized coumarin scaffold

Bader I. Huwaimel^{1,2}  | Sravan K. Jonnalagadda¹ | Shirisha Jonnalagadda¹  | Shikha Kumari¹ | Alessio Nocentini³ | Claudiu T. Supuran³  | Paul C. Trippier^{1,4,5} 

¹Department of Pharmaceutical Sciences, College of Pharmacy, University of Nebraska Medical Center, Omaha, Nebraska, USA

²Department of Pharmaceutical Chemistry, College of Pharmacy, University of Ha'il, Ha'il, Saudi Arabia

³Polo Scientifico, Laboratorio di Chimica Bioinorganica, Università degli Studi di Firenze, Sesto Fiorentino, Florence, Italy

⁴Fred & Pamela Buffett Cancer Center, University of Nebraska Medical Center, Omaha, Nebraska, USA

⁵UNMC Center for Drug Discovery, University of Nebraska Medical Center, Omaha, Nebraska, USA

Correspondence

Paul C. Trippier, Department of Pharmaceutical Sciences, College of Pharmacy, University of Nebraska Medical Center, Omaha, NE 68106, USA.

Email: paul.trippier@unmc.edu

Funding information

National Cancer Institute, Grant/Award Numbers: R01CA226436, P30CA036727; National Institute for Child Health and Human Development, Grant/Award Number: R01HD106590

Abstract

Inhibition of specific carbonic anhydrase (CA) enzymes is a validated strategy for the development of agents to target cancer. The CA isoforms IX and XII are overexpressed in various human solid tumors wherein they play a critical role in regulating extracellular tumor acidification, proliferation, and progression. A series of novel sulfonamides based on the coumarin scaffold were designed, synthesized and characterized as potent and selective CA inhibitors. Selected compounds show significant activity and selectivity over CA I and CA II to target the tumor-associated CA IX and CA XII with high inhibition activity at the single digit nanomolar level. Twelve compounds were identified to be more potent compared with acetazolamide (AAZ) control to inhibit CA IX while one was also more potent than AAZ to inhibit CA XII. Compound **18f** (K_i's = 955 nM, 515 nM, 21 nM and 5 nM for CA's I, II, IX, and XII, respectively) is highlighted as a novel CA IX and XII inhibitor for further development.

KEYWORDS

carbonic anhydrase IX inhibitors, carbonic anhydrases XII inhibitor, coumarin, structure–activity relationship

1 | INTRODUCTION

The carbonic anhydrases (CA) are a family of ubiquitous zinc enzymes which play a catalytic role in the reversible hydration of carbon dioxide (CO₂) and water (H₂O) to bicarbonate (HCO₃⁻) and a proton (Supuran, 2008). In humans, the CA enzymes have 15 isoforms that vary by localization and catalytic activity including; the cytosolic CAs; CA I, CA II, CA III, CA VII, CA XIII; the membrane-bound CAs; CA IV,

CA IX, CA XII, CA XIV, CA XV (not present in primates, only in rodents and other animals/fish); CA VA and CA VB are mitochondrial, and CA VI is secreted in saliva and colostrum. In addition, three catalytically inactive forms of CA are known (CA VIII, CA X, and CA XI) referred to as CA related-proteins (Lomelino et al., 2016). CA IX and CA XII are highly overexpressed genes in response to hypoxia in a variety of human solid tumors such as breast, colorectal, glioblastoma and lung, where they play a critical role in regulating tumor

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2023 The Authors. *Drug Development Research* published by Wiley Periodicals LLC.

acidification, proliferation, and progression (Lee & Griffiths, 2020; Neri & Supuran, 2011).

The overexpression of CA IX and CA XII induces cancer cell growth, activation of the metastatic cascade, and reduced response to chemotherapy (Supuran, 2010). Targeting both CA IX and CA XII in cancers that overexpress these biomarkers, and suppressing their activity has been shown to be therapeutically beneficial in the treatment of tumors (McDonald et al., 2022). The classic CA inhibitors contain the sulfonamide pharmacophore and have been determined to exhibit potent CA IX and XII inhibition with high potency to attenuate cancer cell growth both in vitro and in vivo. Examples of this class include acetazolamide, dichlorphenamide and dorzolamide (Table 1) (Supuran, 2008). Recently, coumarin-based small molecules (**1.1** and **1.2**) were reported as nonclassical CA inhibitors with high efficacy and selectivity for the physiologically dominant tumor-associated isoenzymes CA IX and CA XII (Maresca et al., 2010; Supuran, 2008, 2020; Thacker et al., 2019; Tousni et al., 2011; Williams & Gieling, 2019). CA inhibitors with combined sulfonamide and coumarin moieties have been reported to possess high efficacy for inhibiting the enzymatic activity of CA IX (**1.3–1.7**, Table 1) (Wang et al., 2013). Moreover, sulfonamides containing coumarin moieties have potent anticancer activity in the MCF-7 breast cancer cell line (Wang et al., 2013). Although several substituted coumarins have been described in the literature, little information is known about the importance of these coumarin structures as CA inhibitors.

Herein, the synthesis of a series of CA inhibitors is reported based around the coumarin chemotype with substituted sulfonamide moieties to further investigate the amalgamation of these two pharmacophores as CA inhibitors. Several of the synthesized derivatives possess high potency for inhibiting the tumor-associated CA IX and CA XII with nanomolar activity while possessing selectivity over CA I and II.

2 | EXPERIMENTAL

2.1 | Chemistry

All reactions were carried out in oven- or flame-dried glassware under positive nitrogen pressure unless otherwise noted. Reaction progress was monitored by thin-layer chromatography (TLC) carried out on silica gel plates (2.5 cm × 7.5 cm, 200 μm thick, 60 F254) and visualized by using UV (254 nm) or by potassium permanganate or phosphomolybdic acid stain as indicator. Flash column chromatography was performed with silica gel (40–63 μm, 60 Å) or on a Biotage[®] automated system (Biotage[®] Selekt). Commercial grade solvents and reagents were purchased from Fisher Scientific or Sigma Aldrich and were used without further purification except as indicated. Anhydrous solvents were purchased from Acros Organics and stored under an atmosphere of dry nitrogen over molecular sieves.

¹H and ¹³C NMR spectra were recorded in the indicated deuterated solvent on a Bruker Advance III HD spectrometer at

400 or 500 for ¹H and 100 or 126 MHz for ¹³C respectively with solvent peak as an internal standard. Multiplicities are indicated by s (single), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), m (multiplet), and br (broad). Chemical shifts (δ) are reported in parts per million (ppm) and coupling constants (*J*) in Hertz. High-resolution mass spectroscopy (HRMS) was performed on a 6230 LC/TOF (Agilent) using an ESI source. The spectral data was extracted from total ion chromatogram (TIC).

2.1.1 | Ethyl 7-hydroxy-2-oxo-2H-chromene-3-carboxylate (2a)

To a mixture of 2,4-dihydroxybenzaldehyde (**1a**, 2 g, 14.48 mmol) in ethanol (5 mL) at room temperature, diethyl malonate (5.81 g, 2.43 mL, 15.23 mmol) was added along with piperidine (0.29 mL, 2.9 mmol). The mixture was stirred and heated to 60°C for 2 h. Then, it was cooled to room temperature and filtered, washed with water and ethanol, and air dried to yield a yellow powder (2.954 g, 87% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 1.30 (t, *J* = 7.1 Hz, 3H), 4.24–4.29 (q, *J* = 7.1 Hz, 2H), 6.73 (s, 1H), 6.86 (d, *J* = 8.5 Hz, 1H), 7.77 (d, *J* = 8.5 Hz, 1H), 8.67 (s, 1H), 11.06 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 14.6, 61.2, 102.2, 110.8, 112.5, 114.4, 132.5, 149.8, 156.8, 157.5, 163.4, 164.5. HRMS (ESI): *m/z* calcd for C₁₂H₁₀O₅ [M+Na]⁺: 257.1939, found: 257.1932.

2.1.2 | 2-Chloro-*N*-(4-sulfamoylphenyl)acetamide (4a)

Potassium carbonate (K₂CO₃) (1.2 g, 8.72 mmol) was added to a solution of sulfanilamide (**3a**, 1 g, 5.81 mmol) in THF (20 mL). Chloroacetyl chloride (0.56 mL, 6.97 mmol) was added to the above solution dropwise and under N₂ atmosphere at 0°C with stirring. After that, the reaction mixture was stirred for 2 h and then water was added to quench the reaction. The reaction mixture was extracted with ethyl acetate, the organic layer washed with brine and dried over sodium sulfate, the solvent was removed in vacuo and purified by recrystallization in hexane: MeOH to yield a white powder (1.349 g, 93% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 4.30 (s, 2H), 7.27 (s, 2H), 7.74–7.81 (q, *J* = 7.8 Hz, 4H), 10.61 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 44.0, 119.4, 127.2, 139.4, 141.7, 165.6. HRMS (ESI): *m/z* calcd for C₈H₉ClN₂O₃S [M+Na]⁺: 271.6736, found: 271.6731.

2.1.3 | 2-Chloro-*N*-(4-(*N*-(5,6-dimethoxypyrimidin-4-yl)sulfamoyl)phenyl)acetamide (4b)

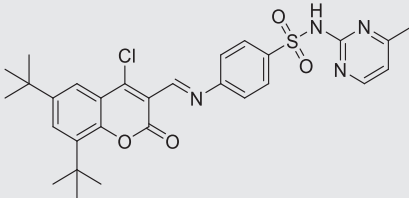
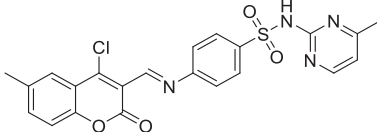
The powder compound was obtained from sulfadoxine (**3b**, 1 g, 3.22 mmol) by following the experimental conditions described for **4a** (0.983 g, 79% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 3.70 (s, 3H), 3.90 (s, 3H), 4.29 (s, 2H), 7.78 (d, *J* = 8.7 Hz, 2H), 7.96 (d, *J* = 8.7 Hz,

TABLE 1 Structures and inhibition profile of selected known carbonic anhydrase inhibitors.

Compound	Structure	K_i (μM)		
		CA II	CA IX	CA XII
Acetazolamide (AAZ)		0.012	0.026	0.006
SLC-0111 (U-104)		9.6	0.045	0.004
Dichlorophenamide		0.038	0.05	0.05
Dorzolamide		0.009	0.052	0.004
1.1		>100	0.2	0.2
1.2		94.3	0.61	7.7
1.3		0.023	0.124	NA
1.4		0.173	0.090	NA
1.5		0.103	0.074	NA

(Continues)

TABLE 1 (Continued)

Compound	Structure	K_i (μM)		
		CA II	CA IX	CA XII
1.6		0.063	0.024	NA
1.7		0.061	0.048	NA

Abbreviation: NA, not available.

2H), 8.11 (s, 1H), 10.67 (s, 1H), 11.04 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 43.9, 54.5, 60.6, 119.2, 127.7, 129.3, 135.5, 142.8, 150.7, 151.0, 162.1, 165.7. HRMS (ESI): m/z calcd for $\text{C}_{14}\text{H}_{15}\text{ClN}_4\text{O}_5\text{S}$ [$\text{M}+\text{Na}$] $^+$: 409.7968, found: 409.7947.

2.1.4 | *N*-(4-(*N*-Acetylsulfamoyl)phenyl)-2-chloroacetamide (4c)

The white powder was obtained from sulfacetamide (3c, 1 g, 4.67 mmol) by following the experimental conditions described for 4a (1.076 g, 80% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.32 (s, 3H), 4.31 (s, 2H), 7.78 (d, J = 8.7 Hz, 2H), 7.89 (d, J = 8.7 Hz, 2H), 10.72 (s, 1H), 11.99 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 23.6, 44.0, 119.3, 129.4, 134.0, 143.4, 165.8, 169.1. HRMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_{11}\text{ClN}_2\text{O}_4\text{S}$ [$\text{M}+\text{Na}$] $^+$: 313.7102, found: 313.7101.

2.1.5 | Ethyl 2-oxo-7-(2-oxo-2-((4-sulfamoylphenyl)amino)ethoxy)-2*H*-chromene-3-carboxylate (5a)

To a solution of ethyl 7-hydroxy-2-oxo-2*H*-chromene-3-carboxylate (2a 0.2 g, 0.85 mmol) in dry *N,N*-dimethylformamide (15 mL), anhydrous K_2CO_3 (0.18 g, 1.28 mmol) was added. The solution was stirred for 15 min at 70–80°C and 2-chloro-*N*-(4-sulfamoylphenyl) acetamide (4a, 0.23 g, 0.94 mmol) was added, followed by a pinch of potassium iodide (KI), and heated overnight. After that, the water (10 mL) was added to the reaction mixture, followed by 1 mL 6 N HCl. The resulting solid was filtered, washed with water, and air dried, and purification by flash column chromatography (hexane/EtOAc 20:1) afforded the title compound as a brown powder (0.21 g, 55% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 1.30 (t, J = 7.1 Hz, 3H), 3.70 (s, 3H), 4.90 (s, 3H), 4.27–4.29 (q, J = 7.3 Hz, 2H), 4.95 (s, 2H), 7.09 (dd, J = 7.1, 2.2 Hz, 2H), 7.27 (s, 2H), 7.79 (s, 4H), 7.87 (d, J = 8.5 Hz, 1H), 8.73 (s, 1H), 10.52 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 14.5,

61.4, 67.7, 101.6, 112.4, 113.9, 114.2, 119.7, 127.1, 132.1, 139.3, 141.6, 149.5, 156.6, 157.1, 163.2, 163.6, 166.6. HRMS (ESI): m/z calcd for $\text{C}_{20}\text{H}_{18}\text{N}_2\text{O}_8\text{S}$ [$\text{M}+\text{Na}$] $^+$: 469.4179, found: 469.4138.

2.1.6 | Ethyl 7-(2-((4-(*N*-(5,6-dimethoxypyrimidin-4-yl)sulfamoyl)phenyl)amino)-2-oxoethoxy)-2-oxo-2*H*-chromene-3-carboxylate (5b)

The yellow powder was obtained from 2-chloro-*N*-(4-(*N*-(5,6-dimethoxypyrimidin-4-yl)sulfamoyl)phenyl)acetamide (4b) by following the experimental conditions described for 5a (0.197 g, 61% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 1.31 (t, J = 7.0 Hz, 3H), 4.25–4.30 (q, J = 7.0 Hz, 2H), 4.95 (s, 2H), 7.07 (dd, J = 7.3, 2.1 Hz, 2H), 7.81 (d, J = 8.8 Hz, 2H), 7.87 (d, J = 8.5 Hz, 1H), 7.95 (d, J = 8.8 Hz, 2H), 8.11 (s, 1H), 8.73 (s, 1H), 10.56 (s, 1H), 11.04 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 14.5, 54.5, 60.7, 61.4, 67.7, 101.6, 112.4, 113.9, 114.2, 119.4, 127.6, 129.3, 132.1, 135.4, 149.5, 150.8, 151.1, 157.1, 162.1, 163.2, 163.6, 166.7. HRMS (ESI): m/z calcd for $\text{C}_{26}\text{H}_{24}\text{N}_4\text{O}_{10}\text{S}$ [$\text{M}+\text{Na}$] $^+$: 607.5418, found: 607.5406.

2.1.7 | Ethyl 7-(2-((4-(*N*-acetylsulfamoyl)phenyl)amino)-2-oxoethoxy)-2-oxo-2*H*-chromene-3-carboxylate (5c)

The yellow powder was obtained from *N*-(4-(*N*-acetylsulfamoyl)phenyl)-2-chloroacetamide (4c) by following the experimental conditions described for 5a (0.22 g, 58% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 1.28–1.32 (m, 3H), 1.91 (s, 1H), 4.25–4.30 (q, J = 7.3 Hz, 2H), 4.96 (s, 2H), 7.08 (d, J = 9.9 Hz, 2H), 7.83–7.89 (m, 5H), 8.73 (d, J = 2.3 Hz, 1H), 10.63 (s, 1H), 11.98 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 14.5, 23.6, 61.4, 67.6, 101.6, 112.4, 113.9, 114.2, 119.5, 119.6, 129.3, 132.1, 133.9, 149.5, 156.6, 157.1, 163.2, 163.6, 166.8, 169.1. HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{20}\text{N}_2\text{O}_9\text{S}$ [$\text{M}+\text{Na}$] $^+$: 511.4545, found: 511.4518.

2.1.8 | 7-Hydroxy-2-oxo-2H-chromene-3-carboxylic acid (7a)

To a solution of ethyl 7-hydroxy-2-oxo-2H-chromene-3-carboxylate (**2a**, 2 g, 8.54 mmol) in MeOH (15 mL) and water (12 mL) was added 2 N NaOH solution (40 mL). The solution was heated to reflux for 12 h, then cooled and concentrated in vacuo. The crude product was diluted with water (10 mL) and acidified with an aqueous solution of 6 N HCl. The resulting solid was filtered, washed with water, and air-dried to provide the product as a yellow solid. (1.71 g, 97% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 6.73 (s, 1H), 6.85 (dd, J = 8.6, 2.1 Hz, 1H), 7.72 (d, J = 8.6 Hz, 1H), 8.66 (s, 1H), 11.10 (br-s, 1H), 12.93 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 103.1, 111.0, 113.0, 114.5, 132.4, 149.7, 157.4, 158.3, 164.1, 164.5 ppm. HRMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_6\text{O}_5$ $[\text{M}+\text{Na}]^+$: 229.1406, found: 229.1401.

2.1.9 | 6-Chloro-2-oxo-2H-chromene-3-carboxylic acid (7b)

The white powder was obtained from ethyl 6-chloro-2-oxo-2H-chromene-3-carboxylate (**2b**, 2 g, 7.92 mmol) by following the experimental conditions described for **7a** (1.725 g, 97% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.49 (d, J = 8.8 Hz, 1H), 7.75 (dd, J = 8.8, 2.5 Hz, 1H), 8.04 (d, J = 2.5 Hz, 1H), 8.69 (s, 1H), 13.38 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 118.6, 119.8, 120.0, 128.8, 129.4, 134.0, 147.4, 153.5, 156.5, 164.1. HRMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_5\text{ClO}_4$ $[\text{M}+\text{Na}]^+$: 247.5861, found: 247.5859.

2.1.10 | 6-Bromo-2-oxo-2H-chromene-3-carboxylic acid (7c)

The yellow powder was obtained from ethyl 6-bromo-2-oxo-2H-chromene-3-carboxylate (**2c**, 2 g, 6.73 mmol) by following the experimental conditions described for **7a** (1.8 g, 99% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.42 (d, J = 8.8 Hz, 1H), 7.86 (dd, J = 8.8, 2.5 Hz, 1H), 8.17 (d, J = 2.5 Hz, 1H), 8.69 (s, 1H), 13.37 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 118.6, 119.8, 120.1, 128.9, 129.3, 134.1, 148.0, 153.1, 156.3, 164.6. RMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_5\text{BrO}_4$ $[\text{M}+\text{Na}]^+$: 292.0370, found: 292.0361.

2.1.11 | 6,8-Dichloro-2-oxo-2H-chromene-3-carboxylic acid (7d)

The yellow powder was obtained from ethyl 6,8-dichloro-2-oxo-2H-chromene-3-carboxylate (**2d**, 2 g, 6.97 mmol) by following the experimental conditions described for **7a** (1.65 g, 91% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 8.04 (d, J = 2.5 Hz, 1H), 8.06 (d, J = 2.5 Hz, 1H), 8.71 (s, 1H), 13.51 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 116.7, 118.44, 120.02, 129.0, 129.6,

133.9, 148.5, 153.4, 156.8, 164.0. HRMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_4\text{Cl}_2\text{O}_4$ $[\text{M}+\text{Na}]^+$: 282.0308, found: 282.0301.

2.1.12 | 2-Oxo-2H-chromene-3-carboxylic acid (7e)

The white powder was obtained from ethyl 2-oxo-2H-chromene-3-carboxylate (**2e**, 2 g, 9.17 mmol) by following the experimental conditions described for **7a** (1.7 g, 98% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.37–7.44 (m, 2H), 7.73 (t, J = 7.6 Hz, 1H), 7.91 (d, J = 7.7 Hz, 1H), 8.74 (s, 1H), 13.24 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 116.5, 118.4, 118.7, 125.3, 130.6, 134.6, 148.7, 154.8, 157.1, 164.3. HRMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_6\text{O}_4$ $[\text{M}+\text{Na}]^+$: 213.1414, found: 213.1410.

2.1.13 | 6-Methoxy-2-oxo-2H-chromene-3-carboxylic acid (7f)

The yellow powder was obtained from ethyl 6-methoxy-2-oxo-2H-chromene-3-carboxylate (**2f**, 2 g, 8.06 mmol) by following the experimental conditions described for **7a** (1.71 g, 96% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.80 (s, 3H), 4.78 (br-s, 1H), 7.31 (dd, J = 9, 3 Hz, 1H), 7.35 (d, J = 9 Hz, 1H), 7.44 (d, J = 3 Hz, 1H), 8.66 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.2, 112.3, 117.7, 118.8, 118.9, 122.5, 148.5, 149.3, 156.1, 157.4, 164.4. HRMS (ESI): m/z calcd for $\text{C}_{11}\text{H}_8\text{O}_5$ $[\text{M}+\text{Na}]^+$: 243.1673, found: 243.1652.

2.1.14 | 7-(Diethylamino)-2-oxo-2H-chromene-3-carboxylic acid (7g)

The brown powder was obtained from ethyl 7-(diethylamino)-2-oxo-2H-chromene-3-carboxylate (**2g**, 2 g, 6.91 mmol) by following the experimental conditions described for **7a** (1.53 g, 84% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 1.14 (t, J = 7.0 Hz, 6H), 3.47 (q, J = 7.0 Hz, 4H), 6.56 (d, J = 2.0 Hz, 1H), 6.81 (dd, J = 9.0, 2.0 Hz, 1H), 7.63 (d, J = 9.0 Hz, 1H), 8.58 (s, 1H), 12.49 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 12.7, 44.8, 96.3, 107.6, 107.8, 110.5, 132.3, 149.9, 153.3, 158.3, 159.9, 164.9. HRMS (ESI): m/z calcd for $\text{C}_{14}\text{H}_{15}\text{NO}_4$ $[\text{M}+\text{Na}]^+$: 284.2621, found: 284.2619.

2.1.15 | 7-Methoxy-2-oxo-2H-chromene-3-carboxylic acid (7h)

The yellow powder was obtained from ethyl 7-methoxy-2-oxo-2H-chromene-3-carboxylate (**2h**, 2 g, 8.06 mmol) by following the experimental conditions described for **7a** (1.54 g, 91% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.81 (s, 3H), 7.33 (dd, J = 8.8, 2.5 Hz, 1H), 7.40 (d, J = 8.9 Hz, 1H), 7.48 (d, J = 2.5 Hz, 1H), 8.69 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.3, 112.3, 117.7, 118.8, 119.0, 122.4, 148.5, 149.4, 156.2, 157.4, 164.4. HRMS (ESI): m/z calcd for $\text{C}_{11}\text{H}_8\text{O}_5$ $[\text{M}+\text{Na}]^+$: 243.1673, found: 243.1631.

2.1.16 | 6-Fluoro-2-oxo-2H-chromene-3-carboxylic acid (7i)

The white powder was obtained from ethyl 6-fluoro-2-oxo-2H-chromene-3-carboxylate (**2i**, 2 g, 7.92 mmol) by following the experimental conditions described for **7a** (1.62 g, 95% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.40 (d, J = 8.8 Hz, 1H), 7.86 (dd, J = 8.8, 2.5 Hz, 1H), 8.17 (d, J = 2.5 Hz, 1H), 8.69 (s, 1H), 13.37 (br-s, 1H). ^{13}C NMR (126 MHz, DMSO- d_6): δ = 115.4, 115.6, 118.6, 118.7, 119.3, 119.3, 119.9, 121.8, 122.0, 147.7, 147.8, 151.4, 151.4, 156.8, 157.5, 159.4, 164.3. HRMS (ESI): m/z calcd for $\text{C}_{10}\text{H}_5\text{FO}_4$ [$M+\text{Na}$] $^+$: 231.1328, found: 231.1318.

2.1.17 | 8-(Tert-butyl)-2-oxo-2H-chromene-3-carboxylic acid (7j)

The white powder was obtained from ethyl 8-(tert-butyl)-2-oxo-2H-chromene-3-carboxylate (**2j**, 2 g, 7.92 mmol) by following the experimental conditions described for **7a** (1.42 g, 83% yield): ^1H NMR (500 MHz, DMSO- d_6): δ = 1.46 (s, 9H), 7.35 (t, J = 8.0 Hz, 1H), 7.68 (d, J = 7.5 Hz, 1H), 7.78 (d, J = 8.0 Hz, 1H), 8.74 (s, 1H), 13.24 (br-s, 1H). ^{13}C NMR (126 MHz, DMSO- d_6): δ = 29.9, 35.0, 117.9, 119.0, 124.9, 129.2, 132.0, 137.0, 149.7, 153.6, 156.8, 164.4. HRMS (ESI): m/z calcd for $\text{C}_{14}\text{H}_{14}\text{O}_4$ [$M+\text{Na}$] $^+$: 269.2488, found: 269.2453.

2.1.18 | 7-Hydroxy-2-oxo-N-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (9a)

A solution of 7-hydroxy-2-oxo-2H-chromene-3-carbonyl chloride (**8a**, 0.2 g, 0.89 mmol) and sulfanilamide (**3a**, 0.17 g, 0.98 mmol) in the presence pyridine or triethylamine (1 mL) in DMF (5 mL) was stirred under reflux for 12 h. The solution was cooled, and 5 mL of 6 N HCl was added, and the resulting solid was filtered off and washed with water (10 mL) and air dried to yield a white powder (0.211 g, 66% yield). ^1H NMR (400 MHz, DMSO- d_6): δ = 6.84 (d, J = 2.4 Hz, 1H), 6.90 (dd, J = 8.6, 2.4 Hz, 1H), 7.30 (t, J = 9.1, 2H), 7.81–7.92 (m, 5H), 8.87 (d, J = 8.6 Hz, 1H), 10.89 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 102.4, 106.4, 111.5, 113.9, 115.2, 120.0, 127.3, 132.7, 139.5, 141.3, 149.1, 157.0, 161.2, 161.7, 165.0. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{12}\text{N}_2\text{O}_6\text{S}$ [$M+\text{Na}$] $^+$: 383.3285, found: 383.3253.

2.1.19 | 6-Chloro-2-oxo-N-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (9b)

The white powder was obtained from 6-chloro-2-oxo-2H-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.287 g, 92% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.32 (s, 2H), 7.61 (d, J = 8.9 Hz, 1H), 7.83 (d, J = 8.7 Hz, 2H), 7.85–7.95 (m, 3H), 7.89 (d, J = 8.9 Hz, 2H), 8.16

(d, J = 2.6 Hz, 1H), 8.78 (s, 1H), 10.86 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 118.8, 120.1, 120.2, 121.6, 127.3, 129.4, 129.6, 134.2, 139.9, 141.1, 146.7, 153.0, 160.2, 160.6. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{11}\text{ClN}_2\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 401.7737, found: 401.7717.

2.1.20 | 6-Bromo-2-oxo-N-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (9c)

The white powder was obtained from 6-bromo-2-oxo-2H-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.243 g, 83% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.32 (s, 2H), 7.61 (d, J = 8.8 Hz, 1H), 7.83 (dd, J = 8.8, 2.1 Hz, 2H), 7.85 (s, 1H), 7.89 (d, J = 8.5 Hz, 2H), 8.15 (d, J = 2.5 Hz, 1H), 8.87 (s, 1H), 10.86 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 117.2, 119.0, 120.1, 120.7, 121.6, 127.3, 132.6, 137.0, 139.9, 141.1, 146.6, 153.4, 160.1, 160.6. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{11}\text{BrN}_2\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 446.2247, found: 446.2236.

2.1.21 | 6,8-Dichloro-2-oxo-N-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (9d)

The white powder was obtained from 6,8-dichloro-2-oxo-2H-chromene-3-carbonyl chloride (**8d**, 0.2 g, 0.72 mmol) by following the experimental conditions described for **9a** (0.214 g, 72% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.33 (s, 2H), 7.86 (d, J = 8.8 Hz, 2H), 7.89 (d, J = 8.8 Hz, 2H), 8.14 (s, 2H), 8.84 (s, 1H), 10.78 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 120.2, 121.2, 121.4, 122.7, 127.3, 128.7, 129.3, 133.5, 140.0, 141.0, 148.7, 159.1, 160.4, 162.7. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{10}\text{Cl}_2\text{N}_2\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 436.2178, found: 436.2172.

2.1.22 | 6-Methoxy-2-oxo-N-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (9f)

The yellow powder was obtained from 2-oxo-2H-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.298 g, 95% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.84 (s, 1H), 7.32 (s, 2H), 7.39 (d, J = 8.2 Hz, 1H), 7.49 (d, J = 8.9 Hz, 1H), 7.59 (d, J = 2.2 Hz, 1H), 7.85 (d, J = 8.6 Hz, 2H), 7.90 (d, J = 8.6 Hz, 2H), 8.89 (s, 1H), 10.95 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.3, 112.3, 117.9, 119.3, 120.1, 120.2, 122.6, 127.3, 139.8, 141.1, 148.1, 148.9, 156.5, 160.8, 160.9. HRMS (ESI): m/z calcd for $\text{C}_{17}\text{H}_{14}\text{N}_2\text{O}_6\text{S}$ [$M+\text{Na}$] $^+$: 397.3553, found: 397.3518.

2.1.23 | 7-(Diethylamino)-2-oxo-N-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (9g)

To a solution of 7-(diethylamino)-2-oxo-2H-chromene-3-carboxylic acid (**7g**, 0.2 g, 0.77 mmol), 1-ethyl-3-(3-dimethylaminopropyl)

carbodiimide hydrochloride (EDC) (0.14 g, 0.92 mmol), 1-hydroxybenzotriazole hydrate (HOBt•H₂O) (0.124 g, 0.92 mmol) and triethylamine (TEA) (0.5 mL) in DMF (7 mL) was added sulfanilamide (**3a**, 0.132 g, 0.77 mmol). The solution was stirred at room temperature for 12 h. The water (5 mL) was added to the solution with few drops of 6 N HCl and the resulting solid was filtered off and washed with water and air-dried to yield a yellow powder (0.233 g, 73% yield). ¹H NMR (400 MHz, DMSO-*d*₆): δ = 1.15 (t, *J* = 7.1 Hz, 6H), 3.52 (q, *J* = 9.1 Hz, 4H), 6.68 (d, *J* = 2.1 Hz, 1H), 6.86 (dd, *J* = 8.9, 2.1 Hz, 1H), 7.29 (s, 2H), 7.73 (d, *J* = 9 Hz, 1H), 7.82 (d, *J* = 8.6 Hz, 2H), 7.88 (d, *J* = 8.6 Hz, 2H), 8.77 (s, 1H), 10.98 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 12.8, 44.9, 96.4, 108.4, 109.1, 111.0, 119.8, 127.3, 132.4, 139.3, 141.5, 148.9, 153.4, 157.9, 161.6, 162.6. HRMS (ESI): *m/z* calcd for C₂₀H₂₁N₃O₅S [M+Na]⁺: 438.4500, found: 438.4498.

2.1.24 | 7-Methoxy-2-oxo-*N*-(4-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (**9h**)

The yellow powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8h**, 0.2 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.283 g, 91% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 3.93 (s, 1H), 7.09 (d, *J* = 2.2 Hz, 1H), 7.20 (d, *J* = 8.2 Hz, 1H), 7.31 (s, 2H), 7.84 (d, *J* = 8.2 Hz, 2H), 7.92 (d, *J* = 2.6 Hz, 2H), 7.98 (d, *J* = 8.6 Hz, 1H), 8.93 (s, 1H), 10.89 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 56.8, 100.9, 112.6, 114.4, 115.7, 120.1, 127.3, 132.2, 139.7, 141.3, 148.9, 156.8, 161.1, 161.4, 165.3. HRMS (ESI): *m/z* calcd for C₁₇H₁₄N₂O₆S [M+Na]⁺: 397.3553, found: 397.3537.

2.1.25 | 6-Fluoro-2-oxo-*N*-(4-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (**9i**)

The white powder was obtained from 6-fluoro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8i**, 0.2 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.294 g, 94% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.32 (s, 2H), 7.64–7.69 (m, 2H), 7.85 (d, *J* = 8.7 Hz, 2H), 7.90–7.93 (m, 3H), 8.89 (s, 1H), 10.89 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 115.5, 115.8, 118.9, 119.8, 120.1, 121.5, 122.1, 127.3, 139.9, 141.1, 147.1, 150.8, 159.9, 160.4, 160.7. HRMS (ESI): *m/z* calcd for C₁₆H₁₁ClN₂O₅S [M+Na]⁺: 385.3238, found: 385.3213.

2.1.26 | 8-(*tert*-butyl)-2-oxo-*N*-(4-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (**9j**)

The white powder was obtained from 8-(*tert*-butyl)-2-oxo-2*H*-chromene-3-carbonyl chloride (**8j**, 0.2 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.196 g, 71% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 1.49 (s, 9H), 7.32 (s, 2H), 7.42 (t, *J* = 8.7 Hz, 1H), 7.73 (dd, *J* = 7.6, 2.1 Hz, 1H), 7.83–7.92 (m, 5H),

8.89 (s, 1H), 10.91 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 29.9, 35.0, 119.39, 119.47, 120.1, 125.4, 127.3, 129.3, 132.2, 137.2, 139.8, 141.3, 148.9, 153.0, 160.2, 160.9. HRMS (ESI): *m/z* calcd for C₂₀H₂₀N₂O₅S [M+Na]⁺: 423.4398, found: 423.4351.

2.1.27 | *N*-(4-(*N*-(4,6-dimethylpyrimidin-2-yl)sulfamoyl)phenyl)-7-hydroxy-2-oxo-2*H*-chromene-3-carboxamide (**10a**)

The white powder was obtained from 7-hydroxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8a**, 0.2 g, 0.84 mmol) and sulfamethazine (**3e**, 0.233 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.21 g, 54% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 2.26 (s, 6H), 6.76 (s, 1H), 6.85 (d, *J* = 2.1 Hz, 1H), 6.92 (dd, *J* = 8.6, 2.1 Hz, 1H), 7.85–7.89 (m, 3H), 7.98 (d, *J* = 8.7 Hz, 2H), 8.86 (s, 1H), 10.89 (s, 1H), 11.18 (s, 1H), 11.66 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 23.3, 102.4, 111.6, 114.3, 115.0, 119.4, 129.8, 132.7, 135.9, 141.9, 149.1, 156.6, 156.9, 161.1, 161.6, 164.6. HRMS (ESI): *m/z* calcd for C₂₂H₁₈N₄O₆S [M+Na]⁺: 489.4538, found: 489.4517.

2.1.28 | 6-Chloro-*N*-(4-(*N*-(4,6-dimethylpyrimidin-2-yl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (**10b**)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and sulfamethazine (**3e**, 0.228 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.254 g, 63% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 2.26 (s, 6H), 6.75 (s, 1H), 7.60 (d, *J* = 8.8 Hz, 1H), 7.79 (dd, *J* = 8.6, 2.5 Hz, 1H), 7.89 (d, *J* = 8.8 Hz, 2H), 8.00 (d, *J* = 8.8 Hz, 2H), 8.14 (d, *J* = 2.5 Hz, 1H), 8.84 (s, 1H), 10.85 (s, 1H), 11.81 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 23.3, 112.5, 118.7, 119.4, 119.8, 120.1, 121.6, 129.4, 129.6, 129.8, 130.3, 134.2, 136.3, 141.7, 146.7, 150.2, 152.9, 156.6, 160.1, 160.6. HRMS (ESI): *m/z* calcd for C₂₂H₁₇ClN₄O₅S [M+Na]⁺: 507.8985, found: 507.8975.

2.1.29 | 6-Bromo-*N*-(4-(*N*-(4,6-dimethylpyrimidin-2-yl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (**10c**)

The white powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and sulfamethazine (**3e**, 0.194 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.269 g, 72% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 2.26 (s, 6H), 6.76 (s, 1H), 7.54 (d, *J* = 8.8 Hz, 1H), 7.89 (d, *J* = 8.8 Hz, 2H), 7.94 (dd, *J* = 8.8, 2.3 Hz, 1H), 8.00 (d, *J* = 8.8 Hz, 2H), 8.27 (d, *J* = 2.3 Hz, 1H), 8.84 (s, 1H), 10.85 (s, 1H), 11.68 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 23.3, 112.5, 117.2, 119.0, 119.5,

120.6, 121.6, 129.8, 129.9, 132.6, 137.0, 141.7, 146.6, 153.4, 156.6, 160.1, 160.6 ppm. HRMS (ESI): m/z calcd for $C_{22}H_{17}BrN_4O_5S$ $[M+Na]^+$: 552.3499, found: 552.3443.

2.1.30 | *N*-(4-(*N*-(4,6-dimethylpyrimidin-2-yl)sulfamoyl)phenyl)-6-methoxy-2-oxo-2*H*-chromene-3-carboxamide (10f)

The white powder was obtained from 6-methoxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and sulfamethazine (**3e**, 0.234 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.367 g, 91% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 2.27 (s, 6H), 3.84 (s, 3H), 6.77 (s, 1H), 7.39 (dd, J = 9.0, 2.9 Hz, 1H), 7.50 (d, J = 9.0 Hz, 1H), 7.59 (d, J = 2.9 Hz, 1H), 7.90 (d, J = 8.8 Hz, 2H), 8.02 (d, J = 8.8 Hz, 2H), 8.87 (s, 1H), 10.96 (s, 1H), 11.59 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 23.3, 56.3, 112.3, 113.8, 117.8, 119.3, 119.4, 120.1, 122.8, 123.4, 136.2, 141.8, 148.1, 148.9, 156.5, 160.8, 160.9. HRMS (ESI): m/z calcd for $C_{23}H_{20}N_4O_6S$ $[M+Na]^+$: 503.4807, found: 503.4802.

2.1.31 | 7-Hydroxy-*N*-(4-(*N*-(6-methoxy-2-oxo-2*H*-chromene-3-carboxamido)phenyl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (11a)

The white powder was obtained from 7-hydroxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8a**, 0.2 g, 0.84 mmol) and sulfamethoxy-2-oxo-2*H*-chromene-3-carboxamide (**3f**, 0.235 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.237 g, 60% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 3.84 (s, 3H), 6.85 (d, J = 2.0 Hz, 1H), 6.91 (dd, J = 8.6, 2.0 Hz, 1H), 7.38 (d, J = 8.5 Hz, 1H), 7.76 (br-s, 1H), 7.82–7.88 (m, 6H), 8.87 (s, 1H), 10.87 (s, 1H), 11.22 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 54.9, 102.3, 111.1, 111.6, 114.2, 115.1, 120.2, 127.9, 132.7, 141.5, 149.1, 153.6, 156.9, 161.1, 161.6, 164.7. HRMS (ESI): m/z calcd for $C_{21}H_{16}N_4O_7S$ $[M+Na]^+$: 491.4265, found: 491.4241.

2.1.32 | 6-Chloro-*N*-(4-(*N*-(6-methoxy-2-oxo-2*H*-chromene-3-carboxamido)phenyl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (11b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and sulfamethoxy-2-oxo-2*H*-chromene-3-carboxamide (**3f**, 0.228 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.326 g, 82% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 3.84 (s, 3H), 7.39 (br-s, 1H), 7.62 (d, J = 8.7 Hz, 1H), 7.82–7.91 (m, 5H), 8.15 (d, J = 2.2 Hz, 1H), 8.85 (s, 1H), 10.84 (s, 1H), 13.84 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 54.9, 118.8, 120.2, 120.7, 121.6, 127.8, 129.4, 129.7, 134.2, 141.4, 146.1, 146.7, 153.0, 160.1, 160.6. HRMS (ESI): m/z calcd for $C_{21}H_{15}ClN_4O_6S$ $[M+Na]^+$: 509.8711, found: 509.8709.

2.1.33 | 6-Bromo-*N*-(4-(*N*-(6-methoxy-2-oxo-2*H*-chromene-3-carboxamido)phenyl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (11c)

The white powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and sulfamethoxy-2-oxo-2*H*-chromene-3-carboxamide (**3f**, 0.196 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.309 g, 83% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 3.84 (s, 3H), 7.40 (br-s, 1H), 7.55 (d, J = 8.8 Hz, 1H), 7.84–7.94 (m, 4H), 7.92 (dd, J = 8.8, 2.4 Hz, 2H), 8.28 (d, J = 2.4 Hz, 1H), 8.84 (s, 1H), 10.84 (s, 1H), 13.81 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 54.9, 117.2, 119.0, 120.2, 120.7, 121.6, 127.8, 132.6, 137.0, 141.3, 146.6, 153.4, 160.1, 160.6 ppm. HRMS (ESI): m/z calcd for $C_{21}H_{15}BrN_4O_6S$ $[M+Na]^+$: 554.3226, found: 554.3213.

2.1.34 | 6,8-Dichloro-*N*-(4-(*N*-(6-methoxy-2-oxo-2*H*-chromene-3-carboxamido)phenyl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (11d)

The white powder was obtained from 6,8-dichloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8d**, 0.2 g, 0.72 mmol) and sulfamethoxy-2-oxo-2*H*-chromene-3-carboxamide (**3f**, 0.202 g, 0.72 mmol) by following the experimental conditions described for **9a** (0.263 g, 70% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 3.85 (s, 3H), 7.40 (br-s, 1H), 7.87 (m, 5H), 8.13 (s, 2H), 8.82 (s, 1H), 10.77 (s, 1H), 13.83 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 54.9, 120.2, 121.2, 121.4, 122.7, 127.9, 128.2, 128.6, 129.3, 133.4, 141.3, 146.1, 148.7, 151.4, 153.9, 159.1, 160.4. HRMS (ESI): m/z calcd for $C_{21}H_{14}Cl_2N_4O_6S$ $[M+Na]^+$: 544.3155, found: 544.3108.

2.1.35 | 6-Methoxy-*N*-(4-(*N*-(6-methoxy-2-oxo-2*H*-chromene-3-carboxamido)phenyl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (11f)

The white powder was obtained from 6-methoxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and sulfamethoxy-2-oxo-2*H*-chromene-3-carboxamide (**3f**, 0.235 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.328 g, 81% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 3.83 (d, J = 4.0 Hz, 6H), 7.37 (dd, J = 6.4, 2.1 Hz, 2H), 7.47 (d, J = 8.8 Hz, 1H), 7.56 (d, J = 4.0 Hz, 1H), 7.84–7.89 (m, 4H), 7.89 (s, 1H), 8.88 (s, 1H), 10.93 (s, 1H), 13.93 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 54.9, 56.3, 107.2, 112.3, 117.8, 119.3, 120.1, 122.8, 127.9, 129.8, 130.1, 135.6, 141.4, 148.0, 148.9, 156.5, 160.8. HRMS (ESI): m/z calcd for $C_{22}H_{18}N_4O_7S$ $[M+Na]^+$: 505.4531, found: 505.4522.

2.1.36 | 6-Chloro-2-oxo-*N*-(4-(*N*-(pyridin-2-yl)sulfamoyl)phenyl)-2*H*-chromene-3-carboxamide (12b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and sulfapyridine (**3d**,

0.204 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.298 g, 80% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 6.88 (t, J = 7.5 Hz, 1H), 7.17 (d, J = 8.6 Hz, 1H), 7.61 (d, J = 8.6 Hz, 1H), 7.72 (t, J = 8.5 Hz, 1H), 7.83–7.89 (m, 6H), 8.02 (br-s, 1H), 8.15 (s, 1H), 8.84 (s, 1H), 10.84 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 101.9, 113.1, 119.0, 119.7, 120.4, 120.8, 121.6, 128.2, 132.6, 137.7, 141.2, 149.6, 153.4, 160.1, 160.9. HRMS (ESI): m/z calcd for $\text{C}_{21}\text{H}_{14}\text{ClN}_3\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 478.8573, found: 478.8539.

2.1.37 | 6-Bromo-2-oxo-*N*-(4-(*N*-(pyridin-2-yl)sulfamoyl)phenyl)-2*H*-chromene-3-carboxamide (**12c**)

The white powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and sulfapyridine (**3d**, 0.18 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.288 g, 82% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 6.88 (t, J = 7.5 Hz, 1H), 7.17 (d, J = 8.6 Hz, 1H), 7.54 (d, J = 8.6 Hz, 1H), 7.72 (t, J = 7.5 Hz, 1H), 7.85–7.94 (m, 6H), 8.02 (br-s, 1H), 8.28 (d, J = 2.4 Hz, 1H), 8.84 (s, 1H), 10.3 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 101.6, 112.8, 117.2, 119.0, 120.1, 120.7, 121.6, 128.3, 132.6, 137.0, 141.5, 146.5, 153.4, 160.0, 160.6. HRMS (ESI): m/z calcd for $\text{C}_{21}\text{H}_{14}\text{BrN}_3\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 523.3089, found: 523.3059.

2.1.38 | 6,8-Dichloro-2-oxo-*N*-(4-(*N*-(pyridin-2-yl)sulfamoyl)phenyl)-2*H*-chromene-3-carboxamide (**12d**)

The white powder was obtained from 6,8-dichloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8d**, 0.2 g, 0.72 mmol) and sulfapyridine (**3d**, 0.18 g, 0.72 mmol) by following the experimental conditions described for **9a** (0.210 g, 59% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 6.88 (t, J = 6.6 Hz, 1H), 7.17 (d, J = 8.4 Hz, 1H), 7.73 (dt, J = 7.1, 1.9 Hz, 1H), 7.84–7.91 (m, 4H), 8.02 (d, J = 2.4 Hz, 1H), 8.12 (s, 2H), 8.81 (s, 1H), 10.76 (s, 1H), 12.02 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 114.1, 120.1, 121.2, 121.4, 122.7, 128.3, 128.6, 129.3, 133.4, 137.6, 140.9, 141.4, 146.1, 148.7, 153.5, 159.0, 160.4. HRMS (ESI): m/z calcd for $\text{C}_{21}\text{H}_{13}\text{Cl}_2\text{N}_3\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 513.3020, found: 513.3007.

2.1.39 | 2-Oxo-*N*-(4-(*N*-(pyridin-2-yl)sulfamoyl)phenyl)-2*H*-chromene-3-carboxamide (**12e**)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8e**, 0.2 g, 0.96 mmol) and sulfapyridine (**3d**, 0.24 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.389 g, 96% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 6.87 (t, J = 6.2 Hz, 1H), 7.17 (d, J = 8.6 Hz, 1H), 7.47 (t, J = 7.4 Hz, 1H), 7.56 (d, J = 8.4 Hz, 1H), 7.78 (dt, J = 6.3 Hz, 1H), 7.84–7.90 (q, J = 6.2, 4H), 8.02 (d, J = 7.0 Hz, 2H), 8.90 (s, 1H), 10.88 (s, 1H), 11.94 (br, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 114.1,

116.0, 116.7, 118.8, 120.0, 120.3, 120.4, 125.7, 128.3, 130.8, 134.9, 137.3, 140.7, 141.6, 148.1, 154.4, 160.6, 160.9. HRMS (ESI): m/z calcd for $\text{C}_{21}\text{H}_{15}\text{N}_3\text{O}_5\text{S}$ [$M+\text{Na}$] $^+$: 444.4133, found: 444.4103.

2.1.40 | 2-Oxo-*N*-(4-(*N*-(pyridin-2-yl)sulfamoyl)phenyl)-2*H*-chromene-3-carboxamide (**12f**)

The white powder was obtained from 6-methoxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and sulfapyridine (**3d**, 0.21 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.249 g, 66% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.84 (s, 3H), 6.88 (t, J = 6.1 Hz, 1H), 7.18 (d, J = 8.6 Hz, 1H), 7.38 (dd, J = 7.4, 2.0 Hz, 1H), 7.50 (d, J = 7.6 Hz, 1H), 7.59 (d, J = 2.1 Hz, 1H), 7.73 (dt, J = 6.6, 1.9 Hz, 1H), 7.86–7.91 (q, J = 7.3 Hz, 4H), 8.02 (d, J = 7.0 Hz, 2H), 8.87 (s, 1H), 10.93 (s, 1H), 11.93 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.3, 112.3, 114.1, 117.8, 119.3, 120.1, 120.2, 122.8, 128.4, 140.8, 141.5, 148.1, 148.9, 156.5, 160.8. HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{17}\text{N}_3\text{O}_6\text{S}$ [$M+\text{Na}$] $^+$: 474.4393, found: 474.4329.

2.1.41 | 6-Chloro-*N*-(4-(*N*-(5-methyl-1,3,4-thiadiazol-2-yl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (**13b**)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and sulfamethizole (**3g**, 0.222 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.288 g, 74% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 2.47 (s, 3H), 7.62 (d, J = 8.8 Hz, 1H), 7.80 (d, J = 8.8 Hz, 2H), 7.88 (d, J = 8.8 Hz, 3H), 8.15 (d, J = 2.4 Hz, 1H), 8.84 (s, 1H), 10.86 (s, 1H), 13.94 (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 16.5, 118.8, 120.0, 120.3, 121.6, 127.5, 129.4, 129.6, 134.2, 137.5, 141.6, 146.7, 153.0, 155.1, 160.1, 160.6, 168.3. HRMS (ESI): m/z calcd for $\text{C}_{19}\text{H}_{13}\text{ClN}_4\text{O}_5\text{S}_2$ [$M+\text{Na}$] $^+$: 499.8982, found: 499.8936.

2.1.42 | 6-Bromo-*N*-(4-(*N*-(5-methyl-1,3,4-thiadiazol-2-yl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (**13c**)

The white powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and sulfamethizole (**3g**, 0.189 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.277 g, 76% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 2.47 (s, 3H), 7.55 (d, J = 8.8 Hz, 1H), 7.82 (d, J = 8.8 Hz, 2H), 7.90 (d, J = 7.8 Hz, 2H), 7.94 (dd, J = 7.4, 2.1 Hz, 1H), 8.28 (d, J = 2.4 Hz, 1H), 8.84 (s, 1H), 10.86 (s, 1H), 13.94 ppm (br-s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 16.5, 117.2, 119.0, 120.3, 120.7, 121.6, 127.5, 132.6, 137.0, 137.5, 141.7, 146.6, 153.4, 155.0, 160.1, 160.6, 168.3. HRMS (ESI): m/z calcd for $\text{C}_{19}\text{H}_{13}\text{BrN}_4\text{O}_5\text{S}_2$ [$M+\text{Na}$] $^+$: 544.3507, found: 544.3501.

2.1.43 | 6-Chloro-*N*-(4-(*N*-(2,6-dimethoxypyrimidin-4-yl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (14b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and sulfadimethoxine (**3h**, 0.254 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.312 g, 73% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 3.77 (s, 3H), 3.80 (s, 3H), 5.96 (s, 1H), 7.61 (d, J = 8.8 Hz, 1H), 7.81 (dd, J = 8.8, 2.4 Hz, 1H), 7.92–7.97 (q, J = 7.6, 4H), 8.14 (d, J = 2.4 Hz, 1H), 8.85 (s, 1H), 10.90 (s, 1H), 11.59 (br-s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 54.2, 55.0, 85.0, 118.8, 120.1, 120.2, 121.6, 129.1, 129.2, 129.6, 134.2, 135.2, 142.4, 146.7, 153.0, 160.0, 160.3, 160.8, 164.6, 172.1. HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{17}\text{ClN}_4\text{O}_7\text{S}$ [$M+\text{Na}$] $^+$: 539.8971, found: 539.8957.

2.1.44 | 6-Bromo-*N*-(4-(*N*-(2,6-dimethoxypyrimidin-4-yl) sulfamoyl)phenyl) -2-oxo-2*H*-chromene-3-carboxamide (14c)

The white compound was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and sulfadimethoxine (**3h**, 0.217 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.322 g, 82% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 3.77 (s, 3H), 3.80 (s, 3H), 5.96 (s, 1H), 7.54 (d, J = 8.8 Hz, 1H), 7.91–7.98 (m, 5H), 8.15 (d, J = 2.4 Hz, 1H), 8.84 (s, 1H), 10.90 (s, 1H), 11.56 (br-s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 54.2, 55.0, 85.0, 117.2, 119.0, 120.2, 120.6, 121.6, 129.1, 132.6, 137.0, 142.4, 146.6, 153.4, 160.0, 160.3, 160.8, 162.7, 172.1. HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{17}\text{BrN}_4\text{O}_7\text{S}$ [$M+\text{Na}$] $^+$: 584.3486, found: 584.3497.

2.1.45 | *N*-(4-(*N*-(2,6-dimethoxypyrimidin-4-yl) sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (14e)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8e**, 0.2 g, 0.96 mmol) and sulfadimethoxine (**3h**, 0.298 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.377 g, 81% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 3.77 (s, 3H), 3.80 (s, 3H), 5.96 (s, 1H), 7.47 (t, J = 7.6 Hz, 1H), 7.56 (d, J = 8.4 Hz, 1H), 7.56 (t, J = 8.6 Hz, 1H), 7.92–8.01 (m, 5H), 8.90 (s, 1H), 10.94 (s, 1H), 11.57 (br-s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 54.2, 55.0, 85.0, 116.7, 118.8, 120.2, 120.2, 125.7, 129.1, 130.8, 134.9, 135.1, 142.5, 148.1, 154.4, 160.3, 160.5, 151.1, 164.7, 172.1. HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{18}\text{N}_4\text{O}_7\text{S}$ [$M+\text{Na}$] $^+$: 505.4531, found: 505.4509.

2.1.46 | *N*-(4-(*N*-acetylsulfamoyl)phenyl)-6-chloro-2-oxo-2*H*-chromene-3-carboxamide (15b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and sulfacetamide

(**3c**, 0.176 g, 0.82 mmol) by following the experimental conditions described for **9a** (0.302 g, 88% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 1.93 (s, 3H), 7.61 (d, J = 8.8 Hz, 1H), 7.81 (dd, J = 8.8, 2.5 Hz, 1H), 7.91–7.96 (m, 4H), 8.14 (d, J = 2.5 Hz, 1H), 8.86 (s, 1H), 10.93 (s, 1H), 12.04 (br-s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 23.6, 116.6, 118.8, 120.1, 120.6, 121.7, 129.5, 134.3, 134.6, 139.0, 142.7, 146.7, 153.0, 160.1, 160.6, 169.2. HRMS (ESI): m/z calcd for $\text{C}_{18}\text{H}_{13}\text{ClN}_2\text{O}_6\text{S}$ [$M+\text{Na}$] $^+$: 443.8101, found: 443.8095.

2.1.47 | *N*-(4-(*N*-acetylsulfamoyl)phenyl)-6-bromo-2-oxo-2*H*-chromene-3-carboxamide (15c)

The white powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and sulfacetamide (**3c**, 0.15 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.285 g, 87% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 1.93 (s, 3H), 7.54 (d, J = 8.8 Hz, 1H), 7.90–7.95 (m, 5H), 8.28 (d, J = 2.4 Hz, 1H), 8.85 (s, 1H), 10.93 (s, 1H), 12.05 (s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 23.6, 117.2, 119.1, 120.1, 120.6, 121.6, 129.4, 132.6, 134.6, 137.1, 142.7, 146.7, 153.4, 160.0, 160.8, 169.1. HRMS (ESI): m/z calcd for $\text{C}_{18}\text{H}_{13}\text{BrN}_2\text{O}_6\text{S}$ [$M+\text{Na}$] $^+$: 488.2614, found: 488.2601.

2.1.48 | *N*-(4-(*N*-acetylsulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (15e)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8e**, 0.2 g, 0.96 mmol) and sulfacetamide (**3c**, 0.21 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.329 g, 89% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 1.93 (s, 3H), 7.49 (t, J = 7.9 Hz, 1H), 7.58 (d, J = 8.8 Hz, 1H), 7.80 (t, J = 8.8 Hz, 1H), 7.91–7.97 (q, J = 7.6, 4H), 8.02 (dd, J = 6.9, 2.1 Hz, 1H), 8.93 (s, 1H), 10.97 (s, 1H), 12.05 (br-s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 23.7, 116.7, 118.8, 120.1, 120.3, 125.8, 129.4, 130.8, 134.6, 134.9, 142.8, 148.2, 154.4, 160.5, 161.1, 169.2. HRMS (ESI): m/z calcd for $\text{C}_{18}\text{H}_{14}\text{N}_2\text{O}_6\text{S}$ [$M+\text{Na}$] $^+$: 409.3660, found: 409.3641.

2.1.49 | *N*-(4-(*N*-acetylsulfamoyl)phenyl)-6-methoxy-2-oxo-2*H*-chromene-3-carboxamide (15f)

The white powder was obtained from 6-methoxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and sulfacetamide (**3c**, 0.18 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.297 g, 85% yield): $^1\text{H NMR}$ (400 MHz, DMSO- d_6): δ = 1.93 (s, 3H), 3.85 (s, 3H), 7.40 (dd, J = 9.0, 2.8 Hz, 1H), 7.50 (d, J = 9.0 Hz, 1H), 7.59 (dd, J = 2.8 Hz, 1H), 7.91–7.96 (m, 4H), 8.89 (s, 1H), 11.01 (s, 1H), 12.05 (br-s, 1H). $^{13}\text{C NMR}$ (100 MHz, DMSO- d_6): δ = 23.7, 56.3, 112.4, 117.9, 119.3, 120.1, 120.3, 122.8, 129.4, 134.5, 142.8, 148.1, 148.9, 156.5, 160.8, 161.1, 169.2. HRMS (ESI): m/z calcd for $\text{C}_{19}\text{H}_{16}\text{N}_2\text{O}_7\text{S}$ [$M+\text{Na}$] $^+$: 439.3920, found: 439.3901.

2.1.50 | 6-Chloro-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (16b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and 3-aminobenzenesulfonamide (**3i**, 0.141 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.273 g, 85% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.43 (s, 2H), 7.59–7.62 (m, 3H), 7.82 (dd, J = 8.8, 2.5 Hz, 1H), 7.87 (t, J = 7.8, 1H), 8.16 (d, J = 2.5 Hz, 1H), 8.28 (s, 1H), 8.85 (s, 1H), 10.83 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 117.4, 118.8, 120.2, 121.8, 123.3, 129.4, 129.5, 130.2, 134.2, 138.7, 145.3, 146.4, 153.0, 160.1, 160.6, 162.7. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{11}\text{ClN}_2\text{O}_5\text{S}$ [$\text{M}+\text{Na}$] $^+$: 401.7737, found: 401.7703.

2.1.51 | 6-Bromo-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (16c)

The white powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and 3-aminobenzenesulfonamide (**3i**, 0.121 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.267 g, 90% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.43 (s, 2H), 7.52–7.60 (m, 2H), 7.92 (dd, J = 8.8, 2.5 Hz, 3H), 8.29 (s, 2H), 8.84 (s, 1H), 10.82 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 117.2, 117.4, 119.0, 120.7, 121.8, 123.3, 130.2, 132.6, 136.6, 138.7, 145.3, 146.4, 153.0, 160.1, 160.5, 162.7. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{11}\text{BrN}_2\text{O}_5\text{S}$ [$\text{M}+\text{Na}$] $^+$: 446.2247, found: 446.2211.

2.1.52 | 2-Oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (16e)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8e**, 0.2 g, 0.96 mmol) and 3-aminobenzenesulfonamide (**3i**, 0.17 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.248 g, 75% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.43 (s, 2H), (t, J = 8.3 Hz, 1H), 7.56–7.61 (m, 3H), 7.80 (t, J = 8.3 Hz, 1H), 7.90 (d, J = 6.7 Hz, 1H), 8.03 (d, J = 7.7 Hz, 1H), 8.29 (s, 1H), 8.91 (s, 1H), 10.87 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 116.7, 117.3, 118.8, 120.5, 121.7, 123.3, 125.7, 130.2, 130.8, 134.8, 138.7, 145.3, 147.9, 154.3, 160.6, 160.8. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{12}\text{N}_2\text{O}_5\text{S}$ [$\text{M}+\text{Na}$] $^+$: 367.3292, found: 367.3224.

2.1.53 | 6-Methoxy-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (16f)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and 3-aminobenzenesulfonamide (**3i**, 0.145 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.211 g, 69% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.85 (s, 3H), 7.40 (dd, J = 6.8, 2.1 Hz, 1H), 7.43 (s, 2H), 7.50 (dd, J = 8.2, 2.1 Hz,

1H), 7.58–7.61 (m, 3H), 7.89 (dd, J = 8.7, 2.0 Hz, 1H), 8.30 (s, 1H), 8.88 (s, 1H), 10.91 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.3, 112.35, 117.3, 117.9, 119.3, 120.4, 121.7, 122.8, 123.3, 130.2, 138.7, 145.3, 147.8, 148.9, 156.5, 160.8. HRMS (ESI): m/z calcd for $\text{C}_{17}\text{H}_{14}\text{N}_2\text{O}_6\text{S}$ [$\text{M}+\text{Na}$] $^+$: 397.3553, found: 397.3516.

2.1.54 | 7-Methoxy-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (16h)

The white powder was obtained from 3-oxo-2*H*-chromene-3-carbonyl chloride (**8h**, 0.2 g, 0.84 mmol) and 3-aminobenzenesulfonamide (**3i**, 0.145 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.246 g, 84% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.93 (s, 3H), 7.10 (dd, J = 6.6, 2.0 Hz, 1H), 7.19 (d, J = 7.6 Hz, 1H), 7.42 (s, 2H), 7.58 (d, J = 7.4 Hz, 2H), 7.89 (d, J = 7.6 Hz, 1H), 7.97 (d, J = 7.6 Hz, 1H), 8.29 (s, 1H), 8.91 (s, 1H), 10.85 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.8, 100.92, 112.6, 114.3, 115.8, 117.3, 121.6, 123.3, 130.2, 132.2, 138.8, 145.3, 148.6, 156.8, 161.0, 161.3, 165.2. HRMS (ESI): m/z calcd for $\text{C}_{17}\text{H}_{14}\text{N}_2\text{O}_6\text{S}$ [$\text{M}+\text{Na}$] $^+$: 397.3553, found: 397.3529.

2.1.55 | 6-Fluoro-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (16i)

The white powder was obtained from 6-fluoro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8i**, 0.2 g, 0.82 mmol) and 3-aminobenzenesulfonamide (**3i**, 0.141 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.291 g, 93% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 7.43 (s, 2H), 7.59–7.68 (m, 4H), 7.88–6.93 (m, 2H), 8.29 (s, 1H), 8.87 (s, 1H), 10.86 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 115.7, 117.4, 118.9, 119.7, 121.8, 123.3, 130.2, 138.7, 145.3, 146.4, 150.8, 157.5, 159.9, 160.3, 160.6. HRMS (ESI): m/z calcd for $\text{C}_{16}\text{H}_{11}\text{FN}_2\text{O}_5\text{S}$ [$\text{M}+\text{Na}$] $^+$: 385.3238, found: 385.3224.

2.1.56 | *N*-(4-(*N*-(5,6-dimethoxypyrimidin-4-yl)sulfamoyl)phenyl)-7-hydroxy-2-oxo-2*H*-chromene-3-carboxamide (17a)

The white powder was obtained from 7-hydroxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8a**, 0.2 g, 0.96 mmol) and sulfadoxin (**3b**, 0.298 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.17 g, 41% yield): ^1H NMR (400 MHz, DMSO- d_6): δ = 3.71 (s, 3H), 3.91 (s, 3H), 6.85 (d, J = 7.8 Hz, 1H), 6.91 (dd, J = 7.6, 2.0 Hz, 1H), 7.85–7.92 (m, 3H), 8.00 (d, J = 7.8 Hz, 2H), 8.13 (s, 1H), 8.87 (s, 1H), 10.91 (s, 1H), 11.16 (br-s, 2H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 54.5, 60.7, 102.4, 111.6, 114.3, 115.1, 119.8, 127.8, 129.3, 132.8, 135.9, 150.8, 156.9, 161.3, 161.5, 162.1, 164.6. HRMS (ESI): m/z calcd for $\text{C}_{22}\text{H}_{18}\text{N}_4\text{O}_8\text{S}$ [$\text{M}+\text{Na}$] $^+$: 521.4571, found: 521.4541.

2.1.57 | *N*-(4-(*N*-(5,6-dimethoxypyrimidin-4-yl)sulfamoyl)phenyl)-2-oxo-2*H*-chromene-3-carboxamide (17e)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8e**, 0.2 g, 0.96 mmol) and sulfadoxin (**3b**, 0.298 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.299 g, 64% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 3.71 (s, 3H), 3.91 (s, 3H), 7.49 (t, *J* = 7.4 Hz, 1H), 7.57 (d, *J* = 8.4 Hz, 1H), 7.79 (t, *J* = 7.7 Hz, 1H), 7.93 (d, *J* = 8.7 Hz, 2H), 8.01 (d, *J* = 8.7 Hz, 3H), 8.13 (s, 1H), 8.91 (s, 1H), 10.93 (s, 1H), 11.11 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 54.5, 60.7, 116.7, 118.8, 119.9, 120.3, 125.8, 127.7, 129.3, 130.8, 134.9, 136.0, 142.2, 148.1, 150.7, 151.1, 154.4, 160.6, 161.1, 162.1. HRMS (ESI): *m/z* calcd for C₂₂H₁₈N₄O₇S [M+Na]⁺: 505.4531, found: 505.4510.

2.1.58 | *N*-(2,4-disulfamoyl-5-(trifluoromethyl)phenyl)-6-methoxy-2-oxo-2*H*-chromene-3-carboxamide (18f)

The yellow powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and 4-amino-6-(trifluoromethyl)benzene-1,3-disulfonamide (**3j**, 0.268 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.217 g, 49% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 3.85 (s, 3H), 7.43 (dd, *J* = 8.9, 2.6 Hz, 1H), 7.55 (d, *J* = 9.0 Hz, 1H), 7.64 (d, *J* = 2.6 Hz, 1H), 7.89 (m, 4H), 8.66 (s, 1H), 8.86 (s, 1H), 9.06 (s, 1H), 11.54 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 56.3, 117.1, 118.7, 119.2, 120.4, 121.6, 123.1, 125.9, 130.7, 131.2, 135.1, 139.9, 143.4, 147.1, 156.3, 160.2, 160.9. HRMS (ESI): *m/z* calcd for C₁₈H₁₄F₃N₃O₈S₂ [M+Na]⁺: 544.4306, found: 544.4302.

2.1.59 | 7-Hydroxy-2-oxo-*N*-(4-(*N*-(pyrimidin-2-yl)sulfamoyl)phenyl)-2*H*-chromene-3-carboxamide (19a)

The white powder was obtained from 7-hydroxy-2-oxo-2*H*-chromene-3-carbonyl chloride (**8a**, 0.2 g, 0.84 mmol) and Sulfadiazine (**3h**, 0.233 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.19 g, 47% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 6.85 (d, *J* = 2.5 Hz, 1H), 6.93 (dd, *J* = 7.6, 2.1 Hz, 1H), 7.05 (t, *J* = 8.6 Hz, 1H), 7.86–7.91 (m, 3H), 7.98 (d, *J* = 8.7 Hz, 2H), 8.52 (d, *J* = 7.6 Hz, 2H), 8.87 (s, 1H), 10.91 (s, 1H), 11.20 (br-s, 1H), 11.74 (s, 1H). HRMS (ESI): *m/z* calcd for C₂₀H₁₄N₄O₆S [M+Na]⁺: 461.4051, found: 461.4019.

2.1.60 | 6-Chloro-2-oxo-*N*-(4-sulfamoylphenethyl)-2*H*-chromene-3-carboxamide (20b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and 4-(2-aminoethyl)

benzenesulfonamide (**3h**, 0.17 g, 0.94 mmol) by following the experimental conditions described for **9a** (0.289 g, 94% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 2.94 (t, *J* = 7.6 Hz, 2H), 3.58–3.63 (q, *J* = 7.7 Hz, 2H), 7.30 (s, 2H), 7.47 (d, *J* = 8.6 Hz, 2H), 7.54 (d, *J* = 7.8 Hz, 1H), 7.75–7.78 (dd, *J* = 7.6, 2.1 Hz, 3H), 8.12 (d, *J* = 2.4 Hz, 1H), 8.76 (t, *J* = 7.6 Hz, 1H), 8.81 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 35.11, 118.2, 120.3, 120.4, 129.2, 129.5, 129.6, 133.9, 142.6, 143.8, 146.7, 152.9, 160.3, 161.2. HRMS (ESI): *m/z* calcd for C₁₈H₁₅ClN₂O₅S [M+Na]⁺: 429.8312, found: 429.8303.

2.1.61 | 6-Methoxy-2-oxo-*N*-(4-sulfamoylphenethyl)-2*H*-chromene-3-carboxamide (20f)

The yellow powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and 4-(2-aminoethyl)benzenesulfonamide (**3h**, 0.17 g, 0.94 mmol) by following the experimental conditions described for **9a** (0.212 g, 76% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 2.94 (t, *J* = 7.6 Hz, 2H), 3.58–3.63 (q, *J* = 7.6 Hz, 2H), 3.38 (s, 3H), 7.30 (s, 2H), 7.36 (dd, *J* = 7.6, 2.1 Hz, 1H), 7.44–7.47 (m, 3H), 7.57 (d, *J* = 2.4 Hz, 1H), 7.77 (t, *J* = 7.6 Hz, 1H), 8.83 (s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 35.11, 118.2, 120.3, 120.4, 129.2, 129.5, 129.6, 133.9, 142.6, 143.8, 146.7, 152.9, 160.3, 161.2. HRMS (ESI): *m/z* calcd for C₁₉H₁₈N₂O₆S [M+Na]⁺: 425.4126, found: 425.4112.

2.1.62 | 6-Chloro-2-oxo-*N*-(2-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (21b)

The white powder was obtained from 6-chloro-2-oxo-2*H*-chromene-3-carbonyl chloride (**8b**, 0.2 g, 0.82 mmol) and 2-aminobenzenesulfonamide (**3j**, 0.141 g, 0.84 mmol) by following the experimental conditions described for **9a** (0.255 g, 76% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.37 (t, *J* = 6.8 Hz, 1H), 7.52 (s, 2H), 7.60–7.68 (m, 2H), 7.82 (dd, *J* = 8.6, 2.2 Hz, 1H), 7.89 (d, *J* = 7.6 Hz, 1H), 8.18 (d, *J* = 2.1 Hz, 1H), 8.21 (s, 1H), 9.00 (s, 1H), 11.10 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 118.7, 120.3, 120.8, 125.2, 125.6, 127.8, 129.4, 129.8, 132.9, 134.4, 134.7, 147.4, 153.1, 160.1, 160.3. HRMS (ESI): *m/z* calcd for C₁₆H₁₁ClN₂O₅S [M+Na]⁺: 401.7737, found: 401.7716.

2.1.63 | 6-Bromo-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (21c)

The yellow powder was obtained from 6-bromo-2-oxo-2*H*-chromene-3-carbonyl chloride (**8c**, 0.2 g, 0.70 mmol) and 2-aminobenzenesulfonamide (**3j**, 0.121 g, 0.70 mmol) by following the experimental conditions described for **9a** (0.219 g, 79% yield): ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.37 (t, *J* = 6.8 Hz, 1H), 7.52 (s, 2H), 7.56 (d, *J* = 2.1 Hz, 1H), 7.66 (t, *J* = 6.4 Hz, 1H), 7.89–7.95 (m, 2H), 8.16 (d, *J* = 7.6 Hz, 1H), 8.35 (d, *J* = 2.4 Hz, 1H), 9.00 (s, 1H), 11.10 (s, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 117.2, 118.9, 120.7, 120.8, 125.2, 125.6, 127.8, 132.8, 132.9,

134.6, 134.7, 137.1, 147.3, 153.5, 160.1, 160.3. HRMS (ESI): m/z calcd for $C_{16}H_{11}BrN_2O_5S$ $[M+Na]^+$: 446.2247, found: 446.2209.

2.1.64 | 2-Oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (21e)

The white powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8e**, 0.2 g, 0.96 mmol) and 2-aminobenzenesulfonamide (**3j**, 0.17 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.265 g, 81% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 7.36 (t, J = 6.8, Hz, 1H), 7.46–7.58 (m, 4H), 7.67 (t, J = 6.7, Hz, 1H), 7.78 (dd, J = 7.6, 2.1 Hz, 1H), 7.92 (d, J = 7.6, 1H), 8.07 (d, J = 2.1 Hz, 1H), 8.19 (d, J = 2.4 Hz, 1H), 9.03 (s, 1H), 11.14 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 116.6, 118.9, 119.6, 125.1, 125.6, 125.7, 127.8, 131.0, 134.6, 134.7, 135.0, 148.7, 154.5, 160.6, 160.7. HRMS (ESI): m/z calcd for $C_{16}H_{12}N_2O_5S$ $[M+Na]^+$: 367.3292, found: 367.3244.

2.1.65 | 6-Methoxy-2-oxo-*N*-(3-sulfamoylphenyl)-2*H*-chromene-3-carboxamide (21f)

The yellow powder was obtained from 2-oxo-2*H*-chromene-3-carbonyl chloride (**8f**, 0.2 g, 0.84 mmol) and 2-aminobenzenesulfonamide (**3j**, 0.145 g, 0.96 mmol) by following the experimental conditions described for **9a** (0.247 g, 74% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 3.85 (s, 3H), 7.34–7.41 (m, 2H), 7.51 (d, J = 6.4 Hz, 3H), 7.64–7.68 (m, 2H), 7.90 (dd, J = 7.6, 2.1 Hz, 1H), 8.17 (d, J = 7.4 Hz, 1H), 8.99 (s, 1H), 11.17 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 56.3, 112.5, 117.8, 119.4, 119.8, 122.8, 125.1, 125.7, 127.8, 132.9, 134.7, 148.5, 149.0, 156.5, 160.7. HRMS (ESI): m/z calcd for $C_{17}H_{14}N_2O_6S$ $[M+Na]^+$: 397.3553, found: 397.3539.

2.1.66 | 2-Oxo-2-((4-sulfamoylphenyl)amino)ethyl-7-(diethylamino)-2-oxo-2*H*-chromene-3-carboxylate (22)

To a solution of ethyl 7-(diethylamino)-2-oxo-2*H*-chromene-3-carboxylate (**2g**, 0.2 g, 0.77 mmol) in dry DMF (15 mL), anhydrous K_2CO_3 (0.18 g, 1.28 mmol) was added. The mixture was stirred for 15 min at 70–80°C and 2-chloro-*N*-(4-sulfamoylphenyl) acetamide (**4a**, 0.21 g, 0.85 mmol) was added, followed by a pinch of KI, and heated overnight. Water (10 mL) was added to the reaction mixture, followed by 1 mL 6 N HCl. The resulting solid was filtered, washed with water and air dried, and purification by flash column chromatography (hexane/EtOAc 20:1) afforded the title compound as a yellow powder (0.198 g, 54% yield): 1H NMR (400 MHz, DMSO- d_6): δ = 1.15 (t, J = 6.8 Hz, 6H), 3.50–3 (q, J = 6.8 Hz 4H), 4.88 (s, 2H), 6.58 (s, 1H), 6.82 (d, J = 8.3 Hz, 1H), 7.27 (s, 1H), 7.70 (d, J = 9.0 Hz, 1H), 7.79 (s, 4H), 8.19 (br-s, 1H), 8.68 (s, 1H), 10.53 (s, 1H). ^{13}C NMR (100 MHz, DMSO- d_6): δ = 12.8, 44.8, 63.2, 96.3, 106.6, 107.5, 110.5,

119.2, 127.2, 132.5, 139.2, 141.8, 146.8, 153.6, 157.8, 162.8, 163.4, 165.6. HRMS (ESI): m/z calcd for $C_{22}H_{23}N_3O_7S$ $[M+Na]^+$: 496.4863, found: 496.4835.

2.2 | CA assay

Catalyzed CO_2 hydration activity was evaluated using a photophysics stopped-flow instrument using phenol red (0.2 mM) as an indicator and UV detection at 557 nm as previously described (Ewies et al., 2022). 4-(2-Hydroxyethyl)-1-piperazineethanesulfonic acid (20 mM, pH 7.5) was used as a buffer, and 20 mM Na_2SO_4 to keep its ionic strength constant. Initial rates of the CA-catalyzed CO_2 hydration reaction were applied for 10–100 s. Then, CO_2 concentrations (1.7–17 mM) were applied for kinetic parameters and inhibition constant calculation. The initial velocity for each inhibitor was determined by using a minimum of six traces of the initial 5%–10% of the reaction. Simultaneously, the uncatalyzed rates were likewise measured to be subtracted from the total observed rates.

Stock solutions (0.1 mM) of the new compounds and the reference compound, AAZ, were prepared and subsequently diluted with distilled-deionized water to obtain 0.01 nM concentration. Prior incubation of the mixture of the compounds and enzyme solutions for 15 min at room temperature was carried out until the formation of enzyme–inhibitor complex. Nonlinear least-squares methods were used to calculate the inhibition constants using the Cheng–Prusoff equation.

2.3 | Cell culture

Cells were maintained at 37°C with 5% CO_2 in a humidified environment. Human embryonic kidney cells (HEK293) were purchased from American Type Culture Collection (ATCC). The cells were cultured in Eagle's Minimum Essential Medium (ATCC®, Catalog No. 30-2003™) supplemented with fetal bovine serum (ATCC®, Catalog No. 30-2020) to a final concentration of 10% and Penicillin-Streptomycin Solution (Corning™, Catalog No. MT30001CI) according to the supplier's recommended protocol. Stock solutions of **18f** were prepared in DMSO and were serially diluted for cell culture treatment maintaining the final DMSO concentration at less than 1%. CellTiter 96 aQueous one solution cell proliferation assay (3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2*H*-tetrazolium) (MTS) assay kit (Catalog No. G3580) was purchased from Promega.

2.4 | Cytotoxicity studies

To determine the cell growth inhibition ability of the synthesized compounds, MTS assay was used according to the manufacturer's recommended protocol. Stock solutions of **18f** were prepared in DMSO and were serially diluted for cell culture treatment maintaining

the final DMSO concentration at less than 1%. Cells were seeded at a density of 1×10^5 cells in 96-well plates. After 24 h, cells were treated at the indicated concentrations of **18f**. After incubation at 37°C in an environment of 5% CO₂ for 72 h, 10 μL of MTS reagent (CellTiter 96® AQueous One Solution Reagent) was added to each well and incubated at the above-mentioned conditions for 2–4 h. Absorbance was recorded at 570 nm on a BioTek Synergy Mx multimode plate reader and the viability of cells was plotted as percentage of controls.

3 | RESULTS AND DISCUSSION

3.1 | Chemistry

Coumarin derivatives (2*H*-chromen-2-one derivatives) can be accessed by a number of reported syntheses (Chimenti et al., 2009; He et al., 2014; Yang et al., 2019). Herein, coumarin ethyl ester was synthesized according to the route depicted in Scheme 1 (Ahmed et al., 2019; Sağlık et al., 2019). Commercially available 2,4-dihydroxybenzaldehyde (**1a**) was reacted with diethyl malonate in the presence of piperidine to afford ethyl 7-hydroxy-2-oxo-2*H*-chromene-3-carboxylate (**2a**). The synthesis of sulfonamide derivatives with the chloroacetamide linker were obtained from commercially available sulfonamides (**3a–c**), avoiding the need for Pd/C reduction of the respective nitro intermediates (Kinarivala & Trippier, 2014). These compounds were reacted with 2-chloroacetyl chloride at 0°C in the presence of anhydrous potassium carbonate and a catalytic amount of potassium iodide to afford the corresponding chloroacetamide derivatives (**4a–c**) in good yield (Angeli et al., 2019). Williamson ether synthesis employing **2a** with **4a–c** afforded

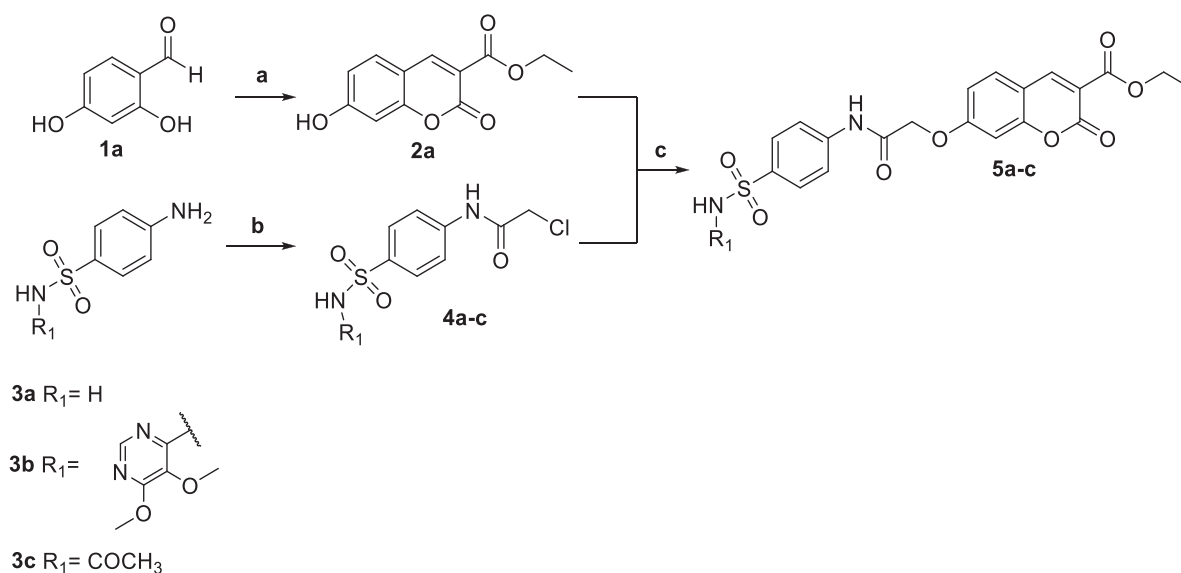
coumarin sulfonamides (**5a–c**) in moderate yields (Table 2) (Jonnalagadda et al., 2022; Yang et al., 2019).

Further coumarin derivatives (**9a–j**) were accessed through hydrolysis of the respective substituted ester of type **2a**. The respective acid chlorides (**8a–j**) were afforded through exposure to refluxing thionyl chloride. The desired coumarin sulfonamide derivatives were obtained via reacting commercially available sulfonamides with either acid **7a–j** through HOBt mediated amide formation or direct reaction with acid chlorides **8a–j** (Scheme 2) (Table 3) (Endo et al., 2017; Mincione et al., 2001; Nishikawa et al., 2019).

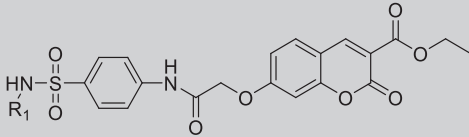
A third derivative class was designed to incorporate a linker between the coumarin acid and the sulfonamide moiety. This derivative was synthesized based on the rationale of a higher rate of hydrolysis and the potential to release subunits of sulfonamide and coumarin which could individually inhibit the activity of CA. 2-Oxo-2-((4-sulfamoylphenyl)amino)ethyl-7-(diethylamino)-2-oxo-2*H*-chromene-3-carboxylate (**22**) was obtained by reaction of 7-(diethylamino)-2-oxo-2*H*-chromene-3-carboxylic acid (**7g**) with 2-chloro-*N*-(4-sulfamoylphenyl)acetamide (**4a**) in the presence of anhydrous potassium carbonate and a catalytic amount of potassium iodide to afford a moderate yield of **22** (Scheme 3) (Yang et al., 2019).

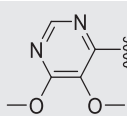

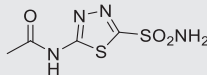
3.2 | Structure–activity relationship

Many reports describe that sulfonamide or coumarin-based molecules are able to inhibit the CA enzymes IX and XII and effect tumor pH, leading to inhibition of growth in both primary tumors and metastatic sites (Emami & Dadashpour, 2015; Lomelino et al., 2016; Neri & Supuran, 2011). Herein we report a number of synthesized compounds show inhibition of tumor associated human CAs IX and XII with selective nanomolar activity.



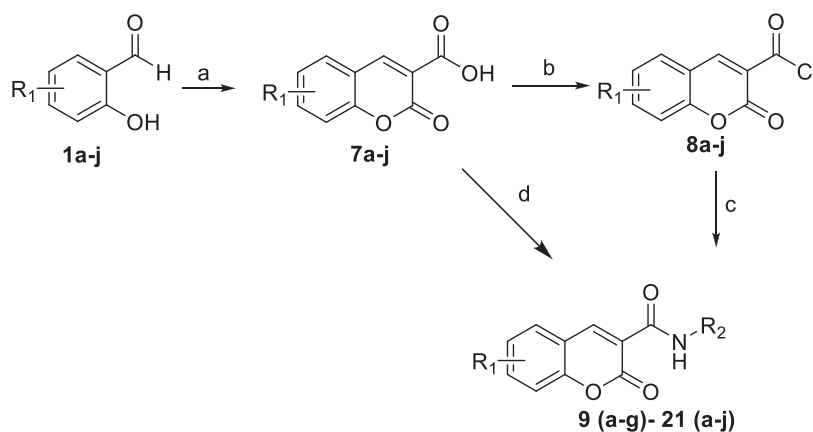
SCHEME 1 Synthesis of carbonic anhydrase inhibitors **5a–c**. Reagents and conditions: (a) Diethyl malonate, piperidine, 3 h, 60°C, 87%; (b) chloroacetyl chloride, K₂CO₃, KI, THF, 2 h, 0°C, 76%–93%; (c) Na₂CO₃, DMF, 155°C, 12 h, 49%–61%.

TABLE 2 Structure, molecular weight, cLogP, polar surface area and CA I, II, IX and XII isoform inhibition activity of synthesized carbonic anhydrase inhibitors from Scheme 1.


Compound	R ₁	Mw	cLogP ^a	PSA ^b	K _i (μM, unless stated) ^c			
					CA I	CA II	CA IX	CA XII
5a	H	446.43	1.37	151.09	738.1 nM	9.1 nM	26.9 nM	20.3 nM
5b		584.56	3.17	180.28	>100	>100	>100	>100
5c		488.47	0.97	154.17	>100	>100	6.1	1.9
AAZ		222.24	-0.98	113.99	250 nM	12.5 nM	25.0 nM	5.7 nM

^aCalculated by ChemDraw Professional 16.0.^bPolar surface area (pH 7.4), calculated by ChemDraw Professional 16.0.^cMean from three different assays, by a stopped flow technique (errors were in the range of ±5%–10% of the reported values).

SCHEME 2 Synthesis of carbonic anhydrase inhibitors **9 (a–g)**–**18 (a–g)**. Reagents and conditions: (a) (i) Diethyl malonate, piperidine, 3 h, 60°C, 70%–92%; (ii) NaOH, methanol, RT, 84%–99%; (b) SOCl₂, 75°C, 5 h, 71%–96%; (c) Appropriate sulfonamide, pyridine, DMF, 12 h, 80°C, 49%–96%; (d) Appropriate sulfonamide, EDC, HOBT•H₂O, Et₃N, DMF, RT, 5 h, 73%.



Twelve of these compounds proved to be more potent compared with acetazolamide (AAZ) control to inhibit CA IX (**9g**, **9i**, **16b–i**, **18f**, **22**), while one was also more potent than AAZ to inhibit CA XII (**18f**). These compounds possess significant selectivity for CA IX and CA XII over CA I and/or CAII.

Ethyl 2-oxo-7-(2-oxo-2-((4-sulfamoylphenyl)amino)ethoxy)-2H-chromene-3-carboxylate (**5a**) possessing a primary sulfonamide with a hydroxyl group in position 7 of the coumarin ring provides selective nanomolar inhibition of CA II, IX, and XII with K_i values of 9.1 nM, 26.9 nM and 20.3 nM, respectively. Conversion of the primary sulfonamide to secondary, such as adding Sulfadoxine (**5b**) and *N*-((4-aminophenyl)sulfonyl)acetamide (**5c**) lead to decrease in this activity in agreement with the literature (Table 2) (Wang et al., 2013).

A range of coumarin side chains featuring various sulfonamide substitutions were synthesized. 7-Hydroxy-2-oxo-*N*-(4-sulfamoylphenyl)-2H-chromene-3-carboxamide (**9a**) is a more potent and selective CA XII inhibitor than the similar compounds **9b–j** with K_i value of 6.2 nM (Table 3). Addition of halide substitution to the coumarin ring results in decreased activity to inhibit CA II, IX, and XII (compounds **9b**, **9c**, **9d**, and **9i**) compared to hydroxyl substitution. (Huwaimel et al., 2021, 2022) Converting the 7-hydroxy in the coumarin ring to 7-diethylamino (**9g**) increases activity to inhibit CA IX with K_i = 13.9 nM compared to AAZ which possesses a K_i of 25 nM. Addition of a second chlorine to the coumarin ring (**9d**) lead to a decrease in activity to inhibit CA II, IX, and XII compared with a single chlorine (**9b**). Homologation of the linker between coumarin and sulfonamide from zero (**9b** and **9f**) to ethyl (**20b** and **20f**)

TABLE 3 Inhibition data of human CA isoforms I, II, IX and XII, structure, molecular weight, calculated logP and polar surface area of synthesized carbonic anhydrase inhibitors from Schemes 2–3.

Compound	Structure	Mw	cLogP ^a	PSA ^b	KI (μM, unless stated) ^c			
					CA I	CA II	CA IX	CA XII
9a		360.34	0.44	135.79	265.4 nM	12.4 nM	29.2 nM	6.2 nM
10a		466.47	2.11	146.52	64.8	29.1	17.4	8.3
11a		468.44	1.57	155.75	68.9	24.0	8.1	4.7
17a		498.47	2.24	165	>100	>100	>100	>100
19a		438.41	1.11	147	>100	>100	25.8	50.1
9b		378.78	1.64	115.56	818.4 nM	63.5 nM	52.3 nM	28.4 nM
10b		484.91	3.31	126.29	>100	>100	>100	>100
11b		486.88	2.77	135.52	>100	>100	>100	>100
12b		455.87	3.05	113.93	>100	>100	28.4	9.1
13b		476.91	2.63	126.29	>100	>100	58.8	>100
14b		516.91	4.19	144.75	>100	>100	>100	>100
15b		420.82	1.32	118.64	>100	37.2	6.1	3.4
16b		378.78	1.64	115.56	639.2 nM	44.9 nM	8.4 nM	29.6 nM

TABLE 3 (Continued)

Compound	Structure	Mw	cLogP ^a	PSA ^b	KI (μM, unless stated) ^c			
					CA I	CA II	CA IX	CA XII
20b		406.84	1.92	115.56	23.5 nM	8.6 nM	35.8 nM	46.0 nM
21b		378.78	1.64	115.56	726.5 nM	139.4 nM	56.2 nM	66.6 nM
9c		423.24	1.79	115.56	1422 nM	85.4 nM	67.6 nM	16.2 nM
10c		529.37	3.46	126.29	>100	>100	>100	>100
11c		531.34	2.92	135.52	>100	>100	>100	>100
12c		500.32	3.20	113.93	>100	>100	39.1	>100
13c		521.36	2.78	126.29	>100	>100	>100	>100
14c		561.36	4.34	144.75	>100	>100	>100	>100
15c		465.27	1.47	118.64	>100	46.8	0.91	10.6
16c		423.24	1.79	115.56	1961 nM	76.3 nM	14.4 nM	54.2 nM
21c		423.24	1.79	115.56	1569 nM	436.8 nM	79.5 nM	156.7 nM
9d		413.23	2.35	115.56	2322 nM	238.4 nM	104.9 nM	46.3 nM
11d		521.33	3.48	135.52	>100	>100	>100	>100

(Continues)

TABLE 3 (Continued)

Compound	Structure	Mw	cLogP ^a	PSA ^b	KI (μM, unless stated) ^c			
					CA I	CA II	CA IX	CA XII
12d		490.31	3.76	113.93	>100	>100	>100	>100
12e		421.43	2.32	113.93	>100	>100	39.1	>100
14e		482.47	3.47	144.75	>100	>100	>100	>100
15e		386.38	0.60	118.64	>100	10.3	0.79	6.3
16e		344.34	0.92	115.56	192.8 nM	53.5 nM	18.9 nM	69.8 nM
17e		482.47	2.72	144.75	>100	>100	>100	>100
21e		344.34	0.92	115.56	552.4 nM	126.5 nM	75.2 nM	42.0 nM
9f		374.37	0.89	179.24	1571 nM	99.9 nM	60.1 nM	29.0 nM
10f		480.50	2.56	135.52	>100	>100	>100	>100
11f		482.47	2.02	144.75	>100	>100	>100	>100
12f		451.45	2.30	123.16	>100	>100	23.2	>100
15f		416.40	0.58	127.87	90.1	28.7	14.5	3.7
16f		374.37	0.89	124.79	774.1 nM	212.3 nM	5.2 nM	52.1 nM

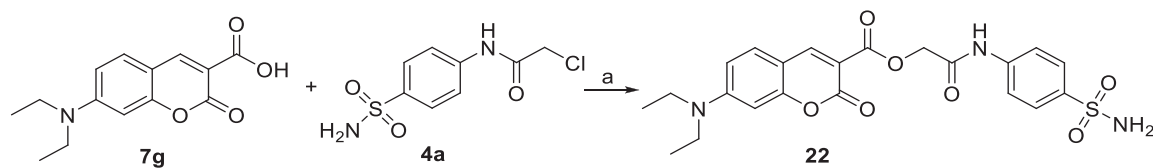
TABLE 3 (Continued)

Compound	Structure	Mw	cLogP ^a	PSA ^b	KI (μM, unless stated) ^c			
					CA I	CA II	CA IX	CA XII
18f		521.44	-0.30	184.95	954.6 nM	514.5 nM	21.1 nM	5.1 nM
20f		402.42	1.17	124.79	654.4 nM	56.8 nM	18.7 nM	45.8 nM
21f		374.37	0.89	124.79	832.1 nM	110.9 nM	35.7 nM	61.2 nM
9g		415.46	2.20	118.80	590.1 nM	26.8 nM	13.9 nM	22.1 nM
9h		374.37	0.89	124.79	575.3 nM	42.7 nM	31.8 nM	22.9 nM
16h		374.37	0.89	124.79	185.3 nM	68.7 nM	34.2 nM	18.3 nM
9i		362.33	1.07	115.56	665.2 nM	51.2 nM	25.0 nM	31.4 nM
16i		362.33	1.07	115.56	225.4 nM	75.2 nM	15.2 nM	49.5 nM
9j		400.45	2.74	115.56	52.0 nM	16.4 nM	28.7 nM	22.9 nM
22		473.50	2.22	145.10	74.4 nM	23.0 nM	19.6 nM	65.7 nM
AAZ		222.24	-0.98	113.99	250 nM	12.5 nM	25.0 nM	5.7 nM

^aCalculated by ChemDraw Professional 16.0.

^bPolar surface area (pH 7.4), calculated by ChemDraw Professional 16.0.

^cMean from three different assays, by a stopped flow technique (errors were in the range of ±5%–10% of the reported values).



SCHEME 3 Synthesis of carbonic anhydrase inhibitor **22**. Reagents and conditions: (a) K₂CO₃, KI, DMF, 155°C, 12 h, 54%.

lead to a approximate two-fold decrease in the activity to inhibit CA XII (Ki's of [9b] 28 nM to [20b] 46 nM and [9f] 29 nM to [20f] 46 nM) while increasing activity to inhibit CA II and IX (Ki's of [9b] 64 and 52 nM to [20b] 9 and 36 nM and [9f] 100 and 60 nM to [20f] 57 and 19 nM, respectively). 6-Chloro-2-oxo-N-(4-sulfamoylphenethyl)-2H-chromene-3-carboxamide (20b) is more potent and selective to CA II than other synthesized compounds with a Ki of 8.6 nM compared to AAZ with a Ki of 12.5 nM.

A substituent screen of the terminal sulfonamide of 6-chloro substituted coumarins revealed a preference of *meta* (16b) > *para* (9b) > *ortho* (21b) in terms of activity for CA I, II, and IX, with Ki's of 8.4, 52, and 68 nM, respectively for CA IX. For CA XII *ortho* > *para* = *meta* with Ki's of 16, 28, and 30 nM for 16b, 9b, and 21b, respectively. The greater activity of the *meta* position for CA IX inhibition was confirmed when the 6-chloro substituent of the coumarin ring was converted to 6-methoxy with compound 16f possessing a Ki of 5 nM for CA IX which was greater than *ortho* (21f) and *para* (9f). *Meta*-sulfonamides 16b and 16f represent the most potent CA IX inhibitors identified, at least three-fold more potent than AAZ control. Changing the methoxy group in the coumarin ring from position 6 (16f) to position 7 (16h) lead to decreased CA IX inhibition with Ki values 5.2 to 34.2 nM, respectively (Table 3).

Increasing the number of sulfonamides to two groups on the terminal phenyl ring with 2,4-disubstitution and addition of a CF₃ moiety at the 5-position (18f) lead to further increased activity and selectivity with Ki's = 955, 515, 21, and 5 nM for CA's I, II, IX, and XII, respectively. Compound 18f represents the most active CA XII inhibitor synthesized, equipotent with AAZ control (5.7 nM). Interestingly, addition of an ester linkage which would be expected to be metabolically labile in vivo systems, results in a pan CA I, II, IX, and XII inhibitor with Ki's of 74, 23, 20, and 66 nM, respectively.

As expected, most compounds possessing secondary sulfonamides possessed generally lower activity in inhibiting CA enzymes than their primary sulfonamide counterparts. As shown in Table 3, most of the secondary sulfonamides, especially those with functionalized aromatic heterocycles, proved inactive to inhibit CA, with Ki values >100 μM (10-14a-e, 17a-e, 19a). *N*-acetyl functionalized secondary sulfonamides 15b-f possess some activity to inhibit CA's I, II, IX, and XII. *N*-(4-(*N*-acetylsulfamoyl)phenyl)-6-bromo-2-oxo-2H-chromene-3-carboxamide (15c) and *N*-(4-(*N*-acetylsulfamoyl)phenyl)-2-oxo-2H-chromene-3-carboxamide (15e) possess Ki's of 910 and 790 nM, respectively, to inhibit CA XI.

To determine the cytotoxic effect of the most active compound against low tumorigenic cells, human embryonic kidney (HEK293) cells were incubated with varying concentrations (0, 1, 10, 20, 30, 50, and 100 μM) of 18f for 72 h and cell viability measured with MTS assay (Figure 1). The highly potent CA inhibitor elicited no cytotoxicity up to 50 μM and 97% cell viability at 100 μM.

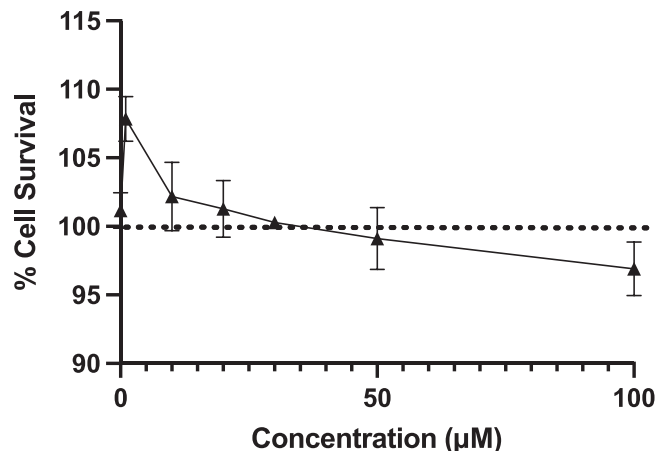


FIGURE 1 Cell viability of HEK293 cells treated with varying concentrations of 18f (0–100 μM) for 72 h. Dashed line indicates 100% cell viability.

4 | CONCLUSION

In summary, a library of sulfonamide functionalized coumarins has been synthesized that revealed several compounds with high activity to inhibit human CAs I, II, IX, and XII with many displaying selectivity to the tumor-associated CA IX and CA XII. These CA inhibitors are exemplified by compound 16b possessing Ki's of 8.4, 30, 45, and 639 nM to CA IX, XII, II, and I, respectively with 5- to 76-fold selectivity for the cancer-associated CAs and compound 16f possessing Ki's of 5, 52, 212, and 774 nM for CA IX, XII, II, and I, respectively with 42- to 155-fold selectivity for cancer associated CAs. Both compounds are more potent than the clinical CA inhibitor acetazolamide. Compound 18f (Ki's = 5, 21, 515, and 955 nM for CA's XII, IX, II, and I, respectively) is representative of a novel CA IX and XII inhibitor that possess no general cytotoxic effect in low tumorigenic cells. All three compounds represent novel structures for further development.

ACKNOWLEDGMENTS

We thank the National Cancer Institute of the National Institutes of Health (R01 CA226436), the Eunice Kennedy Shriver National Institute of Child Health and Human Development of the National Institutes of Health (R01 HD106590), University of Nebraska Medical Center, and Fred and Pamela Buffett Cancer Center Grant (P30 CA036727) for funding (P.C.T.). B.I.H thanks the University of Hail, Hail, Saudi Arabia for a graduate scholarship. The content is solely the responsibility of the authors and does not represent the official views of the National Institutes of Health.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

The data that supports the findings of this study are available in the supplementary material of this article. ¹H and ¹³C NMR spectra of the

designed compounds are available in the supporting information material of this article.

ORCID

Bader I. Huwaimel  <http://orcid.org/0000-0002-7813-117X>

Shirisha Jonnalagadda  <http://orcid.org/0000-0002-5403-4383>

Claudiu T. Supuran  <http://orcid.org/0000-0003-4262-0323>

Paul C. Trippier  <http://orcid.org/0000-0002-4947-5782>

REFERENCES

- Ahmed, A., Channar, P. A., Saeed, A., Kalesse, M., Kazi, M. A., Larik, F. A., Abbas, Q., Hassan, M., Raza, H., & Seo, S. Y. (2019). Synthesis of sulfonamide, amide and amine hybrid pharmacophore, an entry of new class of carbonic anhydrase II inhibitors and evaluation of chemo-informatics and binding analysis. *Bioorganic Chemistry*, 86, 624–630. <https://doi.org/10.1016/j.bioorg.2019.01.060>
- Angeli, A., Di Cesare Mannelli, L., Ghelardini, C., Peat, T. S., Bartolucci, G., Menicatti, M., Carta, F., & Supuran, C. T. (2019). Benzensulfonamides bearing spirohydantoin moieties act as potent inhibitors of human carbonic anhydrases II and VII and show neuropathic pain attenuating effects. *European Journal of Medicinal Chemistry*, 177, 188–197. <https://doi.org/10.1016/j.ejmech.2019.05.058>
- Chimenti, F., Secci, D., Bolasco, A., Chimenti, P., Bizzarri, B., Granese, A., Carradori, S., Yáñez, M., Orallo, F., Ortuso, F., & Alcaro, S. (2009). Synthesis, molecular modeling, and selective inhibitory activity against human monoamine oxidases of 3-carboxamido-7-substituted coumarins. *Journal of Medicinal Chemistry*, 52(7), 1935–1942. <https://doi.org/10.1021/jm801496u>
- Emami, S., & Dadashpour, S. (2015). Current developments of coumarin-based anti-cancer agents in medicinal chemistry. *European Journal of Medicinal Chemistry*, 102, 611–630. <https://doi.org/10.1016/j.ejmech.2015.08.033>
- Endo, S., Xia, S., Suyama, M., Morikawa, Y., Oguri, H., Hu, D., Ao, Y., Takahara, S., Horino, Y., Hayakawa, Y., Watanabe, Y., Gouda, H., Hara, A., Kuwata, K., Toyooka, N., Matsunaga, T., & Ikari, A. (2017). Synthesis of potent and selective inhibitors of Aldo-Keto reductase 1B10 and their efficacy against proliferation, metastasis, and cisplatin resistance of lung cancer cells. *Journal of Medicinal Chemistry*, 60(20), 8441–8455. <https://doi.org/10.1021/acs.jmedchem.7b00830>
- Ewies, E. F., Sabry, E., Bekheit, M. S., Fouad, M. A., Vullo, D., & Supuran, C. T. (2022). Click chemistry-based synthesis of new benzenesulfonamide derivatives bearing triazole ring as selective carbonic anhydrase II inhibitors. *Drug Development Research*, 83(6), 1281–1291. <https://doi.org/10.1002/ddr.21957>
- He, X., Chen, Y. Y., Shi, J. B., Tang, W. J., Pan, Z. X., Dong, Z. Q., Song, B. A., Li, J., & Liu, X. H. (2014). New coumarin derivatives: Design, synthesis and use as inhibitors of hMAO. *Bioorganic & Medicinal Chemistry*, 22(14), 3732–3738. <https://doi.org/10.1016/j.bmc.2014.05.002>
- Huwaimel, B. I., Bhakta, M., Kulkarni, C. A., Milliken, A. S., Wang, F., Peng, A., Brookes, P. S., & Trippier, P. C. (2021). Discovery of halogenated benzothiadiazine derivatives with anticancer activity. *ChemMedChem*, 16(7), 1143–1162. <https://doi.org/10.1002/cmdc.202000729>
- Huwaimel, B. I., Jonnalagadda, S., Jonnalagadda, S., Zahra, F. T., Nocentini, A., Supuran, C. T., Mikelis, C. M., & Trippier, P. C. (2022). Chlorinated benzothiadiazines inhibit angiogenesis through suppression of VEGFR2 phosphorylation. *Bioorganic & Medicinal Chemistry*, 67, 116805. <https://doi.org/10.1016/j.bmc.2022.116805>
- Jonnalagadda, S. K., Huwaimel, B. I., Jonnalagadda, S., Garrison, J. C., & Trippier, P. C. (2022). Access to highly strained tricyclic ketals derived from coumarins. *The Journal of Organic Chemistry*, 87(6), 4476–4482. <https://doi.org/10.1021/acs.joc.2c00018>
- Kinarivala, N., & Trippier, P. C. (2014). Exploration of relative chemoselectivity in the hydrodechlorination of 2-chloropyridines. *Tetrahedron Letters*, 55(39), 5386–5389. <https://doi.org/10.1016/j.tetlet.2014.08.008>
- Lee, S. H., & Griffiths, J. R. (2020). How and why are cancers acidic? carbonic anhydrase IX and the homeostatic control of tumour extracellular pH. *Cancers*, 12(6), 1616. <https://doi.org/10.3390/cancers12061616>
- Lomelino, C., Supuran, C., & McKenna, R. (2016). Non-classical inhibition of carbonic anhydrase. *International Journal of Molecular Sciences*, 17(7), 1150. <https://doi.org/10.3390/ijms17071150>
- Maresca, A., Temperini, C., Pochet, L., Masereel, B., Scozzafava, A., & Supuran, C. T. (2010). Deciphering the mechanism of carbonic anhydrase inhibition with coumarins and thiocoumarins. *Journal of Medicinal Chemistry*, 53(1), 335–344. <https://doi.org/10.1021/jm901287j>
- McDonald, P. C., Chafe, S. C., Supuran, C. T., & Dedhar, S. (2022). Cancer therapeutic targeting of hypoxia induced carbonic anhydrase IX: From bench to bedside. *Cancers*, 14(14), 3297. <https://doi.org/10.3390/cancers14143297>
- Mincione, F., Starnotti, M., Menabuoni, L., Scozzafava, A., Casini, A., & Supuran, C. T. (2001). Carbonic anhydrase inhibitors: 4-sulfamoyl-benzenecarboxamides and 4-chloro-3-sulfamoyl-benzenecarboxamides with strong topical antiglaucoma properties. *Bioorganic & Medicinal Chemistry Letters*, 11(13), 1787–1791. [https://doi.org/10.1016/s0960-894x\(01\)00303-1](https://doi.org/10.1016/s0960-894x(01)00303-1)
- Neri, D., & Supuran, C. T. (2011). Interfering with pH regulation in tumours as a therapeutic strategy. *Nature Reviews Drug Discovery*, 10(10), 767–777. <https://doi.org/10.1038/nrd3554>
- Nishikawa, Y., Miki, T., Awa, M., Kuwata, K., Tamura, T., & Hamachi, I. (2019). Development of a nitric oxide-responsive labeling reagent for proteome analysis of live cells. *ACS Chemical Biology*, 14(3), 397–404. <https://doi.org/10.1021/acschembio.8b01021>
- Sağlık, B. N., Çevik, U. A., Osmaniye, D., Levent, S., Çavuşoğlu, B. K., Demir, Y., İlgin, S., Özkay, Y., Kopal, A. S., Beydemir, Ş., & Kaplançıklı, Z. A. (2019). Synthesis, molecular docking analysis and carbonic anhydrase I-II inhibitory evaluation of new sulfonamide derivatives. *Bioorganic Chemistry*, 91, 103153. <https://doi.org/10.1016/j.bioorg.2019.103153>
- Supuran, C. T. (2008). Carbonic anhydrases: Novel therapeutic applications for inhibitors and activators. *Nature Reviews Drug Discovery*, 7(2), 168–181. <https://doi.org/10.1038/nrd2467>
- Supuran, C. T. (2010). Carbonic anhydrase inhibitors. *Bioorganic & Medicinal Chemistry Letters*, 20(12), 3467–3474. <https://doi.org/10.1016/j.bmcl.2010.05.009>
- Supuran, C. T. (2020). Coumarin carbonic anhydrase inhibitors from natural sources. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 35(1), 1462–1470. <https://doi.org/10.1080/14756366.2020.1788009>
- Thacker, P. S., Alvola, M., Arifuddin, M., Angeli, A., & Supuran, C. T. (2019). Design, synthesis and biological evaluation of coumarin-3-carboxamides as selective carbonic anhydrase IX and XII inhibitors. *Bioorganic Chemistry*, 86, 386–392. <https://doi.org/10.1016/j.bioorg.2019.02.004>
- Touissni, N., Maresca, A., McDonald, P. C., Lou, Y., Scozzafava, A., Dedhar, S., Winum, J. Y., & Supuran, C. T. (2011). Glycosyl coumarin carbonic anhydrase IX and XII inhibitors strongly attenuate the growth of primary breast tumors. *Journal of Medicinal Chemistry*, 54(24), 8271–8277. <https://doi.org/10.1021/jm200983e>

- Wang, Z. C., Qin, Y. J., Wang, P. F., Yang, Y. A., Wen, Q., Zhang, X., Qiu, H. Y., Duan, Y. T., Wang, Y. T., Sang, Y. L., & Zhu, H. L. (2013). Sulfonamides containing coumarin moieties selectively and potently inhibit carbonic anhydrases II and IX: Design, synthesis, inhibitory activity and 3D-QSAR analysis. *European Journal of Medicinal Chemistry*, 66, 1–11. <https://doi.org/10.1016/j.ejmech.2013.04.035>
- Williams, K. J., & Gieling, R. G. (2019). Preclinical evaluation of ureidosulfamate carbonic anhydrase IX/XII inhibitors in the treatment of cancers. *International Journal of Molecular Sciences*, 20(23), 6080. <https://doi.org/10.3390/ijms20236080>
- Yang, F., Zhao, N., Song, J., Zhu, K., Jiang, C., Shan, P., & Zhang, H. (2019). Design, synthesis and biological evaluation of novel coumarin-based hydroxamate derivatives as histone deacetylase (Hdac) inhibitors with antitumor activities. *Molecules*, 24(14), 2569. <https://doi.org/10.3390/molecules24142569>

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Huwaimel, B. I., Jonnalagadda, S. K., Jonnalagadda, S., Kumari, S., Nocentini, A., Supuran, C. T., & Trippier, P. C. (2023). Selective carbonic anhydrase IX and XII inhibitors based around a functionalized coumarin scaffold. *Drug Development Research*, 84, 681–702. <https://doi.org/10.1002/ddr.22049>