

Beneath the surface of colorectal cancer: Unmasking the evolving nature of (Neo)RAS

Giulia Massaro^{a,1}, Jacopo Venturini^{a,1}, Daniele Rossini^{b,c,*}, Agnese Vannini^a, Marco Brugia^a, Daniele Lavacchi^a, Cristiana Conticello^a, Irene Valle^a, Delia Ravizza^a, Serena Pillozzi^d, Lorenzo Antonuzzo^{a,b,c}

^a Clinical Oncology Unit, Careggi University Hospital, Florence, Italy

^b Department of Experimental and Clinical Medicine, University of Florence, Florence, Italy

^c Department of Health Science, University of Florence, Florence, Italy

^d Department of Experimental and Clinical Biomedical Sciences 'Mario Serio', University of Florence, Florence, Italy

ARTICLE INFO

Keywords:

MCRC
KRAS/NRAS mutations
Anti-EGFR therapy
CtDNA
NeoRAS WT phenomenon

ABSTRACT

Metastatic colorectal cancer (mCRC) remains a major clinical challenge, despite therapeutic advancements. Mutations in KRAS and NRAS (RAS) oncogenes drive resistance to anti-EGFR drugs, necessitating RAS mutational analysis prior to treatment. While tissue biopsy remains the gold standard for molecular profiling, it has limitations such as invasiveness, intra-tumoral heterogeneity, and delayed results. Liquid biopsy (LB), on the other hand, offers a non-invasive alternative by analyzing circulating tumor DNA (ctDNA) and it provides a dynamic view of molecular changes over time. Indeed, ctDNA analysis has expanded the understanding of the mCRC's molecular landscape, revealing that RAS mutated (MT) subclones undergo both a positive and a negative selection during treatment. This negative selection has been described as the "NeoRAS WT phenomenon." The temporary disappearance of RAS mutations opens "RAS WT windows," thus making potential candidates for anti-EGFR therapies even patients initially diagnosed as RAS MT. This review examines numerous studies investigating the clinical significance of the "NeoRAS WT phenomenon" as a distinct pathological entity. It also highlights the key limitations arising from the variability in study designs, detecting methods and ctDNA shedding rates. The results of ongoing prospective trials are necessary to determine whether NeoRAS WT stands as a reliable marker for guiding anti-EGFR treatment strategies in RAS-mutated patients.

1. Introduction

Metastatic colorectal cancer (mCRC) remains a major clinical challenge. Advances in understanding its molecular complexity have led to significant improvements in clinical outcomes. KRAS and NRAS exon 2, 3, and 4 (RAS) oncogenic mutations occur in about 45 % of patients with mCRC, activating the RAS/RAF/MEK/ERK pathway and conferring resistance to anti-epidermal growth factor receptor (EGFR) monoclonal antibodies (moAbs) (Porru et al., 2018), (Martinelli et al., 2020). For RAS mutant (RAS MT) mCRC, current clinical guidelines recommend first-line chemotherapy with either the cytotoxic FOLFOXIRI triplet or the FOLFOX/CAPOX/FOLFIRI doublets, in combination with anti-vascular endothelial growth factor (anti-VEGF) agents (Douillard et al., 2013; Bokemeyer et al., 2015; Cervantes et al., 2023; Benson et al.,

2024). Conversely, patients with RAS and RAF wild type (WT) mCRC are eligible for either FOLFOX or FOLFIRI combined with anti-EGFR moAbs (i.e. Panitumumab or Cetuximab), particularly for left-sided tumors (Cervantes et al., 2023; Benson et al., 2024; Lenz et al., 2014; Stintzing et al., 2016). As a second-line strategy, anti-angiogenic agents (i.e. Bevacizumab, Aflibercept, or Ramucirumab) in combination with chemotherapy represent the standard of care option for both RAS MT and WT patients. In the presence of BRAF V600 gene mutations, a targeted approach with Encorafenib and Cetuximab has been approved (Cervantes et al., 2023; Benson et al., 2024). Among third-line treatment options, regorafenib, trifluridine/tipiracil, and fruquintinib have demonstrated limited efficacy in clinical trials, yielding modest objective response rates (ORR) and relatively short progression-free survival (PFS) and overall survival (OS) (Mayer et al., 2015; Prager et al., 2023;

* Corresponding author at: Department of Experimental and Clinical Medicine, University of Florence, Florence, Italy.

E-mail address: daniele.rossini@unifi.it (D. Rossini).

¹ Joint first authors.

<https://doi.org/10.1016/j.critrevonc.2025.104746>

Received 26 October 2024; Received in revised form 23 April 2025; Accepted 24 April 2025

Available online 26 April 2025

1040-8428/© 2025 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Dasari et al., 2023; Grothey et al., 2013). Given these limitations, ESMO guidelines recommend rechallenging pretreated RAS WT mCRC patients with anti-EGFR MoAbs, especially those who responded well initially (Cervantes et al., 2023). This approach, supported by liquid biopsy-driven selection, has shown promising results in small, non-randomized studies (Martinelli et al., 2021; Cremolini et al., 2019). In contrast, RAS MT patients are not eligible for the same strategy due to their well-established intrinsic resistance mechanisms (Cervantes et al., 2023). However, emerging evidence suggests that liquid biopsy (LB) may overcome these limitations (Gazzaniga et al., 2018). In response to treatment pressure and tumor microenvironment changes, mCRC's mutational landscape exhibits a dynamic and heterogeneous nature, underscoring the need for new targeting strategies (Nicolazzo et al., 2022). To address these challenges, this review explores RAS mutation dynamics in the era of LB, focusing on current and emerging evidence

regarding anti-EGFR blockade in RAS MT patients.

1.1. The ctDNA revolution in CRC

In the era of precision medicine, the rapid evolution of targeted therapies demands advanced diagnostics to optimize patient selection. Despite being the gold standard for molecular profiling, tumor tissue biopsy carries notable limitations, including invasiveness and risks of severe complications (i.e. pain, bleeding, pneumothorax, pancreatitis, etc.) (Overman et al., 2013). Additionally, tumor cell yield may be insufficient for analysis, and sample preservation can be challenging (Diaz and Bardelli, 2014). Intratumoral heterogeneity, which varies spatially and temporally, further complicates tumor characterization, as single-site biopsies may miss distinct tumor subpopulations dispersed within a lesion, while the impracticality of serial sampling limits the

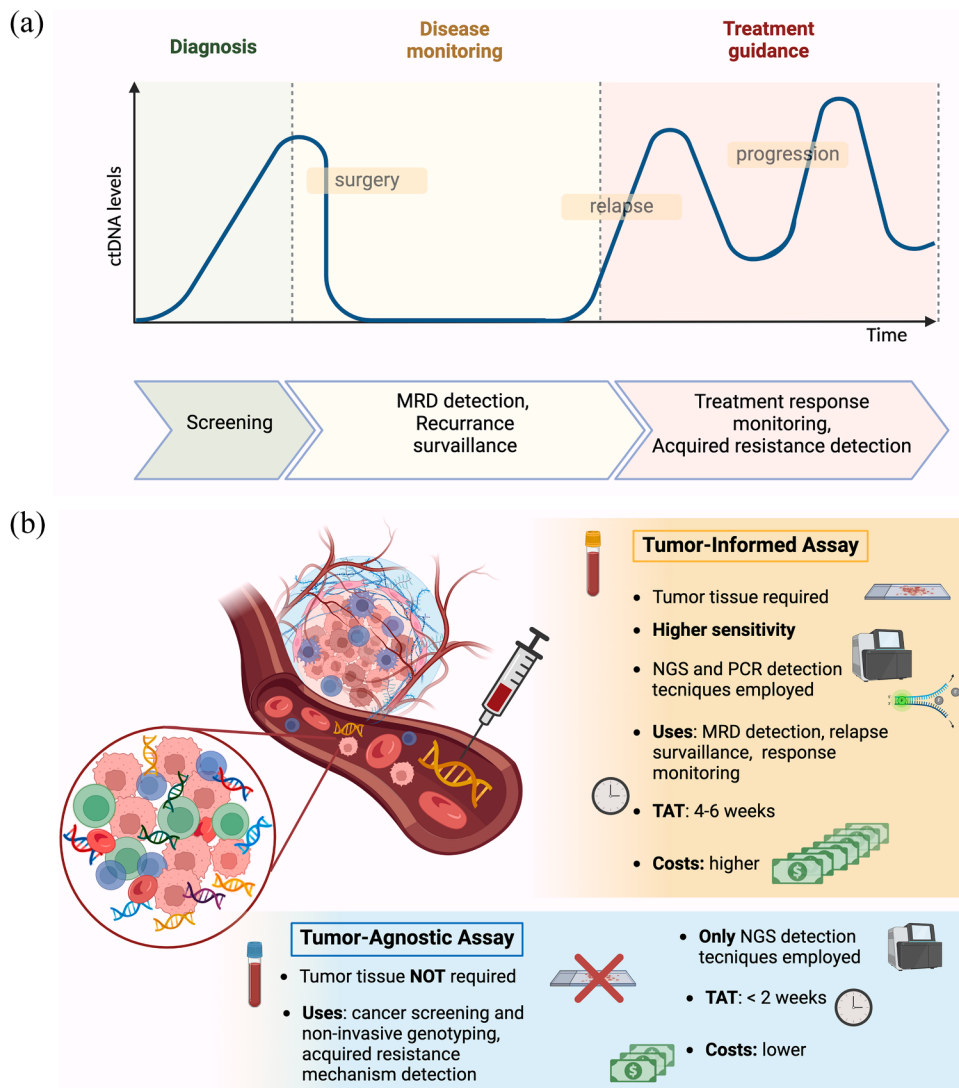


Fig. 1. A. The different clinical settings for ctDNA's applicability. LB finds a wide range of uses, starting from cancer screening to treatment monitoring in advanced stages, thus portraying dynamically the disease's evolution in response to surgical and medical interventions. Abbreviations NGS, Next-Generation Sequencing, PCR, Polymerase Chain Reaction, MRD, Minimal Residual Disease, ctDNA Circulating Tumor DNA; LB, liquid biopsy. Image created with BioRender.com, accessed on 28 August 2024. B. Types of ctDNA testing modalities. Tumor-informed assays track specific plasmatic mutations based on previously sequenced data from tissue samples, enhancing sensitivity at the cost of higher expenses and longer turnaround times (TAT). Conversely, tumor-agnostic methods permit a broader but less targeted plasma analysis, without prior knowledge of the tumor's characteristics. Abbreviations NGS, Next-Generation Sequencing, PCR, Polymerase Chain Reaction, MRD, Minimal Residual Disease, ctDNA Circulating Tumor DNA; LB, liquid biopsy; TAT, turnaround time. Image created with BioRender.com, accessed on 28 August 2024.

detection of dynamic changes over time (Gerlinger et al., 2012; Findlay et al., 2016; Venesio et al., 2018; Dagogo-Jack and Shaw, 2018). Lastly, the prolonged turnaround time (TAT) for histological reports delay treatment decision-making (Lone et al., 2022).

To address these challenges, LB has emerged as a non-invasive tool for tumor analysis, enabling the detection of cancer-derived components in the bloodstream, including circulating cell-free DNA (cfDNA), circulating tumor cells (CTCs), extracellular vesicles (EVs), and regulatory RNAs (siRNAs, miRNAs), aiming to identify DNA mutations, copy number variations (CNVs) of crucial genes, epigenetic patterns, and the proteome's and metabolome's characteristics (Gazzaniga et al., 2018; Overman et al., 2013; Diaz and Bardelli, 2014; Venesio et al., 2018; Nakamura et al., 2019; Siravegna et al., 2019a; Kim et al., 2016; Mayers et al., 2014). Circulating tumor DNA (ctDNA) represents the tumor-specific fraction of cfDNA, accounting for 0.1–10 % of total cfDNA and characterized by shorter DNA fragments (~145 bp vs. ~167 bp for cfDNA). Its levels vary widely based on tumor location, histology, disease burden and setting, thus influencing the choice of the detection method (Fig. 1. A). In cases of low shedding rates, sensitive targeted approaches are required, such as PCR-based (i.e., droplet digital PCR (ddPCR), BEAMing-PCR) or Next Generation Sequencing (NGS)-based (Safe-SeqS, CAPP-Seq, and TAmSeq) techniques. Post-surgical minimal residual disease (MRD) detection represents a solid application of these targeted strategies, where plasma analysis is guided by the molecular profiling of the resected tumor (i.e., tumor-informed method; Fig. 1. B). Conversely, high ctDNA levels favor untargeted strategies, such as the NGS-based genome-wide analysis (i.e. whole exome sequencing (WES), whole-genome sequencing (WGS), or whole methylome sequencing), which capture a wide range of mutations but with lower sensitivity. This approach is applied in advanced cancer care to identify new therapeutic targets without relying on tissue specimens (i.e. tumor agnostic technique; Fig. 1. B) (Mauri et al., 2022; Krebs et al., 2022; Xin et al., 2023; Rolfo et al., 2020).

These technologies found several applications in the CRC field, due to its optimal shedding properties compared to other tumors (Mauri et al., 2022; Malla et al., 2022). LB-based MRD detection has proven to be a strong negative prognostic marker, predicting relapse in 95–100 % of cases and preceding radiologically proven recurrence by a median of 8.7–12 months (Diehl et al., 2008; Reinert et al., 2019; Tie et al., 2019). Thus, a ctDNA-based treatment decision making for the CRC adjuvant setting is under evaluation, obtaining promising results in recent randomized trials (Tie et al., 2022; Henriksen et al., 2021; Lonardi et al., 2020; Oki et al., 2023). In advanced stages, ctDNA displayed a high concordance with tissue sampling (84–100 %) and effectively detected drug resistance mechanisms (Nakamura et al., 2020; Loree et al., 2021; Sartore-Bianchi et al., 2019). Additionally, by assessing plasma gene expression levels, LB improved the selection of patients most likely to benefit from targeted therapies (Siravegna et al., 2019a; Oki et al., 2023; Siena et al., 2021; Meric-Bernstam et al., 2019; Sartore-Bianchi et al., 2016; Venturini et al., 2024). Finally, tumor-agnostic WGS profiling showed promising results as a diagnostic tool for early-stage CRC screening (Chung et al., 2024).

1.2. RAS heterogeneity in mCRC and NeoRAS WT concept

Treatment failure in mCRC is driven by its intrinsic heterogeneity (Chan and Buczacki, 2021). Under therapy-induced selective pressure, distinct resistant tumor subclones emerge and predominate (Siena et al., 2018a, 2018b; Morelli et al., 2015; Misale et al., 2012; Siravegna et al., 2019b). LB has demonstrated its ability to track this evolving mutational landscape, capturing the emergence of resistance mechanisms such as acquired RAS, BRAF, MEK, and EGFR mutations, as well as ERBB2 and MET amplifications (Cremolini et al., 2019; Parseghian et al., 2019; Peeters et al., 2015). For instance, emerging evidence now supports anti-EGFR rechallenge after anti-EGFR-based second-line therapies, based on the absence of RAS MT subclones in ctDNA samples (Martinelli

et al., 2021; Cremolini et al., 2019; Ciardiello et al., 2021; Kagawa et al., 2022; Sartore-Bianchi et al., 2022; Sunakawa et al., 2022). The rationale behind this approach is rooted in the pulsatile behavior of RAS WT subclones, whose levels exponentially decline in the absence of EGFR blockade (Morelli et al., 2015; Parseghian et al., 2019). Within this framework, a similar dynamic process has been described in primarily RAS MT mCRC, known as the NeoRAS WT phenomenon. It is hypothesized that, in response to first-line treatments, RAS WT subclones outgrow the initially predominant RAS MT ones, which decline below the detection threshold, thus creating a transient "RAS-WT window" (Fig. 2) (Nicolazzo et al., 2022; Klein-Scory et al., 2020; Sato et al., 2022). This pattern suggests a process of "conversion" to a different subclonal population rather than a true biological "reversion" of RAS MT cells. While some evidence supports the contribution of both chemotherapy and antiangiogenic therapy to RAS WT conversions, (Bouchahda et al., 2020; Nicolazzo et al., 2020; Henry et al., 2020) multiple LB-based studies highlight the greater impact achieved by bevacizumab-containing regimens (Raimondi et al., 2019; Sunakawa et al., 2020; Osumi et al., 2023). Notably, the same NeoRAS WT converting effect of VEGF-inhibition was also confirmed by Epistolio et al. on 28 tissue specimens from liver metastases (Epistolio et al., 2021). To explain this, it has been postulated that bevacizumab-induced hypoxia enhances the sensitivity of RAS MT cells to ROS-mediated cell death (Henry et al., 2020). In light the growing evidence, the following sections will explore the retrospective and prospective investigations into the treatment of NeoRAS WT mCRC as a distinct pathological entity.

2. Current evidence on NeoRAS WT

Research on the NeoRAS WT phenomenon has primarily focused on three key aspects: identifying the most effective diagnostic techniques and optimal timing for assessment, evaluating its predictive value for anti-EGFR treatment response, and determining its prognostic significance following anti-angiogenic therapy. The main findings are detailed in Table 1.

The first studies investigated whether RAS MT subclones clear over time or simply fall below detection thresholds. The study analyzed ctDNA samples from 12 RAS-MT mCRC patients at four key time points: before treatment (T0), after four months of therapy (T1), at first progression (T2), and at second progression (T3). A confirmatory DNA

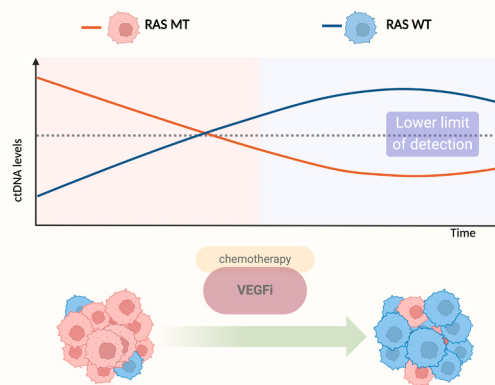


Fig. 2. The “RAS Dynamic Model”. Rather than an independent phenomenon, the NeoRAS WT concept reflects a broader dynamic model, where RAS MT subclones undergo negative selection in response to treatment. This plasmatic clearance, known as the RAS WT window, could offer new therapeutic opportunities with EGFR inhibitors. Abbreviations ctDNA, Circulating Tumor DNA; RAS WT, RAS Wild Type; RAS MT, RAS Mutated; VEGFI Vascular Endothelial Growth Factor inhibitor. Image created with BioRender.com, accessed on 28 August 2024.

Table 1

Summary of trials investigating the NeoRAS WT phenomenon.

	Study type	N patients enrolled	N NeoRAS WT patients (% NeoRAS WT)	Methods	N patients treated with anti-EGFR moAbs	Treatment	PFS	OS	ORR	DCR
Nicolazzo et al., 2020	Prospective	12	7(58 %)	rtPCR Idylla™ + Methylation test	-	-	-	-	-	-
Moati et al., 2020	Prospective	36	2 (5.5 %)	ddPCR, NGS, methylation detection of <i>WIF</i> and <i>NPY</i> genes by ddPCR	-	-	-	-	-	-
Klein-Scory et al., 2020	Prospective	12	10 (91 %)	ddPCR and BEAMing PCR	-	-	-	-	-	-
Garcia De Santiago et al., 2021	Retrospective	23	17 (74 %)	PCR assay: Idylla™ ctKRAS Mutation Assay and Idylla™ct <i>NRAS/BRAF</i> Mutation Assay	-	-	-	-	-	-
Nicolazzo et al., 2023	Prospective	70	42 (60 %)	rtPCR Idylla™ + Methylation test or NGS	-	-	15 mo	42 mo	-	-
Nicolazzo et al., 2021	Prospective	40	14 (35 %)	rtPCR Idylla™ + NGS Oncomine + Methylation test	10	anti-EGFR monotherapy	6,5 mo	-	-	-
Bouchahda et al., 2020	Prospective	16	9 (56 %)	Massarray UltraSeek technology	9	FOLFIRI + cetuximab	9 mo	-	55.6 %	77.8 %
Osumi et al., 2023	Prospective	107	23 (21.5 %)	OncoBEAM™ RAS CRC assay	23	anti-EGFR monotherapy	-	-	-	-
Osumi et al., 2024	Prospective	429	42 (9.8 %)	NGS	6	anti-EGFR monotherapy (N = 2) and chemotherapy+ anti-EGFR (N = 4)	-	-	50 %	20 %
JACCRO CC-1159	Prospective	29	24 (82.2 %)	TaqMan® PCR (castPCR) assays	-	FOLFOXIRI + bevacizumab	-	27mo	-	-
Nicolazzo et al., 2022	Prospective	72	2 (22 %) CT; 41 (82 %) CT+bevacizumab	rtPCR Idylla™ + Methylation test or NGS	-	CT (N = 22) or CT+Bevacizumab (N = 50)	5.9 mo CT; 9.3 mo CT+bevacizumab	-	-	-

Abbreviations EGFR mAb, anti-epidermal growth factor receptor monoclonal antibodies; PFS, Progression Free Survival; OS, Overall Survival; ORR, Objective Response Rate; DCR, Disease Control Rate; NGS, next-generation sequencing; mo, months; ddPCR, droplet-based digital polymerase chain reaction; WIF, WNT-Inhibitory-Factor 1; NPY, Neuropeptide Y; PCR, polymerase chain reaction; rtPCR, Real time PCR; CRC, colon rectal cancer.

methylation test was performed to assess ctDNA presence. At T1, 83 % of patients appeared RAS WT, but the absence of methylated ctDNA suggested that this was due to undetectable ctDNA rather than a biological clearance. By T2, the same 83 % remained RAS WT, but 62.5 % had methylated ctDNA, supporting the hypothesis that in many cases, the mutation had truly disappeared. At T3, 50 % of wild-type cases still had methylated ctDNA, while others, particularly those with lung or brain metastases, had none, likely due to lower ctDNA shedding from these tumor sites. This study highlights the utility of a five-gene methylation test in confirming ctDNA presence (Nicolazzo et al., 2020).

Moati et al. selected 36 patients from the PLACOL study (Nicolazzo et al., 2020) who were diagnosed with RAS MT mCRC through both histology and ctDNA. By performing LB at multiple time points, they analyzed the behavior of RAS MT subclones in response to treatment. RAS mutation clearance was considered valid only after ctDNA persistence confirmation, with NGS-based research on additional gene mutations or ddPCR-based detection of WNT-Inhibitory-Factor 1 (*WIF1*) and Neuropeptide Y (*NPY*) gene methylation. At progression, RAS mutation clearance was observed in 8/36 patients (22.2 %). However, only 2 of

these cases were confirmed using the designated molecular assays (Moati et al., 2020).

A similar strategy was adopted by Nicolazzo et al., using both NGS-mutational panel and PCR-based methylation profiling to confirm the presence of ctDNA after RAS MT clearance detection. The authors prospectively collected data from 40 RAS MT mCRC patients at PD from any line of therapy. Of these, 18/40 cases (45 %) were defined as “RAS converters” since no *RAS/BRAF* ctDNA mutations were found. These plasma samples were then analyzed with the NGS panel, and in 11/18 cases (61 %), at least one somatic mutation other than RAS was identified, classifying them as the “true RAS converters” subgroup. This result was confirmed in 14/18 patients by the methylation assay, showing a concordance of approximately 70 % between the two techniques. Considering the whole population, the authors noted a trend towards improved PFS in patients receiving EGFR blockade as second or further treatment lines compared with those who did not (Nicolazzo et al., 2021).

To address the risk of false-negative results in non-shedding tumors, Klein-Scory et al. employed the more accurate BEAMing PCR assay, as a

confirmatory test for RAS MT ctDNA detected with ddPCR at screening. The authors selected 12 RAS MT mCRC patients who achieved stable disease (SD) or partial response (PR) during first-line therapy. A reduction in plasmatic RAS mutational allele frequency (MAF) was observed in all 12 cases (100 %). In 10/12 patients (91 %), ctDNA-based RAS mutational load dropped below the ddPCR detection threshold ($<0.25\%$), which was interpreted as NeoRAS WT conversion ($p < 0.0001$). Reversion time was not significantly associated with either the tumor's sidedness ($p = 0.955$) or treatment regimen ($p = 0.266$) (Klein-Scory et al., 2020).

Moreover, a high NeoRAS WT conversion rate was reported by García De Santiago et al. in a retrospective analysis of a LB series of 23 RAS MT mCRC patients treated with anti-VEGF therapies. Through PCR assays (i.e., Idylla™ ctKRAS Mutation Assay and Idylla™ct NRAS/BRAF Mutation Assay), the authors observed the disappearance of RAS MT in up to 17/23 patients (73.9 %) (De Santiago et al., 2021).

In 2023, Nicolazzo and colleagues published a prospective trial enrolling 70 patients with RAS MT mCRC refractory to first-line treatment. At first, the authors distinguished the 'NeoRAS WT' (45/70, 64 %) from the 'RAS MT' (25/70, 36 %) population based on plasma ctDNA sampling at PD. To confirm RAS clearance, a sequential validation approach was used: a primary NGS analysis identified at least one additional somatic mutation in 36/45 cases (80 %), while in the remaining 9/45 (20 %) a methylation assay was performed, yielding a positive result in 6/9 cases (67 %). By combining these two methods, a 'true RAS conversion' was confirmed in 42/45 (93 %) samples. Finally, the authors highlighted the prognostic impact of RAS mutation clearance, with both median OS and post-progression survival favoring NeoRAS WT patients over the RAS MT persistent group (64 vs. 27 months, $p < 0.001$, and 42 vs. 16 months, $p < 0.001$, respectively) (Nicolazzo et al., 2023).

The subsequent research delved deeper into the clinical implications of NeoRAS WT patients, focusing on the predictive and prognostic value of RAS conversion and clarifying on the impact of anti-EGFR therapies in this context.

Bouchahda et al. conducted a study enrolling 16 pre-treated, anti-EGFR-naïve mCRC patients harboring RAS mutations at baseline. At disease progression, LB was performed to classify patients into two distinct groups: RAS MT persistent (Group 1, $N = 7$) and RAS WT converted (Group 2, $N = 9$). All patients in Group 2 were treated with the cetuximab + FOLFIRI regimen, while Group 1 received the investigator's choice of therapy. Anti-EGFR treatment was associated with significantly improved outcomes: mPFS was 3.5 months (95 % CI, 0.8–6.1) for Group 1 compared with 9.0 months (95 % CI, 4.7–13.3) for Group 2. Similarly, mOS was 4.7 months (95 % CI, 1.1–8.3) in Group 1, whereas it was not reached at 18.7 months in Group 2. ORR was 42.9 % and DCR 85.7 % in Group 1, while in Group 2, ORR reached 55.6 % and DCR 77.8 % (Bouchahda et al., 2020).

Osumi et al. later expanded on this topic, conducting two key studies. The first was a multi-institutional, observational study analyzing clinicopathological features associated with ctDNA-defined RAS conversion among 107 mCRC patients, refractory to prior chemotherapies. Using BEAMing PCR (OncoBEAM™ RAS CRC assay), 23/107 (21.5 %) patients were identified as NeoRAS WT. In the logistic regression multivariate analysis, factors significantly associated with NeoRAS WT conversion were the absence of liver metastases ($p = 0.019$), smaller tumor diameter ($p = 0.012$), and non KRAS exon 2 mutations ($p = 0.026$). Anti-EGFR monotherapy was attempted only in 3/23 cases, showing poor results: one patient had decreased tumor marker levels but discontinued due to skin toxicity, while for the other two patients mPFS was 2 months (Osumi et al., 2023). Building upon prior findings, some of the same authors investigated a subgroup of 478 tissue-diagnosed RAS MT mCRC patients from the GOZILA study. Plasmatic KRAS and NRAS mutations were detected in 387/478 cases via ctDNA sampling, while the remaining 91 patients (19.0 %) were classified as NeoRAS WT. This population was further divided into two subgroups: Group A, with no

additional detectable mutations, and Group B, carrying other non-clonal somatic alterations (e.g., TP53, ATM, and GNAS). For Group A, the multivariate analysis showed a significant association of NeoRAS conversions with the absence of liver ($p < 0.00001$), and lymph node ($p = 0.033$) metastases. Similarly, in Group B, NeoRAS WT status was significantly correlated with the absence of liver metastasis ($p = 0.0005$) and non KRAS exon 2 mutations ($p = 0.049$). As for the clinical outcome, 6 NeoRAS WT patients were treated with anti-EGFR, ranging from fourth to seventh line. Among them, one achieved PR, while two maintained SD for at least six months (Osumi et al., 2024).

Further prognostic implications were explored by Sunakawa et al., drawing data from the Japanese phase II JACCRO CC-11 trial, where 64 RAS MT patients received modified FOLFOXIRI+bevacizumab as first-line treatment. CtDNA analysis was carried out at three time points in 36 patients: at baseline, week 8, and PD. RAS mutations disappeared in 24/29 (82.8 %) cases at 8 weeks and remained undetectable at PD in 18/29 (62.1 %) cases. Moreover, NeoRAS conversions showed significantly improved clinical outcomes, with mOS of 27 vs 16 months (HR 0.31, $p = 0.0073$) and post-progression survival of 15.1 vs 7.3 months (HR 0.21, $p = 0.0046$). Of note, the NeoRAS WT conversion was associated with a significantly lower tumor burden which could have influenced ctDNA shedding and RAS MT detection (Sunakawa et al., 2022).

The group led by Nicolazzo et al. investigated the role of bevacizumab in RAS conversion in a cohort of 72 patients with unresectable RAS MT mCRC, whose baseline plasma ctDNA mutational status was concordant with that of the primary tumor tissue. In the overall population, 43 cases (60 %) showed NeoRAS WT conversion, with 88 % maintaining it across all time points. Bevacizumab was identified as the only independent factor associated with RAS conversion (OR = 45, 95 % CI 9–230). Among patients receiving chemotherapy alone, RAS conversion was rare (2/22, 9 %), whereas in those treated with chemotherapy plus bevacizumab, 41/50 (82 %) converted to RAS WT. Moreover, patients who lost their RAS mutation had significantly better outcomes than those with persistent RAS mutations in the bevacizumab-treated group (mPFS 9.3 vs. 5.9 months, $p = 0.001$), supporting RAS conversion as a potential positive prognostic marker for treatment response (Nicolazzo et al., 2022).

3. NeoRAS WT mCRC: ongoing trials

Numerous trials are currently underway to prospectively assess the effectiveness of anti-EGFR in the context of LB-defined NeoRAS WT mCRC. These studies vary in terms of design, treatment regimens, and endpoints (Table 2).

The single-arm phase II CETIDYL trial (NCT04189055) is investigating the safety and efficacy of cetuximab vs cetuximab+irinotecan for 107 NeoRAS WT patients, as third or further treatment lines. The study focuses on a very selected population with liver-limited advanced disease.

The phase II randomized MoLiMoR trial (NCT04554836) is assessing the efficacy and safety of FOLFIRI +/- intermittent cetuximab vs FOLFIRI as first-line setting in left-sided mCRC. The study aims to enroll 144 patients, with PFS as the primary endpoint. Unlike other trials, MoLiMoR incorporates real-time molecular monitoring to guide treatment decisions. In the experimental arm, anti-EGFR treatment is either halted or resumed according to the plasma dynamics of RAS MT subclones, detected by ctDNA ddPCR analysis.

The Japanese C-PROWESS single-arm phase II trial (jRCTs031210565) employs BEAMing PCR technology to identify RAS WT mCRC patients at screening. The study is evaluating irinotecan+panitumumab as a third or further-line regimen. The primary endpoint is ORR, with the target enrollment of 30 patients.

The CONVERTIX (2017–003242–25) study was a single-arm, phase II study attempting to assess the efficacy of panitumumab+FOLFIRI as second-line treatment after receiving first-line FOLFOX/CAPOX+bevacizumab. The primary endpoint was PFS, but the trial was

Table 2

Summary of Ongoing Clinical trials assessing NeoRAS WT phenomenon.

Study	Phase	Method	Status	Line of treatment	Estimated N	Primary endpoint	Treatment regimen
CETIDYL trial (NCT04189055)	II	rtPCR	Recruiting	Third or further line	72	ORR	Cetuximab or Cetuximab + Irinotecan
MoLiMoR (NCT04554836)	II	ddPCR	Active, not recruiting	First line	144	PFS	FOLFIRI + intermittent Cetuximab vs FOLFIRI
KAIROS (EudraCT number 2019-001328-36)	II	rtPCR		Second line	112	ORR	Chemotherapy + Cetuximab
C-PROWESS (JRCTs031210565)	II	BEAMing PCR	Recruiting	Third or further line	30	ORR	Cetuximab + Irinotecan
CONVERTIX (2017-003242-25)	II	BEAMing PCR	Closed	Second line	40	PFS	FOLFIRI + Panitumumab
NEORAS-SYSUCC (NCT05962502)	II	NA	Recruiting	Third or further line	54	ORR	Cetuximab + Irinotecan

Abbreviations PFS, Progression Free Survival; ORR, Objective Response Rate; rtPCR, Real time PCR; ddPCR, droplet-based digital polymerase chain reaction; PCR, polymerase chain reaction.

prematurely closed due to insufficient accrual.

NEORAS-SYSUCC (NCT05962502) is a Chinese phase II single-arm trial, testing the efficacy and safety of cetuximab+irinotecan as third-line treatment for NeoRAS WT left-sided mCRC. The primary endpoint is ORR, with a target enrollment of 54 patients. Recruitment began in August 2023.

Finally, the KAIROS study (EudraCT number 2019-001328-36) another single-arm phase II trial, faced early termination due to poor accrual. Designed to assess the efficacy and safety of cetuximab + a chemotherapy doublet as second-line treatment for 112 RAS WT converted mCRC patients, its primary endpoint was ORR.

4. Discussion

Current international guidelines base the mCRC therapeutic algorithm on a dichotomous model, historically shaped by key determinants such as tumor sidedness and mutational profiling (Cervantes et al., 2023; Benson et al., 2024). However, the introduction of LB is reshaping this paradigm, better aligning with the dynamic nature of mCRC, where mutant subclones expand or decline in response to treatment pressure and microenvironmental changes. As described in this article, RAS MT subclones exhibit a pulsatile behavior, undergoing exponential time-dependent decay, not only in baseline RAS WT patients but also in those initially RAS MT, defining the NeoRAS WT phenomenon. This evolving perspective suggests that EGFR blockade could be guided by an LB-defined RAS WT therapeutic window rather than relying solely on baseline tissue genotyping. Supporting this concept, recent data from the FIRE-4 trial showed how LB detected RAS mutations in 13 % of the RAS WT enrolled population. In these cases, switching from anti-EGFR to anti-VEGF therapy resulted in numerically improved outcomes (PFS 10.1 vs. 6.4 m, HR 0.82 (95 % CI 0.47–1.41), $p = 0.54$; OS 24.9 vs. 16.3 mo, HR, 0.57 (95 % CI 0.31–1.07), $p = 0.10$) compared with continuous anti-EGFR therapy, further highlighting the role of LB in addressing mCRC's molecular heterogeneity (Stintzing et al., 2025). The NeoRAS WT phenomenon could therefore be considered part of a broader concept, the "RAS mutation dynamic model," where the tumor's molecular status is tightly linked to the shifting predominance of different subclonal populations.

However, several key limitations must be addressed before considering a change in clinical practice. First, the NeoRAS WT concept lacks a solid biological rationale, as the underlying mechanisms driving RAS conversions remain poorly understood. Current theories suggest that selective treatment pressure, particularly anti-angiogenic therapy, induces tumor microenvironment changes (Osumi et al., 2023). Several studies have hypothesized that bevacizumab-induced hypoxic damage induces a massive and deadly increase of intracellular ROS production, exceeding the high scavenging properties of RAS MT cells (Henry et al.,

2020; Osumi et al., 2023; Lim and Leprivier, 2019; Bartolacci et al., 2021). An additional challenge lies in the lack of standardization in LB diagnostics. The different NeoRAS WT incidences reported across studies stem from inconsistent designs, sample sizes, and, most notably, plasma diagnostic methodologies. Despite the known differences in sensitivity and specificity, both PCR-based and NGS-based detecting techniques have been used across trials, leading to discrepancies in results (Nicolazzo et al., 2022, 2020; Raimondi et al., 2019). Furthermore, diagnostic accuracy is affected by ctDNA shedding variability, which depends on factors such as tumor burden and metastatic sites (Bouchahda et al., 2020; Osumi et al., 2021). Notably, higher ctDNA clearance has been observed in non-liver metastases (e.g., lung, brain, peritoneum), likely due to the presence of biological barriers and increased circulating DNase activity (Uhlén et al., 2015). Similarly, NeoRAS WT conversion correlates with fewer metastatic sites and greater tumor shrinkage, but a reduction in tumor volume alone does not necessarily reflect true RAS MT clearance. False-negative results may occur when ctDNA levels drop below detection thresholds (Bando et al., 2019). To address false negative results for low shedding tumors, several studies adopted confirmatory tests, such as NGS-based screening for truncal APC/TP53 mutations or methylation profiling of WIF1/NPY genes (Klein-Scory et al., 2020; Henry et al., 2020; Sunakawa et al., 2020; Osumi et al., 2023; Nicolazzo et al., 2023). These methods help differentiate true RAS clearance from undetectable ctDNA, refining the actual incidence of NeoRAS WT mCRC. By applying these strategies, Osumi et al. reported a decrease in RAS conversion incidence from 19 % to 9.8 %, while a post-hoc CO.26 trial analysis registered a 50 % reduction (Osumi et al., 2021; Wu et al., 2024). The same authors have highlighted the scarce durability of RAS conversions, with the reacquisition of the baseline RAS mutations within a median time of 162.5 days (Wu et al., 2024). This raises another key debate: the optimal timing for ctDNA sampling. While LB is typically performed at disease progression, the decision to rely solely on plasma analysis or integrate tissue biopsy is challenging. Plasma testing is favored for its non-invasiveness, but in low-shedding tumors, tissue biopsy may be necessary to distinguish true RAS clearance from false negatives. In a heterogeneous disease like mCRC, combining LB with tissue biopsy may offer a more comprehensive approach, but it also raises significant cost-related barriers. High-sensitivity NGS-based ctDNA testing remains expensive, and many centers lack the necessary infrastructure and expertise for reliable real-time molecular monitoring. Without standardized reimbursement policies, its widespread implementation remains largely restricted to clinical trials, thereby limiting broader accessibility. As a final point, despite suggesting a better prognostic potential, the predictive value of RAS MT clearance remains unclear. The transient nature of this phenomenon, the small sample sizes examined, and the inconsistency in diagnostic methods have prevented

a definitive conclusion in the evaluation of the anti-EGFR treatments in this setting (Bouchahda et al., 2020; Henry et al., 2020).

5. Conclusions

While the NeoRAS WT concept holds promise and could redefine treatment strategies in mCRC, its clinical utility needs to be validated through prospective, randomized trials. Addressing current limitations, improving standardization of testing, and ensuring cost-effective implementation will represent crucial steps toward the integration of this approach into routine clinical practice.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

CRediT authorship contribution statement

Giulia Massaro: Conceptualization, Data curation, Formal analysis, Writing – original draft, Writing – review & editing.

Jacopo Venturini: Conceptualization, Data curation, Formal analysis, Writing – original draft, Writing – review & editing.

Daniele Rossini: Analysis, Conceptualization, Data curation, Investigation, Methodology, Supervision, Visualization, Writing – review & editing, Validation.

Agnes Vannini: Data curation, Formal analysis, Writing – original draft.

Marco Brugia: Data curation, Formal analysis, Writing – original draft.

Daniele Lavacchi: Data curation, Formal analysis, Writing – original draft.

Cristiana Conticello: Data curation, Formal analysis, Writing – original draft.

Irene Valle: Data curation, Formal analysis, Writing – original draft.

Delia Ravizza: Data curation, Formal analysis, Writing – original draft.

Serena Pillozzi: Data curation, Formal analysis, Writing – original draft.

Lorenzo Antonuzzo: Conceptualization, Data curation, Formal analysis, Validation.

Declaration of Competing Interest

D.R. received honoraria from Amgen, MSD and Takeda. The other authors declare no competing interests

References

- Bando, H., et al., 2019. A multicentre, prospective study of plasma circulating tumour DNA test for detecting RAS mutation in patients with metastatic colorectal cancer. *Br. J. Cancer* 120 (10). <https://doi.org/10.1038/s41416-019-0457-y>.
- Bartolacci, C., Andreani, C., El-Gammal, Y., Scaglioni, P.P., 2021. Lipid metabolism regulates oxidative stress and ferroptosis in RAS-driven cancers: A perspective on cancer progression and therapy. *Front Mol. Biosci.* <https://doi.org/10.3389/fmolb.2021.706650>.
- Benson, A.B., et al., 2024. Colon Cancer, Version 3.2024, NCCN Clinical Practice Guidelines in Oncology. J. Natl. Compr. Cancer Netw. <https://doi.org/10.6004/jnccn.2024.0029>. PMID: 38862008.
- Bokemeyer, C., et al., 2015. FOLFOX4 plus cetuximab treatment and RAS mutations in colorectal cancer. *Eur. J. Cancer* 51 (10). <https://doi.org/10.1016/j.ejca.2015.04.007>.
- Bouchahda, M., et al., 2020. Undetectable RAS-mutant clones in plasma: Possible implication for anti-EGFR therapy and prognosis in patients with RAS-mutant metastatic colorectal cancer. *JCO Precis Oncol.* (4). <https://doi.org/10.1200/po.19.00400>.
- Cervantes, A., et al., 2023. Metastatic colorectal cancer: ESMO Clinical Practice Guideline for diagnosis, treatment, and follow-up. *Ann. Oncol.* 34 (1). <https://doi.org/10.1016/j.annonc.2022.10.003>.
- Chan, D.K.H., Buczac, S.J.A., 2021. Tumour heterogeneity and evolutionary dynamics in colorectal cancer. *Oncogenesis*. <https://doi.org/10.1038/s41389-021-00342-x>.
- Chung, D.C., et al., 2024. A cell-free DNA blood-based test for colorectal cancer screening. *N. Engl. J. Med.* 390 (11). <https://doi.org/10.1056/nejmoa2304714>.
- Ciardello, D., et al., 2021. Biomarker-guided anti-EGFR rechallenge therapy in metastatic colorectal cancer. *Cancers (Basel)*. <https://doi.org/10.3390/cancers13081941>.
- Cremolini, C., et al., 2019. Rechallenge for patients with RAS and BRAF wild-type metastatic colorectal Cancer with acquired resistance to first-line cetuximab and irinotecan: A phase 2 single-arm clinical trial. *JAMA Oncol.* 5 (3). <https://doi.org/10.1001/jamaoncol.2018.5080>.
- Dagogo-Jack, I., Shaw, A.T., 2018. Tumour heterogeneity and resistance to cancer therapies. *Nat. Rev. Clin. Oncol.* <https://doi.org/10.1038/nrclinonc.2017.166>.
- Dasari, A., et al., 2023. "Fruquintinib versus placebo in patients with refractory metastatic colorectal cancer (FRESCO-2): an international, multicentre, Random, Double-Blind, phase 3 Study," *Lancet* 402 (10395). [https://doi.org/10.1016/S0140-6736\(23\)00772-9](https://doi.org/10.1016/S0140-6736(23)00772-9).
- De Santiago, B.G., et al., 2021. RAS mutational status in advanced colorectal adenocarcinoma treated with anti-angiogenics: Preliminary experience with liquid biopsy. *Vivo (Brooklyn)* 35 (5). <https://doi.org/10.21873/INVIVO.12571>.
- Diaz, L.A., Bardelli, A., 2014. Liquid biopsies: Genotyping circulating tumor DNA. *J. Clin. Oncol.* <https://doi.org/10.1200/JCO.2012.45.2011>.
- Diehl, F., et al., 2008. Circulating mutant DNA to assess tumor dynamics. *Nat. Med.* 14 (9). <https://doi.org/10.1038/nm.1789>.
- Douillard, J.-Y., et al., 2013. Panitumumab-FOLFOX4 treatment and RAS mutations in colorectal cancer. *N. Engl. J. Med.* 369 (11). <https://doi.org/10.1056/NEJMoa1305275>.
- Epistolio, S., et al., 2021. Occurrence of RAS reversion in metastatic colorectal cancer patients treated with bevacizumab. *Oncotarget* 12 (11). <https://doi.org/10.18632/oncotarget.27965>.
- Findlay, J.M., et al., 2016. Differential clonal evolution in oesophageal cancers in response to neo-adjuvant chemotherapy. *Nat. Commun.* 7. <https://doi.org/10.1038/ncomms11111>.
- Gazzaniga, P., Raimondi, C., Urbano, F., Cortesi, E., 2018. EGFR inhibitor as second-line therapy in a Patient with mutant RAS metastatic colorectal cancer: Circulating tumor DNA to personalize treatment. *JCO Precis Oncol.* (2). <https://doi.org/10.1200/po.17.00277>.
- Gerlinger, M., et al., 2012. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N. Engl. J. Med.* 366 (10). <https://doi.org/10.1056/nejmoa1113205>.
- Grothey, A., et al., 2013. Regorafenib monotherapy for previously treated metastatic colorectal cancer (CORRECT): An international, multicentre, randomised, placebo-controlled, phase 3 trial. *Lancet* 381 (9863). [https://doi.org/10.1016/S0140-6736\(12\)61900-X](https://doi.org/10.1016/S0140-6736(12)61900-X).
- Henriksen, T.V.V., et al., 2021. "Serial circulating tumor DNA analysis to assess recurrence risk, benefit of adjuvant therapy, growth rate, and early relapse detection in stage III colorectal cancer patients. *J. Clin. Oncol.* 39 (15). https://doi.org/10.1200/jco.2021.39.15_suppl.3540.
- Henry, J., et al., 2020. NeoRAS: Incidence of RAS reversion from RAS mutated to RAS wild type. *J. Clin. Oncol.* 38 (4). https://doi.org/10.1200/jco.2020.38.4_suppl.180.
- Kagawa, Y., et al., 2022. Plasma RAS dynamics and anti-EGFR rechallenge efficacy in patients with RAS/BRAF wild-type metastatic colorectal cancer: REMARRY and PURSUIT trials. *J. Clin. Oncol.* https://doi.org/10.1200/jco.2022.40.16_suppl.3518.
- Kim, Y., et al., 2016. Targeted proteomics identifies liquid-biopsy signatures for extracapsular prostate cancer. *Nat. Commun.* 7. <https://doi.org/10.1038/ncomms11906>.
- Klein-Scory, S., et al., 2020. Evolution of RAS mutational status in liquid biopsies during first-line chemotherapy for metastatic colorectal cancer. *Front Oncol.* 10. <https://doi.org/10.3389/fonc.2020.01115>.
- Krebs, M.G., et al., 2022. Practical considerations for the use of circulating tumor DNA in the treatment of patients with cancer: A narrative review. *JAMA Oncol.* <https://doi.org/10.1001/jamaoncol.2022.4457>.
- Lenz, H., et al., 2014. CALGB/SWOG 80405: Phase III Trial of Irinotecan/5-FU/Leucovorin (FOLFIRI) or Oxaliplatin/5-FU/Leucovorin (mFOLFOX6) with Bevacizumab (BV) or Cetuximab (Cet) for Patients (Pts) with Expanded RAS Analyses of Untreated Metastatic Adenocarcinoma of the Colon or Rectum (mCRC). *Ann. Oncol.* 25. <https://doi.org/10.1093/annonc/mdu438.13>.
- Lim, J.K.M., Lepriver, G., 2019. The impact of oncogenic RAS on redox balance and implications for cancer development. *Cell Death Dis.* <https://doi.org/10.1038/s41419-019-2192-y>.
- Lonardi, S., et al., 2020. The PEGASUS trial: Post-surgical liquid biopsy-guided treatment of stage III and high risk stage II colon cancer patients (suppl). *J. Clin. Oncol.* 38 (15). doi:10.1200/jco.2020.38.15_suppl.35124.
- Lone, S.N., et al., 2022. Liquid biopsy: A step closer to transforming diagnosis, prognosis, and future of cancer treatments. *Mol. Cancer*. <https://doi.org/10.1186/s12943-022-01543-7>.
- Loree, J.M., et al., 2021. Serial circulating tumor DNA (ctDNA) monitoring in metastatic colorectal cancer (mCRC) reveals dynamic profile of actionable alterations. *J. Clin. Oncol.* 39 (15). https://doi.org/10.1200/jco.2021.39.15_suppl.3572.
- Malla, M., Loree, J.M., Kasi, P.M., Parikh, A.R., 2022. Using circulating tumor DNA in colorectal cancer: Current and evolving practices. *J. Clin. Oncol.* <https://doi.org/10.1200/JCO.21.02615>.
- Martinelli, E., et al., 2020. Implementing anti-epidermal growth factor receptor (EGFR) therapy in metastatic colorectal cancer: Challenges and future perspectives. *Ann. Oncol.* <https://doi.org/10.1016/j.annonc.2019.10.007>.
- Martinelli, E., et al., 2021. Cetuximab Rechallenge Plus Avelumab in Pretreated Patients with RAS Wild-type Metastatic Colorectal Cancer: The Phase 2 Single-Arm Clinical CAVE Trial. *JAMA Oncol.* 7 (10). <https://doi.org/10.1001/jamaoncol.2021.2915>.

- Mauri, G., et al., 2022. Liquid biopsies to monitor and direct cancer treatment in colorectal cancer. *Br. J. Cancer*. <https://doi.org/10.1038/s41416-022-01769-8>.
- Mayer, R.J., et al., 2015. Randomized Trial of TAS-102 for Refractory Metastatic Colorectal Cancer. *N. Engl. J. Med.* 372 (20). <https://doi.org/10.1056/nejmoa1414325>.
- Mayers, J.R., et al., 2014. Elevation of circulating branched-chain amino acids is an early event in human pancreatic adenocarcinoma development. *Nat. Med.* 20 (10). <https://doi.org/10.1038/nm.3686>.
- Meric-Bernstam, F., et al., 2019. Pertuzumab plus trastuzumab for HER2-amplified metastatic colorectal cancer (MyPathway): An updated report from a multicentre, open-label, phase 2a, multiple basket study. *Lancet Oncol.* 20 (4). [https://doi.org/10.1016/S1470-2045\(18\)30904-5](https://doi.org/10.1016/S1470-2045(18)30904-5).
- Misale, S., et al., 2012. Emergence of KRAS mutations and acquired resistance to anti-EGFR therapy in colorectal cancer. *Nature* 486 (7404). <https://doi.org/10.1038/nature11156>.
- Moati, E., et al., 2020. Plasma clearance of RAS mutation under therapeutic pressure is a rare event in metastatic colorectal cancer. *Int. J. Cancer* 147 (4). <https://doi.org/10.1002/ijc.32657>.
- Morelli, M.P., et al., 2015. Characterizing the patterns of clonal selection in circulating tumor DNA from patients with colorectal cancer refractory to anti-EGFR treatment. *Ann. Oncol.* 26 (4). <https://doi.org/10.1093/annonc/mdv005>.
- Nakamura, Y., et al., 2019. TRIUMPH: Primary efficacy of a phase II trial of trastuzumab (T) and pertuzumab (P) in patients (pts) with metastatic colorectal cancer (mCRC) with HER2 (ERBB2) amplification (amp) in tumour tissue or circulating tumour DNA (ctDNA): A GOZILA sub-study. *Ann. Oncol.* 30. <https://doi.org/10.1093/annonc/mdz246.004>.
- Nakamura, Y., et al., 2020. Clinical utility of circulating tumor DNA sequencing in advanced gastrointestinal cancer: SCRUM-Japan GI-SCREEN and GOZILA studies. *Nat. Med.* 26 (12). <https://doi.org/10.1038/s41591-020-1063-5>.
- Nicolazzo, C., et al., 2020. Circulating methylated DNA to monitor the dynamics of RAS mutation clearance in plasma from metastatic colorectal cancer patients. *Cancers (Basel)* 12 (12). <https://doi.org/10.3390/cancers12123633>.
- Nicolazzo, C., et al., 2021. True conversions from RAS mutant to RAS wild-type in circulating tumor DNA from metastatic colorectal cancer patients as assessed by methylation and mutational signature. *Cancer Lett.* Vol. 507. <https://doi.org/10.1016/j.canlet.2021.03.014>.
- Nicolazzo, C., et al., 2022. RAS mutation conversion in bevacizumab-treated metastatic colorectal cancer patients: A liquid biopsy-based study. *Cancers (Basel)* 14 (3). <https://doi.org/10.3390/cancers14030802>.
- Nicolazzo, C., et al., 2023. Genomic landscape and survival analysis of ctDNA 'neo-RAS wild-type' patients with originally RAS mutant metastatic colorectal cancer. *Front Oncol.* 13. <https://doi.org/10.3389/fonc.2023.1160673>.
- Oki, E., et al., 2023. Circulating tumor DNA dynamics as an early predictor of recurrence in patients with radically resected colorectal cancer: Updated results from GALAXY study in the CIRCULATE-Japan. *J. Clin. Oncol.* 41 (16). <https://doi.org/10.1200/jco.2023.41.16.suppl.3521>.
- Osumi, H., et al., 2021. NeORAS wild-type in metastatic colorectal cancer: Myth or truth?—Case series and review of the literature. *Eur. J. Cancer* 153. <https://doi.org/10.1016/j.ejca.2021.05.010>.
- Osumi, H., et al., 2023. A multi-institutional observational study evaluating the incidence and the clinicopathological characteristics of NeORAS wild-type metastatic colorectal cancer. *Transl. Oncol.* 35. <https://doi.org/10.1016/j.tranon.2023.101718>.
- Osumi, H., et al., Dec. 2024. Clinical features associated with NeORAS wild-type metastatic colorectal cancer: A SCRUM-Japan GOZILA substudy. *Nat. Commun.* 15 (1), 500264. <https://doi.org/10.1038/s41467-024>.
- Overman, M.J., et al., 2013. Use of research biopsies in clinical trials: Are risks and benefits adequately discussed? *J. Clin. Oncol.* 31 (1). <https://doi.org/10.1200/JCO.2012.43.1718>.
- Parseghian, C.M., et al., 2019. Anti-EGFR-resistant clones decay exponentially after progression: Implications for anti-EGFR re-challenge. *Ann. Oncol.* 30 (2). <https://doi.org/10.1093/annonc/mdy509>.
- Peeters, M., et al., 2015. Prevalence of RAS mutations and individual variation patterns among patients with metastatic colorectal cancer: A pooled analysis of randomised controlled trials. *Eur. J. Cancer* 51 (13). <https://doi.org/10.1016/j.ejca.2015.05.017>.
- Porru, M., et al., 2018. Targeting KRAS in metastatic colorectal cancer: current strategies and emerging opportunities. *J. Exp. Clin. Cancer Res.* 37 (1). <https://doi.org/10.1186/s13046-018-0719-1>.
- Prager, G.W., et al., 2023. Trifluridine-Tipiracil and Bevacizumab in Refractory Metastatic Colorectal Cancer. *N. Engl. J. Med.* 388 (18). <https://doi.org/10.1056/nejmoa2214963>.
- Raimondi, C., et al., 2019. Transient disappearance of RAS mutant clones in plasma: A counterintuitive Clinical use of EGFR inhibitors in RAS mutant metastatic colorectal cancer. *Cancers (Basel)* 11 (1). <https://doi.org/10.3390/cancers11010042>.
- Reinert, T., et al., 2019. Analysis of plasma cell-free DNA by ultradeep sequencing in patients with stages I to III colorectal cancer. *JAMA Oncol.* 5 (8). <https://doi.org/10.1001/jamaoncol.2019.0528>.
- Rolfo, C., et al., 2020. Challenges and opportunities of cfDNA analysis implementation in clinical practice: Perspective of the International Society of Liquid Biopsy (ISLB). *Crit. Rev. Oncol. Hematol.* <https://doi.org/10.1016/j.critrevonc.2020.102978>.
- Sartore-Bianchi, A., et al., 2016. "Dual-targeted therapy with trastuzumab and lapatinib in treatment refractory, KRAS codon 12/13 wild-type, HER2-positive metastatic colorectal cancer (HERACLES): A proof-of-concept, multicentre, open-label, phase 2 trial." *Lancet Oncol.* 17 (6). [https://doi.org/10.1016/S14702045\(16\)00150-9](https://doi.org/10.1016/S14702045(16)00150-9).
- Sartore-Bianchi, A., et al., 2022. "Circulating tumor DNA to guide rechallenge with panitumumab in Metastatic colorectal cancer: The phase 2 CHRONOS trial." *Nat. Med.* 28 (8). <https://doi.org/10.1038/s41591-02201886-0>.
- Sartore-Bianchi, A., Marsoni, S., Siena, S., 2019. The amount of evidence needed to support ERBB2 as a biomarker for resistance to EGFR inhibitors in metastatic colorectal cancer. *JAMA Oncol.* <https://doi.org/10.1001/jamaoncol.2019.2982>.
- Sato, S., et al., 2022. Chemotherapy-induced reversion of mutant RAS to wild-type RAS in metastatic colorectal cancer. *Anticancer Res* 42 (5). <https://doi.org/10.21873/anticancer.15740>.
- Siena, S., et al., 2018a. Dynamic molecular analysis and clinical correlates of tumor evolution within a phase II trial of panitumumab-based therapy in metastatic colorectal cancer. *Ann. Oncol.* 29 (1). <https://doi.org/10.1093/annonc/mdx504>.
- Siena, S., et al., 2018b. Targeting the human epidermal growth factor receptor 2 (HER2) oncogene in Colorectal cancer. *Ann. Oncol.* <https://doi.org/10.1093/annonc/mdy100>.
- Siena, S., et al., 2021. 386O Exploratory biomarker analysis of DESTINY-CRC01, a phase II, multicenter, open label study of trastuzumab deruxtecan (T-DXd, DS-8201) in patients (pts) with HER2-expressing metastatic colorectal cancer (mCRC). *Ann. Oncol.* 32. <https://doi.org/10.1016/j.annonc.2021.08.908>.
- Siravegna, G., et al., 2019a. Plasma HER2 (ERBB2) copy number predicts response to HER2-targeted therapy in metastatic colorectal cancer. *Clin. Cancer Res.* 25 (10), 3389. <https://doi.org/10.1158/1078-0432.CCR-18>.
- Siravegna, G., et al., 2019b. How liquid biopsies can change clinical practice in oncology. *Ann. Oncol.* <https://doi.org/10.1093/annonc/mdz227>.
- Stintzing, S., et al., 2016. FOLFIRI plus cetuximab versus FOLFIRI plus bevacizumab for metastatic colorectal cancer (FIRE-3): a post-hoc analysis of tumour dynamics in the final RAS wild-type subgroup of this randomised open-label phase 3 trial. *Lancet Oncol.* Vol. 17, no. 10 2045 (16), 30269. <https://doi.org/10.1016/S1470.8>.
- Stintzing, S., et al., Feb. 2025. Baseline liquid biopsy in relation to tissue-based parameters in metastatic colorectal cancer: Results from the randomized FIRE-4 (AIO-KRK-0114) study. *JCO*. <https://doi.org/10.1200/JCO.24.01174>.
- Sunakawa, Y., et al., 2020. RAS mutations in circulating tumor DNA and clinical outcomes of rechallenge treatment with anti-EGFR antibodies in patients with metastatic colorectal cancer. *JCO Precis Oncol.* (4). <https://doi.org/10.1200/po.20.00109>.
- Sunakawa, Y., et al., 2022. Dynamic changes in RAS gene status in circulating tumour DNA: A phase II trial of first-line FOLFIRI plus bevacizumab for RAS-mutant metastatic colorectal cancer (JACCRO CC-11). *ESMO Open* 7 (3). <https://doi.org/10.1016/j.esmoop.2022.100512>.
- Tie, J., et al., 2019. Circulating tumor DNA analyses as markers of recurrence risk and benefit of adjuvant therapy for stage III colon cancer. *JAMA Oncol.* 5 (12). <https://doi.org/10.1001/jamaoncol.2019.3616>.
- Tie, J., et al., 2022. Circulating tumor DNA analysis guiding adjuvant therapy in stage II colon cancer. *N. Engl. J. Med.* 386 (24). <https://doi.org/10.1056/nejmoa2200075>.
- Uhlén, M., et al., 2015. Tissue-based map of the human proteome. *Science* 347 (6220). <https://doi.org/10.1126/science.1260419>.
- Venesio, T., Siravegna, G., Bardelli, A., Sapino, A., 2018. Liquid biopsies for monitoring temporal genomic heterogeneity in breast and colon cancers. *Pathobiology*. <https://doi.org/10.1159/000473882>.
- Venturini, J., et al., Sep. 2024. The emerging HER2 landscape in colorectal cancer: The key to unveil the future treatment algorithm?" *Crit. Rev. Oncol. Hematol.* <https://doi.org/10.1016/j.critrevonc.2024.104515>.
- Wu, F.T.H., Topham, J.T., et al., 2024. Kinetic profiling of RAS mutations with circulating tumor DNA in the Canadian Cancer Trials Group CO.26 trial suggests the loss of RAS mutations in Neo-RAS-wildtype metastatic colorectal cancer is transient. *J. Clin. Oncol.* <https://doi.org/10.1200/PO.24.00031>.
- Xin, L., Yue, Y., Zihan, R., Youbin, C., Tianyu, L., Rui, W., 2023. Clinical application of liquid biopsy based on circulating tumor DNA in non-small cell lung cancer. *Front Physiol.* <https://doi.org/10.3389/fphys.2023.1200124>.

Giulia Massaro, MD, Oncology Resident with clinical and lab research experience. Focus on molecular biology, diagnosis, treatment, and prognosis of gastrointestinal cancers, especially hepato-bilio-pancreatic and colorectal tumors.

Jacopo Venturini, MD., Resident in Oncology with experience in clinical and laboratory projects. The main research projects involve translational oncology and GI cancers.

Daniele Rossini, M.D., Ph.D, associate professor of Medical Oncology at the University of Florence. The main research projects in colorectal cancer.

Agnese Vannini, MD., Medical Oncologist at Careggi University Hospital. The main research projects involve gastrointestinal cancers, in particular colorectal tumors.

Marco Brugia, M.D., Medical Oncologist at Careggi University Hospital with experience in clinical and laboratory projects.

Daniele Lavacchi, M.D. Medical Oncologist at Careggi University Hospital. The main research projects involve gastrointestinal cancers.

Cristiana Conticello, M.D., Oncology Resident. The main research projects involve gastrointestinal cancers. Currently pursuing a Second-Level Master's Degree in Rare Diseases at University of Florence.

Irene Valle, M.D., Resident in Oncology. The main research projects involve gastrointestinal cancers.

Delia Ravizza, M.D., Oncology Resident. The main research projects involve gastrointestinal cancers, especially colorectal tumors.

Serena Pillozzi, PhD, CRO of the Medical Oncology Department at Careggi University Hospital. The main research projects involve cancer biology and metabolomics, translational oncology, gastrointestinal cancers, genitourinary cancers.

Lorenzo Antonuzzo, MD, PhD, professor of Medical Oncology at the Florence University, with experience in gastrointestinal and genitourinary cancers and neuroendocrine tumors