



UNIVERSITÀ
DEGLI STUDI
FIRENZE

FLORE

Repository istituzionale dell'Università degli Studi
di Firenze

**HYPOFUNCTIONALITY OF GI PROTEINS AS AETIOPHATOGENETIC
MECHANISM FOR MIGRAINE AND CLUSTER HEADACHE**

Questa è la Versione finale referata (Post print/Accepted manuscript) della seguente pubblicazione:

Original Citation:

HYPOFUNCTIONALITY OF GI PROTEINS AS AETIOPHATOGENETIC MECHANISM FOR MIGRAINE AND CLUSTER HEADACHE / N. GALEOTTI; C. GHELARDINI; M. ZOPPI; E. DEL BENE; L. RAIMONDI; E. BENEFORTI; A. BARTOLINI. - In: CEPHALALGIA. - ISSN 0333-1024. - STAMPA. - 21:(2001), pp. 38-45. [10.1046/j.1468-2982.2001.00142.x]

Availability:

The webpage <https://hdl.handle.net/2158/2280> of the repository was last updated on 2016-11-09T12:55:17Z

Published version:

DOI: 10.1046/j.1468-2982.2001.00142.x

Terms of use:

Open Access

La pubblicazione è resa disponibile sotto le norme e i termini della licenza di deposito, secondo quanto stabilito dalla Policy per l'accesso aperto dell'Università degli Studi di Firenze (<https://www.sba.unifi.it/upload/policy-oa-2016-1.pdf>)

Publisher copyright claim:

La data sopra indicata si riferisce all'ultimo aggiornamento della scheda del Repository FloRe - The above-mentioned date refers to the last update of the record in the Institutional Repository FloRe

(Article begins on next page)

Hypofunctionality of Gi proteins as aetiopathogenic mechanism for migraine and cluster headache

N Galeotti, C Ghelardini, M Zoppi¹, E Del Bene², L Raimondi, E Beneforti¹ & A Bartolini

Department of Pharmacology, ¹Department of Internal Medicine, Rheumatology Division, and ²Department of Internal Medicine, Headache Centre, University of Florence, Florence, Italy

Cephalalgia

Galeotti N, Ghelardini C, Zoppi M, Del Bene E, Raimondi L, Beneforti E & Bartolini A. Hypofunctionality of Gi proteins as aetiopathogenic mechanism for migraine and cluster headache. *Cephalalgia* 2001; 21:38–45. London. ISSN 0333-1024

The involvement of Gi proteins in the modulation of pain perception has been widely established, and mutations in G-proteins have already been identified as the aetiopathological cause of human diseases. The aim of the present study was to determine whether a deficiency or a hypofunctionality of the Gi proteins occurred in primary headache. The functionality and the level of expression of Gi proteins were investigated in lymphocytes from migraine without aura, migraine with aura and cluster headache sufferers. A reduced capability to inhibit forskolin-stimulated adenylyl cyclase activity in headache patients was observed. Migraine patients also showed basal adenosine cAMP levels about four times higher than controls. The reduced activity of Gi proteins seems not to be related to a reduction of protein levels since no significant reduction of the Gi_α subunits was observed. These results indicate Gi protein hypofunctionality as an aetiopathogenic mechanism in migraine and cluster headache. □ *Migraine, cluster headache, Gi proteins, adenylyl cyclase, lymphocytes*

Professor Alessandro Bartolini, Department of Pharmacology, Viale G. Pieraccini 6, I-50139 Florence, Italy. Tel. 39 55 4271272, fax 39 55 4271272, e-mail bartolini@server1.pharm.unifi.it Received 16 May 2000, accepted 14 November 2000

Introduction

Headache is the most common pain syndrome. It is also the most frequent symptom in neurology, where it may be a disease in itself (primary headache) or indicate an underlying local or systemic disease (secondary headache). In the complete absence of physical and laboratory alterations, diagnosis of primary headache remains purely clinical, based on the detailed description of symptoms by the patients. The criteria for classification of headache disorders was proposed in 1988 by the International Headache Society (IHS) (1) that classified primary headache into three major clinical subtypes: migraine, cluster headache and tension-type headache. Migraine is a multifaceted disorder of which the head pain is only one component. It is an extremely frequent and sometimes incapacitating condition and female preponderance is a characteristic feature (2). The two most frequent varieties of migraine are migraine without aura and migraine with aura in which neurological symptoms precede or accompany the headache (2).

Numerous factors can trigger or predispose to migraine attack. Some are identifiable, such as stress, hormonal and dietary factors, while the majority remain unknown. The exact pathogenesis of primary headache is, indeed, still unknown. Many theories have been elaborated, but none can account for all the clinical features and pathophysiological aspects of this syndrome. The 'vascular' hypothesis, which states that the headache phase of migrainous attacks is caused by vasodilatation and that the neurologic symptoms, occurring during the migraine prodrome, are produced by intracranial vasoconstriction (3), was widely accepted for many years. The 'serotonergic' hypothesis implicated a primary neuronal origin in migraine which may represent a hereditary perturbation of serotonergic neurotransmission. It was found that platelet levels of serotonin fall consistently at the onset of headache and migrainous episodes may be triggered by drugs that release this biogenic amine from tissue store (4). The 'trigemino-vascular' hypothesis is one of the most accepted theories of migraine origin. A depolarization of the trigeminal ganglions or its perivascular nerve terminals

activates the trigeminovascular system, giving rise to central transmission of nociceptive information and retrograde perivascular release of powerful vasoactive neuropeptides, such as CGRP, substance P, neurokinin A (5, 6). In more recent years, however, it was supposed that in migraine both vascular and neural components are relevant and most probably interrelated (7). Recently, 'neuronal hyperexcitability' has been proposed as the biological basis for susceptibility to migraine. Enhanced excitability of cell membrane, perhaps in part genetically determined, renders the brain susceptible to attacks. Factors that increase or decrease neuronal excitability determine the threshold for triggering attacks (8). However, from the numerous studies conducted so far, no single receptor abnormality has emerged that can completely explain why some people are more prone to suffer from migraine than others. These considerations led us to look beyond the receptor at the more universally distributed components of cell signalling that are G-proteins.

Nearly all inhibitory neurotransmitters able to enhance the pain threshold utilize Gi proteins as signal transduction system. G-proteins are heterotrimeric molecules with α , β and γ subunits. The α subunits can be classified into families, depending on whether they are targets for cholera toxin (Gs), pertussis toxin (Gi and Go) or neither (Gq and G₁₂) (9). Gi proteins represent the most widespread modulatory signalling pathway in neurones (10) and are responsible for inhibition of adenylate cyclase activity and modulation of several K⁺ and Ca²⁺ channels in order to reduce cell excitability (11, 12). Involvement of Gi proteins in the modulation of pain perception has been well established. The administration of pertussis toxin, which selectively inactivates Gi proteins (13), produced hyperalgesia and allodynia in laboratory animals (14, 15), clearly indicating that a lack of functionality of Gi proteins enhances the sensitivity to pain. Hypofunctionality of Gi proteins also produced insensitivity to analgesic treatments. It has been observed that pertussis toxin prevents the enhancement of the pain threshold induced by widely used analgesic drugs such as opioids, antihistamines, and tricyclic antidepressants (14, 16, 17).

Mutations in G-proteins have already been identified as the aetiopathological cause of human diseases. Alterations at Gs₂ subunit level have been observed in the Albright hereditary osteodystrophy, an autosomal dominant genetic pathology, as well as in the acromegaly, hyperfunctional thyroid nodules and McCune-Albright syndrome (18).

The aim of the present study was to determine whether a deficiency or a hypofunctionality of the Gi proteins occurs in headache. The functionality and expression levels of Gi proteins were therefore

investigated in lymphocytes from patients with clinically well-defined primary headache.

Patients and methods

Participants

Eighteen healthy volunteers, 21 primary-headache and 12 painful-disease (neuropathic pain, arthrosis, rheumatoid arthritis) sufferers participated in the study after informed consent was obtained from all. Patients were drug-free for a period of at least 1 week and had undergone laboratory and physical examination to exclude the presence of other concomitant infections. Furthermore, healthy subjects had no headache familial history. Patients suffering from primary headache were clinically classified following the criteria established by the IHS (1) and divided into three subgroups: migraine without aura, migraine with aura and cluster headache. Patients belonging to the painful diseases group were classified as suffering from arthrosis, rheumatoid arthritis and neuropathic pain mainly deriving from entrapment syndromes. For all patients the visual analogue scale (VAS) value, which represents an arbitrary indication of the pain experienced during the attack, was evaluated. Blood samples were collected during the quiescent state (period between migraine attacks or cluster periods) and experiments were performed in blind.

Adenylyl cyclase activity assay

Lymphocytes were isolated as described by Böyum (19). Mononuclear cell counts were made. After permeabilization with the detergent digitonin (10 µg/ml) (20), lymphocytes from healthy subjects were preincubated with or without pertussis toxin (PTX) at the concentration of 100 ng/ml at 37°C for 90 min, whereas lymphocytes from headache and painful disease sufferers were all preincubated without PTX. After preincubation, lymphocytes (1–2 × 10⁶ intact cells/assay) were incubated either with the vehicle dimethyl sulphoxide (DMSO) or forskolin 10⁻⁴ M which started the 3',5'-cyclic monophosphate (cAMP) formation (21). To obtain the inhibition curve of cAMP production, the non-hydrolysable analogue of GTP Gpp(NH)p(5'-guanylylimidodiphosphate) was added to the forskolin-containing samples in the range of concentrations from 10 nM to 100 µM in a final volume of 300 µl. After a 15-min incubation at 37°C, the cells were lysed and the sample centrifuged according to Brodde *et al.* (22). The cAMP content was determined in a 100-µl aliquot of the supernatant using an enzyme immunoassay kit. Protein was determined by bicinchoninic acid protein

assay kit with bovine serum albumin (BSA) as standard. All assays were performed in duplicate.

Western blot analysis

Aliquot of lymphocyte lysates (100 µl), or recombinant Gi_{2α} and Gi_{3α}, used as reference proteins, were boiled for 5 min with 30 µl of sample buffer (4% SDS, 20% glycerol, 10% 2-mercaptoethanol, 125 mM Tris-HCl, 0.002% bromophenol blue at pH 8). The mixture (10 mg of proteins for each sample) was separated by electrophoresis on polyacrylamide/SDS gel according to Laemli (23) using 10% acrylamide in the running gel. Following electroblotting to nitrocellulose membranes (Hybond-ECL 0.45 mm; Amersham, Milan, Italy) (24), blots were preblocked for 90 min at room temperature with TBS supplemented with 3% BSA. Following two washes (10 min each) with T-TBS (TBS containing 0.5% Tween 20) at room temperature, blots were incubated overnight at 4°C in TBS supplemented with antisera suspension (a 1:2000 for anti-Gi_{2α} and 1:10 000 working dilutions for Gi_{3α}). Following removal of antisera, blots were washed twice for 10 min with T-TBS at room temperature. Thereafter each blot was incubated with T-TBS supplemented with a second monoclonal goat anti-rabbit peroxidase conjugate antibody (1:5000 working dilution) for 1 h at room temperature. Blots were then extensively washed (1 h) with several changes of T-TBS.

Specific bands were detected using the ECL system (Amersham) according to the manufacturer's instructions. (Densitometric analysis of the bands obtained was carried out using the Scion image program.)

Statistical analysis

Results are given as the means ± SEM. An analysis of variance (ANOVA), followed by Fisher's Protected Least Significant Difference (PLSD) procedure for *post hoc* comparison, was used to verify the significance of differences between two means. Data were analysed with the StatView software for the Macintosh (1992).

Results

Clinical characteristics of patients

As reported in Table 1, all patient groups showed a comparable VAS value indicating homogeneity of intensity of pain experienced. Laboratory values in patients affected by headache were similar to healthy controls (data not shown).

Evaluation of Gi protein functionality

The functionality of Gi proteins was assessed in peripheral blood intact lymphocytes by investigating their ability to inhibit adenylyl cyclase activity. We first stimulated adenylyl cyclase by using forskolin 10⁻⁴ M. The inhibition of cAMP production by Gi proteins was started by administration of Gpp(NH)p in the concentration range of 10⁻⁸–10⁻⁴ M. In lymphocytes from healthy subjects Gpp(NH)p produced a dose-dependent inhibition of forskolin-stimulated adenylyl cyclase activity detectable by a reduction of cAMP levels of about 40%. By contrast, no modification of cAMP levels was observed after preincubation of lymphocytes with PTX. However, when lymphocytes from headache patients were used, Gpp(NH)p failed to inhibit adenylyl cyclase activity, giving an effect similar to that produced by PTX (Fig. 1a). Investigating the subtypes of primary headache, we observed an impaired inhibition of adenylyl cyclase activity in lymphocytes from migraine without aura, migraine with aura and cluster headache sufferers (Fig. 1b).

Increasing concentrations of Gpp(NH)p produced a dose-dependent reduction of cAMP levels in lymphocytes from neuropathic pain sufferers, patients affected by rheumatoid arthritis or arthrosis similar to that observed in healthy subjects. The values indicating the adenylyl cyclase activity were reported as difference (Δ) between forskolin and forskolin + Gpp(NH)p 10⁻⁵ M cAMP values in order to obtain more detailed information about the Gi protein functionality of each patient. The Δ values of all groups of headache sufferers were significantly different from both healthy subjects and patients affected by painful diseases (Fig. 2).

Determination of cAMP basal levels

Under standard conditions of assay, in healthy subjects basal levels of cAMP were 21.2 ± 2.6 pmol/mg protein. Even if there was no statistically significant correlation of cAMP levels with age or sex, older subjects showed tendentially slightly higher levels of cAMP (data not shown). Primary headache patients showed a highly significant enhancement of basal cAMP levels when compared with healthy subjects. This effect was particularly evident in patients suffering from migraine with or without aura rather than in cluster headache sufferers, since the cAMP basal levels were, respectively, about four times and twice higher than controls. By contrast, the basal cAMP levels of patients suffering from neuropathic pain, arthrosis and rheumatoid arthritis were not statistically different from healthy subjects. Patients with arthrosis differed from other groups because, even if they maintain Gi protein

Table 1 Clinical parameters of patients suffering from headache and painful diseases

| Healthy | | |
|------------------------------|------------|---------|
| Sex | Age | VAS |
| M | 29 | 0 |
| M | 32 | 0 |
| M | 43 | 0 |
| F | 58 | 0 |
| F | 49 | 0 |
| F | 36 | 0 |
| F | 25 | 0 |
| M | 27 | 0 |
| M | 41 | 0 |
| F | 43 | 0 |
| F | 25 | 0 |
| M | 50 | 0 |
| F | 25 | 0 |
| M | 48 | 0 |
| M | 44 | 0 |
| M | 62 | 0 |
| F | 25 | 0 |
| F | 31 | 0 |
| 9 M/9 | F 38.5±2.7 | |
| Headache | | |
| Sex | Age | VAS |
| <i>Migraine without aura</i> | | |
| F | 53 | 8.7 |
| F | 31 | 5.7 |
| F | 52 | 8.9 |
| F | 49 | 9.9 |
| F | 34 | 9.9 |
| M | 60 | 7.9 |
| F | 57 | 8.2 |
| F | 40 | 9.1 |
| F | 28 | 7.6 |
| F | 31 | 8.5 |
| 1 M/9 F | 43.5±3.8 | 8.4±0.4 |
| <i>Migraine with aura</i> | | |
| F | 30 | 8.8 |
| F | 73 | 7.5 |
| F | 23 | 7.7 |
| M | 38 | 8.5 |
| M | 23 | 8.3 |
| 2 M/3 F | 37.4±8.3 | 8.2±0.2 |
| <i>Cluster headache</i> | | |
| M | 28 | 9.8 |
| M | 51 | 9.9 |
| M | 37 | 9.9 |
| F | 39 | 9.9 |
| M | 36 | 9.0 |
| M | 27 | 8.3 |
| 5 M/1 F | 36.3±3.2 | 9.4±0.2 |

Table 1 (contd.)

| Painful diseases | | |
|-----------------------------|----------|---------|
| Sex | Age | VAS |
| <i>Neuropathic pain</i> | | |
| F | 60 | 8.0 |
| M | 75 | 9.3 |
| F | 61 | 5.4 |
| F | 72 | 9.3 |
| F | 55 | 7.3 |
| 1 M/4 F | 64.6±3.4 | 7.9±0.6 |
| <i>Arthrosis</i> | | |
| M | 82 | 9.2 |
| F | 60 | 7.1 |
| F | 77 | 9.6 |
| F | 77 | 3.7 |
| 1 M/3 F | 74.0±4.2 | 7.4±1.2 |
| <i>Rheumatoid arthritis</i> | | |
| F | 73 | 4.6 |
| M | 61 | 2.6 |
| F | 75 | 9.2 |
| 1 M/2 F | 69.7±3.6 | 5.5±1.6 |

VAS, Visual analogue scale.

functionality, they showed higher levels of cAMP (Fig. 3).

By comparing forskolin-stimulated levels of cAMP, no statistically significant difference was seen among values observed in healthy subjects and in all the investigated groups of patients (Fig. 3).

Quantitative determination of $G_{i\alpha}$ levels

The expression of $G_{i\alpha}$ and of the subtypes $G_{i2\alpha}$ and $G_{i3\alpha}$ proteins were investigated. As illustrated in Fig. 4, levels of $G_{i\alpha}$, $G_{i2\alpha}$ and $G_{i3\alpha}$, quantified by optical densitometry, were similar in lymphocytes from both healthy subjects and headache sufferers. Some representative examples of Western blot analysis bands are reported as inset.

Discussion

Present results showed that lymphocytes from headache patients showed a reduced capability to inhibit forskolin-stimulated adenylyl cyclase activity, clearly showing a hypofunctionality of Gi proteins. The observed impaired capability to reduce cAMP levels in headache sufferers was similar to that produced by preincubation of lymphocytes from healthy subjects with pertussis toxin at a concentration consistent with a complete ADP-ribosylation of Gi proteins (25),

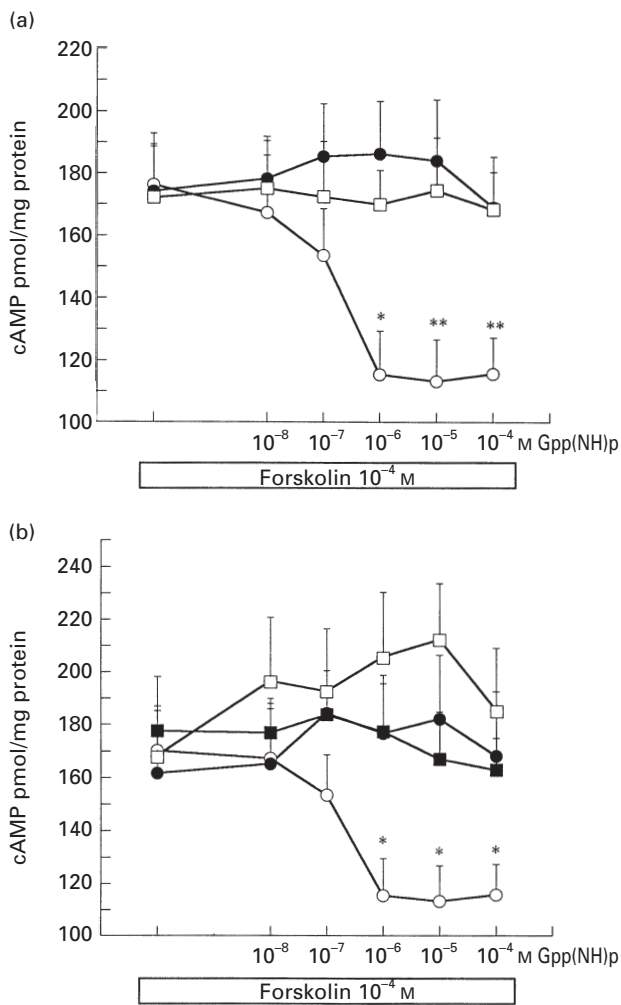


Figure 1 Evaluation of adenylyl cyclase activity in lymphocytes from healthy and migraine subjects. (A) Lymphocytes from healthy subjects were preincubated with or without pertussis toxin (PTX). **P* < 0.05; ***P* < 0.01 vs. forskolin-stimulated lymphocytes. ○, Healthy; ●, headache; □, PTX. (b) Lack of reduction of cAMP levels in patients suffering from migraine without aura (MO; ■), migraine with aura (MA; □) and cluster headache (CH; ●). **P* < 0.05 vs. corresponding value of headache group. ○, Healthy.

further indicating a selective hypofunctionality of Gi proteins. All headache subtypes investigated, even if they had different clinical characteristics, showed a similar reduction of inhibitory functionality, suggesting a fundamental role of Gi proteins in the aetiopathogenesis of both migraine and cluster headache.

Lymphocytes from headache suffering subjects showed higher basal levels of cAMP. These results indicate that, at resting conditions, there is a disequilibrium between the inhibitory and stimulatory system of this enzyme, mediated, respectively, by Gi and Gs proteins, confirming the hypothesis of a hypofunctionality of Gi proteins. An alteration at the level of the

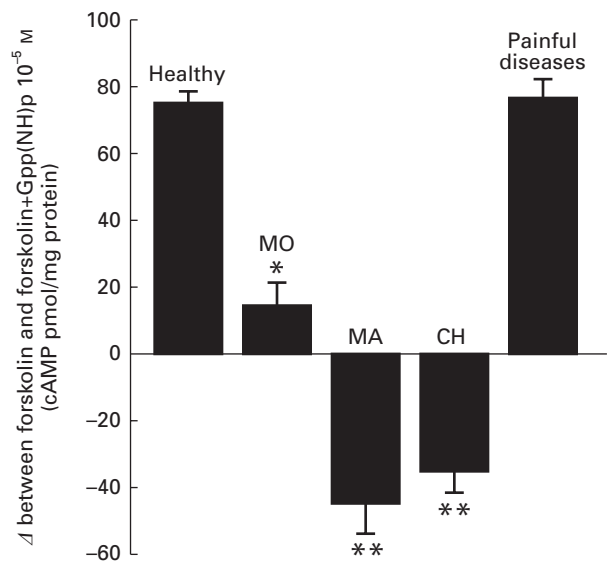


Figure 2 Evaluation of adenylyl cyclase activity in lymphocytes from headache patients in comparison with patients suffering from neuropathic pain, rheumatoid arthritis and arthrosis. **P* < 0.05; ***P* < 0.01 vs. healthy group. MO, Migraine without aura; MA, migraine with aura; CH, cluster headache.

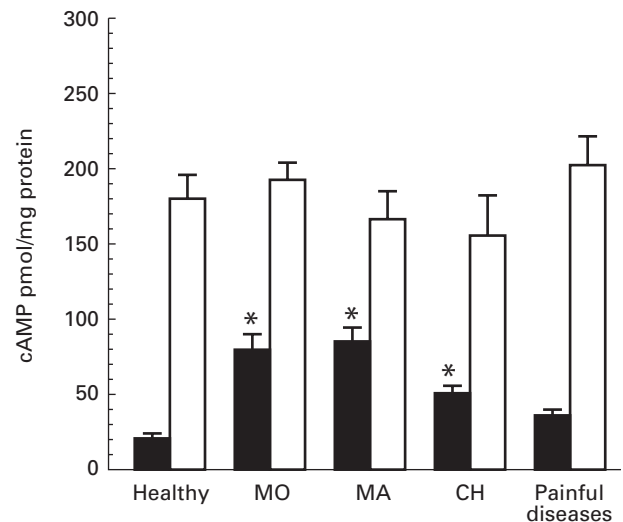


Figure 3 Evaluation of basal (■) and forskolin-stimulated (□) cAMP levels in patients suffering from migraine without aura (MO), migraine with aura (MA), cluster headache (CH), rheumatoid arthritis, neuropathic pain, and arthrosis. **P* < 0.05 vs. healthy subjects.

adenylyl cyclase can also be excluded, since the direct stimulation of the enzyme by forskolin did not reveal any difference in cAMP levels produced between patients and healthy subjects.

Among the several classes of compounds that have been investigated as antimigraine agents, dihydroergotamine or triptans represent the reference drugs for acute

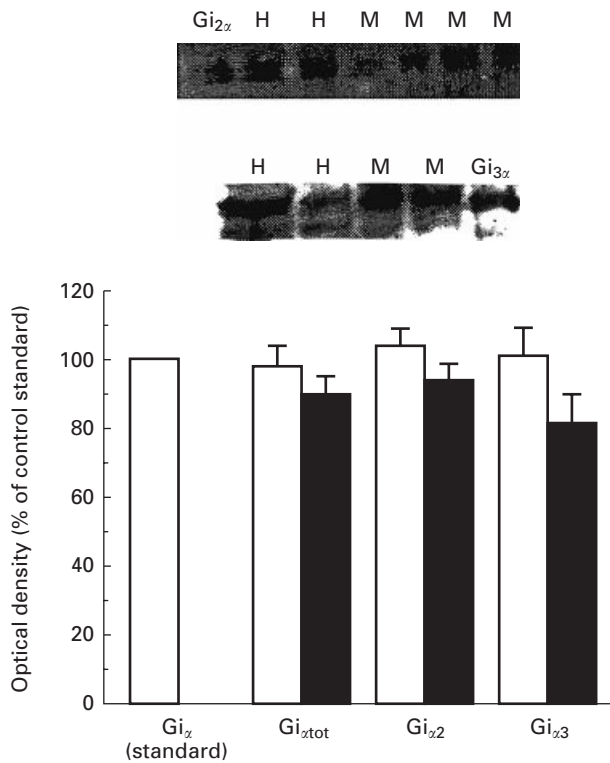


Figure 4 Evaluation of Gi α protein levels in lymphocytes of patients suffering from primary headache (M; ■) in comparison with healthy (H; □) subjects. Data are reported as percentage of corresponding Gi α subtype standard. Inset: bands obtained by Western blot analysis.

migraine treatment (26). These compounds exert their activity by inducing vasoconstriction of cranial vessels, even if it is not completely elucidated whether these drugs exert their therapeutic action through an effect on 5-HT_{1B/D} receptors or via a direct effect on vessels (27). It has long been known that cAMP is a potent vasodilator of cerebral vessels. The relaxation of vascular smooth muscle produced by compounds that activate adenylate cyclase, such as β -adrenergic agents and forskolin, is thought to depend on their capability to increase the concentration of cAMP (28). Rosenblum (29) demonstrated the existence in mice of an adenylyl cyclase-cAMP system for dilating cerebral arterioles. Furthermore, it has been reported that a suffusion with cAMP produced a concentration-dependent sustained vasodilatation of the resting pial arteries in rats (30). The clinical efficacy of vasoconstrictor agents is in agreement with our experimental findings. We observed higher basal levels of cAMP specifically in migraine patients, and this biochemical alteration could represent the origin of the vasodilatation which, in turn, represents the cause triggering the migraine attack. Further confirmation of this hypothesis is provided by the observation that among the drugs employed to

prevent effectively migraine attacks are the β -blockers (3) which are responsible for lowering the intracellular levels of cAMP.

The most recent theory of headache pathogenesis considers brain hyperexcitability as the biological basis for susceptibility to migraine (8). This theory is based on some studies which showed that occipital cortex neurones may be hyperexcitable in some 90% of patients suffering from migraine with aura who experience visual disturbances as part of their aura (31). Furthermore, psychological studies of visual discrimination showed that migraine sufferers had a greater sensitivity for low-level visual processing between attacks, showing functional impairment of inhibitory interneurons (32). It has also been reported that cortical and precortical visual and auditory processing in migraine patients, investigated by means of pattern-reversal visual evoked potentials (VEP) and auditory cortical evoked potentials (IDAP), is impaired between attacks (33, 34). Our results obtained with migraine sufferers are not only in agreement with the above-mentioned hypothesis, but the observed hypofunctionality of Gi proteins can represent the biological mechanism responsible for the cell hyperexcitability. It is well known, indeed, that Gi protein activation reduces cell excitability (12). A deficiency of the Gi protein-mediated inhibitory system increases cell excitability, leading to a hypersensitivity to stimuli that can also represent the main cause of the symptoms experienced by headache sufferers, such as hypersensitivity to pain, photophobia, phonophobia, etc.

Gi proteins are coupled to many receptor types and subtypes. It is also well known that Gi protein levels can be down-regulated at several levels, such as turnover, transcription, etc., by hormones and neurotransmitters. Considering that all subjects included in this study were drug-free for at least 1 week, we can assume that the observed Gi protein hypofunctionality represents an individual's predisposition to headache rather than a down-regulation produced by the pharmacological treatment.

It can also be excluded that the observed hypofunctionality of Gi proteins could be an adaptive response to a condition of chronic pain. Patients suffering from painful diseases such as neuropathic pain, arthrosis or rheumatoid arthritis, who experienced pain sensations comparable to headache sufferers, as indicated by the VAS values, showed an unaltered Gi protein functionality.

As the observed hypofunctionality of Gi proteins could imply a reduction of protein expression, levels of Gi proteins were determined in cell membrane preparations from headache sufferers using Western blot analysis. To this purpose, antibodies against Gi α subunits, which represent the functional subunit of

Gi proteins (9), were employed. Furthermore, specific antibodies against $Gi_{2\alpha}$ and $Gi_{3\alpha}$ subtypes were used, since Gi_2 represents the Gi protein subtype mainly involved in the inhibitory regulation of adenylyl cyclase activity *in vivo* (35) and it plays an important role in the modulation of pain perception. The administration of selective antibodies against $Gi_{2\alpha}$ as well as the inhibition of its expression by the use of specific antisense oligonucleotides prevented the analgesia induced by agonists of μ -opioid receptors (36, 37). Similarly to Gi_2 , Gi_3 has also been reported to be involved in pain processing, since antibodies against $Gi_{3\alpha}$ prevented δ -opioid analgesia (38). No statistically significant reduction of Gi protein levels was observed in headache patients, leading us unable to establish from these data that Gi protein hypofunctionality is subsequent to a reduction of protein levels.

The lack of Gi protein reduction seems to be in contrast to recent data in which a reduction of about 50% of $Gi_{2\alpha}$ mRNA was observed in lymphocytes from migraine patients (39). These results were obtained using Northern blot analysis and it is therefore plausible that this discrepancy could be attributed to the different recognition sites of the probes used for the determination of mRNA levels in comparison with those of the antibodies employed for Western blot analysis. The hypofunctionality of Gi proteins could be, in fact, due to the presence of a mutation in the protein sequence. In this condition, the use of a cDNA probe to quantify mRNA contents, wherever the mutation is located, may show a level reduction. Conversely, using Western blot analysis a reduction of Gi_{α} protein levels might not be shown, since the short segment of the protein with which the employed antibodies interact might not contain the mutation.

A structural alteration of the Gi proteins in headache sufferers can be excluded since there was no difference in the molecular weight of the α subunits of any Gi protein subtype investigated in comparison with healthy subjects.

The investigation into Gi protein functionality was conducted in lymphocytes from peripheral blood taking into account that Gi proteins have an ubiquitous distribution (40) and that blood samples can easily be obtained from patients in a painless manner. Lymphocytes represent an appropriate tool to detect Gi protein functionality since they have cell surface recognition sites for pertussis toxin, the selective inactivator of Gi, which are lacking in other blood cell types such as erythrocytes (41). Lymphocytes have also been widely used to investigate the aetiopathogenesis of human diseases at a biochemical and molecular level (42–46).

In conclusion, our results indicate that lymphocytes from migraine and cluster headache patients are

distinguishable from lymphocytes not only from healthy subjects but also from patients suffering from other painful diseases.

Acknowledgements

The authors thank Dr Silvia Quattrone for her methodological and technical advice, Dr David Beccani for linguistic revision and all subjects and patients for their availability and confidence. The study was partially supported by grants from Boehringer Ingelheim and Florence University (ex 60%).

A diagnostic kit obtained from these results is under patent of the University of Florence.

References

- 1 Classification Committee of the International Headache Society. Classification and diagnostic criteria for headache disorders, cranial neuralgias and facial pain. *Cephalalgia* 1988; 8 (Suppl. 7):1–96.
- 2 Raskin NJ. Headache. In: Martin JD, Fauci AS, Kasper DL, eds. *Harrison's principles of internal medicine*, 13th edn. New York: McGraw-Hill Inc., 1994:65–71.
- 3 Wolff HG. Headache and other head pain. New York: Oxford University Press, 1963.
- 4 Fozard JR. 5-Hydroxytryptamine in the pathophysiology of migraine. In: Bevan JA, ed. *Vascular neuroeffector mechanism*. Amsterdam: Elsevier Science Publishers, 1985:321–8.
- 5 Moskowitz MA. Neurogenic versus vascular mechanisms of sumatriptan and ergot alkaloids in migraine. *Trends Pharmacol Sci* 1992; 13:307–12.
- 6 Goadsby PJ, Edvinsson L. The trigeminovascular system and migraine: studies characterizing cerebrovascular and neuropeptide change seen in man and cat. *Ann Neurol* 1993; 33:48–56.
- 7 Olesen J. Clinical and pathophysiological observations in migraine and tension-type headache explained by integration of vascular, supraspinal and myofascial inputs. *Pain* 1991; 46:125–32.
- 8 Welch KMA. Current opinions in headache pathogenesis: introduction and synthesis. *Curr Opin Neurol* 1998; 11:193–7.
- 9 Simon MI, Strathman MP, Gautman N. Diversity of G proteins in signal transduction. *Science* 1991; 252:802–8.
- 10 Holz GG, Rane SG, Dunlap K. GTP-binding proteins mediate transmitter inhibition of voltage-dependent calcium channels. *Nature* 1986; 319:670–2.
- 11 Hille B. Modulation of ion-channel function by G-protein-coupled receptors. *Trends Neurosci* 1994; 17:531–6.
- 12 Sprang SR. G-protein mechanisms: insights from structural analysis. *Annu Rev Biochem* 1997; 66:639–78.
- 13 Katada T, Ui M. Direct modification of the membrane adenylyl cyclase system by islet-activating protein due to ADP-ribosylation of a membrane protein. *Proc Natl Acad Sci USA* 1982; 79:3129–33.
- 14 Galeotti N, Ghelardini C, Bartolini A. Effect of pertussis toxin on morphine, diphenhydramine, baclofen, clomipramine and physostigmine antinociception. *Eur J Pharmacol* 1996; 308:125–33.
- 15 Womer DE, DeLapp NW, Shannon HE. Intrathecal pertussis toxin produces hyperalgesia and allodynia in mice. *Pain* 1997; 70:223–8.

- 16 Parenti M, Tirone F, Giagnoni G, Pecora N, Parolaro D. Pertussis toxin inhibits the antinociceptive action of morphine in the rat. *Eur J Pharmacol* 1986; 124:357-9.
- 17 Galeotti N, Ghelardini C, Capaccioli S, Quattrone A, Nicolin A, Bartolini A. Blockade of clomipramine and amitriptyline analgesia by an antisense oligonucleotide to mKv1.1, a mouse Shaker-like potassium channel. *Eur J Pharmacol* 1997; 330:15-25.
- 18 Spiegel AM. Defects in G protein-coupled signal transduction in human disease. *Annu Rev Physiol* 1996; 58:143-70.
- 19 Böyum A. Isolation of mononuclear cells and granulocytes from human blood. *Scand J Clin Lab Invest* 1968; 21 (Suppl. 97):77-89.
- 20 Corey SJ, Rosoff PM. Granulocyte-macrophage colony-stimulating factor primes neutrophils by activating a pertussis toxin-sensitive G protein not associated with phosphatidylinositol turnover. *J Biol Chem* 1989; 264:14165-71.
- 21 Griese M, Griese S, Reinhardt D. Inhibitory effects of pertussis toxin on the cAMP generating system in human mononuclear leucocytes. *Eur J Clin Invest* 1990; 20:317-22.
- 22 Brodde O-E, Brinkmann M, Schemuth R, O'Hara N, Daul A. Terbutaline-induced desensitization of human lymphocyte β_2 -adrenoceptors. *J Clin Invest* 1985; 76:1096-101.
- 23 Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 1970; 227:680-5.
- 24 Towbin MT, Staehelin H, Gordon J. Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proc Natl Acad Sci USA* 1979; 76:4350-4.
- 25 Watkins DC, Northup JK, Malbon CC. Pertussis toxin treatment in vivo is associated with a decline in G-protein β -subunits. *J Biol Chem* 1989; 264:4186-94.
- 26 Lobo BL, Cooke SC, Landy SH. Symptomatic pharmacotherapy of migraine. *Clin Ther* 1999; 21:1118-30.
- 27 Diener HC, Kaube H, Limmroth V. Antimigraine drugs. *J Neurol* 1999; 246:515-9.
- 28 Takuwa Y, Takuwa N, Rasmussen H. The effects of isoproterenol on intracellular calcium concentration. *J Biol Chem* 1988; 263:762-8.
- 29 Rosenblum WI. In vivo evidence that an adenylate cyclase-cAMP system dilates cerebral arterioles in mice. *Stroke* 1988; 19:888-91.
- 30 Hong KW, Shin HK, Kim HH, Choi JM, Rhim BY, Lee WS. Metabolism of cAMP to adenosine: role in vasodilation of rat pial artery in response to hypotension. *Am J Physiol* 1999; 276:H376-82.
- 31 Wilkins A, Nimmo-Smith I, Tait A, McManus C, Della Sala A, Tilley A, Lotsch J. A neurological basis for visual discomfort. *Brain* 1984; 107:989-1017.
- 32 Wraya SH, Mijovic-Preleca D, Kosslyna SM. Visual processing in migraineurs. *Brain* 1995; 118:25-35.
- 33 Sandor PS, Afra J, Proietti-Cecchini A, Albert A, Schoenen J. Familial influences on cortical evoked potentials in migraine. *Neuroreport* 1999; 10:1235-8.
- 34 Oelkers R, Grosser K, Lang E, Geisslinger G, Kobal G, Brune K, Lotsch J. Visual evoked potentials in migraine patients: alterations depend on pattern spatial frequency. *Brain* 1999; 122:1147-55.
- 35 Moxham CM, Hod Y, Malbon CC. G_{i2} mediates the inhibitory regulation of adenylate cyclase in vivo: analysis in transgenic mice with G_{i2} suppressed by inducible antisense RNA. *Dev Genet* 1993; 14:266-73.
- 36 Sanchez-Blazquez P, Juarros JL, Martinez-Peña Y, Castro MA, Garzon J. $G_{\alpha_{x/z}}$ and G_{i2} transducer proteins on μ/δ opioid-mediated supraspinal antinociception. *Life Sci* 1993; 53:PL381-6.
- 37 Raffa RB, Martinez RP, Connelly CD. G-protein antisense oligodeoxyribonucleotides and μ -opioid supraspinal antinociception. *Eur J Pharmacol* 1994; 258:R5-7.
- 38 Sanchez-Blazquez P, Garzon J. δ -opioid supraspinal antinociception in mice is mediated by G_{i3} transducer proteins. *Life Sci* 1993; 53:129-34.
- 39 Gardiner IM, Ahmed F, Steiner TJ, McBain A, Kennard C, de Belleruche J. A study of adaptive response in cell signalling in migraine and cluster headache: correlations between headache type and changes in gene expression. *Cephalalgia* 1998; 18:192-6.
- 40 Hepler JR, Gilman AG. G proteins. *Trends Biochem Sci* 1992; 17:383-7.
- 41 Ui M, Nogimori K, Makoto T. Pertussis toxin. New York: Academic Press, 1985.
- 42 Diamond I, Wrubel B, Estrin W, Gordon A. Basal and adenosine receptor-stimulated levels of cAMP are reduced in lymphocytes from alcoholic patients. *Proc Natl Acad Sci USA* 1987; 84:1413-6.
- 43 Maisel AS, Michel MC, Insel PA, Ennis C, Ziegler MG, Phillips C. Pertussis toxin treatment of whole blood. A novel approach to assess G protein function in congestive heart failure. *Circulation* 1990; 81:1198-204.
- 44 Nemoz G, Prigent AF, Aloui R, Charpin G, Gormand F, Gallet H et al. Impaired G-proteins and cyclic nucleotide phosphodiesterase activity in T-lymphocytes from patients with sarcoidosis. *Eur J Clin Invest* 1993; 23:18-27.
- 45 Wand GS, Waltman C, Martin CS, McCaul ME, Levine MA, Wolfgang D. Differential expression of guanosine triphosphate binding proteins in men at high and low risk for the future development of alcoholism. *J Clin Invest* 1994; 94:1004-11.
- 46 Mitchell PB, Manji HK, Chen G, Jolkovsky L, Smith-Jackson E, Denicoff K et al. High levels of G_{α_x} in platelets of euthymic patients with bipolar disorders. *Am J Psychiatry* 1997; 54:218-23.