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In vitro antimycotic activity of some plant extracts towards yeast and yeast-like strains / P. PINELLI; A. ROMANI; D. HEIMLER; F. FRANCONI; A. MARTINI; B. TURCHETTI; P. BUZZINI. - In: PHYTOTHERAPY RESEARCH. - ISSN 0951-418X. - STAMPA. - 19(1):(2005), pp. 44-49. [10.1002/ptr.1622]

Availability:

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Published version:

DOI: 10.1002/ptr.1622

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In vitro Antimycotic Activity of Some Plant Extracts Towards Yeast and Yeast-like Strains

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As part of screening aimed at the selection of novel antimycotic compounds of vegetable origin, leaf extracts of *Camellia sinensis* L., *Cupressus sempervirens* L. and *Pistacia lentiscus* L. and the seed extract of *Glycine soja* Sieb. et Zucc. were tested against yeast and yeast-like species implicated in human mycoses. Of the extracts only those of *C. sinensis* (obtained from a commercial preparation of green tea) exhibited broad activity towards *Candida glabrata*, *Clavispora lusitanae*, *Cryptococcus laurentii*, *Filobasidiella neoformans*, *Issatchenkia orientalis*, *Saccharomyces cerevisiae* and *Prototheca wickerhamii* strains. MICs ranging from 300 to 4800 µg extract/mL (corresponding to 130–2010 µg/mL total polyphenols) were observed. Concentrations of the *C. sinensis* extract over 25 000 µg/mL caused a rapid decrease of viable cells of *Fil. neoformans* and its activity was dose-dependent. Tests carried out using the pure polyphenols present in *C. sinensis* extract composition, showed that only epicatechin-3-O-gallate (ECG) and epigallocatechin-3-O-gallate (EGCG) possess antimycotic activity. Copyright © 2005 John Wiley & Sons, Ltd.

Keywords: antimycotic activity; *Camellia sinensis* L.; yeast and yeast-like strains; ECG; EGCG.

INTRODUCTION

The antimicrobial properties of drugs from medicinal and other edible plants have been recognized since antiquity (Cowan, 1999). Notwithstanding the wide literature concerning the beneficial effects of plant polyphenols in human health (Cowan, 1999; Krauze-Baranowska *et al.*, 1999; Lin *et al.*, 1999; Cassidy *et al.*, 2000; Wang, 2000), to our knowledge little information is available on their antimycotic properties towards yeast and yeast-like microorganisms, which are implicated in many human pathologies (Hazen, 1995; Rex *et al.*, 1995; Warren and Hazen, 1998). In view of the increased number of fungal infections, especially in immunodepressed subjects (Kalowky *et al.*, 1997), and of the development of resistance towards antifungal agents (Hazen, 1995; Rex *et al.*, 1995; Warren and Hazen, 1998; Sanglard and Odds, 2002), there is a considerable scientific and commercial interest in the discovery of novel classes of antimycotic compounds.

Plants represent a good source of novel antimicrobial molecules (Rusia and Srivastava, 1988; Ayoub, 1989; Mahajan *et al.*, 1991; Etkin, 1996; Arora and Ohlan, 1997; Chapman *et al.*, 1997; Johns, 1999; Pieroni, 2000; Buzzini and Pieroni, 2003). Therefore the antimycotic activity was investigated of an extract of *Camellia sinensis* L. leaves and *Glycine soja* Sieb. et Zucc seeds, plants of special importance in the oriental diet which

is very rich in polyphenols. In particular, the former is rich in galloyl-catechins (Riemersma *et al.*, 2001) while the latter is very rich in isoflavones such as genistein and daidzein glycosides (Romani *et al.*, 2003). In order to investigate whether other polyphenol classes may have an antimycotic activity another edible plant of the Mediterranean region was selected, *Pistacia lentiscus*, which contains galloyl-quinic esters and myricetin glycosides as the main compounds (Romani *et al.*, 2002b). Moreover, *Cupressus sempervirens* extract has been investigated because it is rich in biflavonoids, quercetin and kaempferol glycosides (Romani *et al.*, 2002a). The present paper compared the activity of different plant extracts in order to identify what class of polyphenols is able to express antimycotic activity towards yeast and yeast-like microorganisms implicated in many human pathologies (Hazen, 1995; Rex *et al.*, 1995; Warren and Hazen, 1998).

MATERIALS AND METHODS

Plant material. The leaves of the mastic tree and cypress tree were collected from fully developed plants growing in Tuscany (Italy) at the end of June and September (mastic tree) and October (cypress tree) 2002. In both cases the leaves were processed as previously described (Romani *et al.*, 2002a; Romani *et al.*, 2002b). Soy seeds, collected from plants grown under natural conditions, were ground and extracted as previously reported (Romani *et al.*, 2003). Green tea extract (from *Camellia sinensis* leaves) was obtained from Indena S.p.A., Milan, Italy. The extracts were

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stored at -20°C until use and were stable for at least 12 months.

HPLC/DAD and HPLC/MS analysis. The analysis was carried out using a HP-1100 liquid chromatograph equipped with a DAD detector and a HP 1100 MSD API-electrospray (Agilent-Technologies, Palo Alto, USA) operating in positive and negative ionization mode. Analytical conditions were described previously (Romani *et al.*, 2003). Identification of individual polyphenols was carried out on the basis of their retention times, spectroscopic and spectrometric data. Quantification of individual polyphenols was directly performed by HPLC-DAD by using four-point regression curves built with the available standards or isolated compounds. The following standards were used for the identification and characterization of plant extracts: phenolic acid (ellagic acid, gallic acid, caffeic acid, chlorogenic acid and cynarin); secoiridoid (oleuropein); flavones (apigenin, cosmetin, homoorientin, vitexin, luteolin, luteolin-4'-glucoside); flavonols (quercetin, quercetagenin, quercetin-3-arabinoside, quercitrin, isoquercitrin, fisetin, kaempferol, kaempferol-7-neohesperidoside, myricetin, myricitrin, rhamnetin, isorhamnetin and robinin); flavanones (eriodictyol, hesperidin, naringenin, naringenin-7-glucoside and pinocembrin-7-methylther); isoflavones (daidzein and genistein); chalcones (phloridzin) and catechins, epicatechin-3-O-gallate (ECG), epigallocatechin (EGC) and epigallocatechin-3-O-gallate (EGCG). Calibration curves with $r^2 \geq 0.9998$ were considered. The quantification was performed at the maximum wavelength of UV-Vis absorbance by applying the correction for molecular weight.

In particular, in *G. soja* seed extract genistein and glycitein derivatives were determined at 260 nm using genistein as a reference compound, while daidzein derivatives were determined at 305 nm with daidzein as a reference; in *P. lentiscus* leaf extract the galloyl-glycosides and galloyl-quinic acid amounts were calculated at 280 nm using gallic acid as a reference, while myricetin and quercetin glycosides were calibrated at 350 nm with myricitrin and rutin, respectively, as a reference; in *C. sempervirens* leaf extract the flavonol content was determined at 350 nm employing the authentic standard quercitrin and kaempferol, in the same way biflavonoids and methylbiflavonoids were quantified with the authentic standard hinokiflavone and amentoflavone; finally, the commercial extract of green tea was calibrated at 280 nm using the authentic standards of catechin and gallo-catechins, while its flavonol content was calculated at 350 nm using rutin as a reference compound.

Microorganisms. Twenty-four yeast (belonging to 13 species of nine genera) and three yeast-like (*Prototheca* spp.) strains of different origin (conserved in the DBVPG Collection of Industrial Yeasts at the Dipartimento di Biologia Vegetale e Biotecnologie Agroambientali, University of Perugia, Italy, www.agr.unipg.it/dbvpg) (Buzzini and Martini, 2000) were used as target microorganisms (Table 1). Several of these, although belonging to species commonly regarded as species of agro-industrial interest, have gained the status of emerging pathogens in recent years because they are implicated in opportunistic infections (Hazen, 1995; Rex *et al.*, 1995; Warren and Hazen, 1998). Working cultures were maintained on YEPG (g/L: yeast

Table 1. Salient information on yeasts and yeast-like microorganisms used in the present study

Species	Strain	Status	Source of isolation	Locality of isolation
<i>Candida albicans</i>	DBVPG 6133	T	Skin mycosis	Uruguay
<i>Candida albicans</i>	DBVPG 6157	NT of <i>Candida stellatoidea</i>	Unknown	Unknown
<i>Candida glabrata</i>	DBVPG 7212		Unknown	Unknown
<i>Candida glabrata</i>	DBVPG 3828	T	Faeces	Unknown
<i>Pichia guilliermondii</i>	DBVPG 6140	T of <i>Candida guilliermondii</i>	Sputum	Unknown
<i>Candida parapsilosis</i>	DBVPG 6150	T	Ex case of sprue	Puerto Rico
<i>Candida tropicalis</i>	DBVPG 3982	T	Bronchitic patient	Unknown
<i>Candida zeylanoides</i>	DBVPG 6163	NT	Blastomycotic macroglossia	Unknown
<i>Clavispora lusitaniae</i>	DBVPG 6142	T of <i>Candida lusitaniae</i>	Caecum of pig	Portugal
<i>Clavispora lusitaniae</i>	DBVPG 6148		Sputum	Norway
<i>Issatchenkia orientalis</i>	DBVPG 6782		Fruit juice	Unknown
<i>Kluyveromyces marxianus</i>	DBVPG 6141	T of <i>Candida kefir</i>	Kefyr grain	The Netherlands
<i>Saccharomyces cerevisiae</i>	DBVPG 6173	T	Brewer's top yeast	The Netherlands
<i>Saccharomyces cerevisiae</i>	DBVPG 6497		Unknown	Unknown
<i>Saccharomyces cerevisiae</i>	DBVPG 6500		Unknown	Unknown
<i>Yarrowia lipolytica</i>	DBVPG 6053	T	Maize-processing plant	USA
<i>Cryptococcus laurentii</i>	DBVPG 3883		Phlegm of a tuberculosis patient	Unknown
<i>Cryptococcus laurentii</i>	DBVPG 4272		Human oral cavity	Italy
<i>Cryptococcus laurentii</i>	DBVPG 6265	T	Palm wine	Congo
<i>Filobasidiella neoformans</i>	DBVPG 3428		Unknown	Unknown
<i>Filobasidiella neoformans</i>	DBVPG 6010	T of <i>Cryptococcus neoformans</i>	Fermenting fruit juice	Unknown
<i>Filobasidiella neoformans</i>	DBVPG 6225		Spinal fluid	USA
<i>Filobasidiella neoformans</i>	DBVPG 6981		Cerebrospinal fluid	USA
<i>Filobasidiella neoformans</i>	DBVPG 6982		Droppings of cuckoo	Thailand
<i>Prototheca wickerhamii</i>	DBVPG 8879		Unknown	Unknown
<i>Prototheca zopfii</i>	DBVPG 8880		Unknown	Unknown
<i>Prototheca zopfii</i>	DBVPG 8830		Milk from a mastitic cow	Italy

T, Type strain; NT, Neotype.

extract 10, peptone 10, glucose 20, agar 15) or potato dextrose agar (PDA) (Atlas and Parks, 1993) slants at 4 °C until use.

Antimycotic activity test. Antimycotic activity of the crude extracts was evaluated by using the agar diffusion well bioassay (ADWB) (Buzzini and Martini, 2001) with YEPG or yeast nitrogen base (YNB) agar (Difco, Detroit, Michigan, USA) + 2% glucose as substrates.

One mL of 24 h cell suspensions (10^5 cells/mL corresponding to an $A_{580} = 0.1 - 0.2$) of strains were mixed thoroughly with 19 mL molten medium (45 °C) and poured into a Petri dish. After the agar had solidified, six wells (diameter 8 mm) were cut equidistant from the edge of the inoculated dish. An aliquot (100 µL) of extract or standard stock solution was placed in each well. Dishes were checked for the presence of inhibition zones (halos) every 12 h. In this phase of the work, antimicrobial activity was assessed by measuring the diameter of the clear area of inhibition.

Ketoconazole (as control antibiotic) (Calbiochem Inc., USA) and standard compounds, as listed above, were also tested for their antimycotic activity by ADWB. All tests were performed in triplicate.

Determination of minimal inhibitory concentration (MIC). MIC determination was carried out in 96-well microplates (Corning Inc., USA) by a broth microdilution method in agreement with the NCCLS recommendations (NCCLS, 1997). The extracts were diluted in YNB broth + 2% glucose in order to obtain concentrations ranging from 5000 to 100 µg/mL. Stock solutions (100 µg/mL) of ketoconazole (as control antibiotic) in 10% dimethyl sulphoxide (DMSO) (Carlo Erba, Italy) was prepared before use and concentrations ranging from 100 to 0.1 µg/mL were used.

Assessment of fungicidal activity of *C. sinensis* extract.

Cells of *Fil. neoformans* DBVPG 6010, grown for 24 h at 25 °C on YEPG agar slants, were suspended in sterile distilled water (10^6 cells/mL). A quantity of 50 µL of cell suspension was used as the inoculum of 450 µL of YNB broth + 2% glucose. Aliquots of extract were added to obtain increasing concentrations: 1250, 2500, 6250, 12 500, 25 000 and 50 000 µg/mL of extract. Over a period of 32 h, aliquots of cultures were collected at 4 h intervals and viable cells were plated on YEPG agar dishes. Colonies were counted after 48–72 h. A control test (extract free) was also included.

RESULTS AND DISCUSSION

C. sinensis extract was particularly rich in galloyl-derivatives of catechins with epigallocatechin-3-O-gallate, epigallocatechin and epicatechin being the lead compounds (Table 2). Indeed in *P. lentiscus* extract, the most prominent compounds were galloyl-derivatives (galloyl quinic acid, galloyl glycosides), gallic acid and flavonol glycosides (Table 2). *G. soja* extracts were very rich in isoflavones such as daidzein and genistein (Table 2), whereas the flavonoids and biflavonoids were the major compounds in the leaf extract of *C. sempervirens* (Table 2). The polyphenol composition of extracts was in line with previously reported data (Romani *et al.*, 2002a; 2002b; 2003).

The extracts from the cypress tree, mastic tree and soybean did not show any antimycotic activity against yeast and yeast-like strains (data not shown). Thus, galloyl quinic acid, galloyl glycosides, gallic acid, myricetin glycosides present in *P. lentiscus*, isoflavones of *G. soja* and the flavonoids and biflavonoids of *C. sempervirens* had no antimycotic activity.

Table 2. Quantitative analysis of polyphenols (expressed as µg/mL) present in the extracts from leaves of *C. sempervirens*, *C. sinensis* and *P. lentiscus*, and seeds of *G. soja*

Compound	Extract composition (µg/mL)			
	<i>C. sempervirens</i>	<i>C. sinensis</i>	<i>G. soja</i>	<i>P. lentiscus</i>
Genistein	n.d.	n.d.	5.3	n.d.
Genistein-glycosides	n.d.	n.d.	303	n.d.
Daidzein	n.d.	n.d.	13	n.d.
6 OH-Daidzein	n.d.	n.d.	2.7	n.d.
Daidzein glycosides	n.d.	n.d.	490	n.d.
Glycithin glycosides	n.d.	n.d.	28	n.d.
Caffeic acid derivatives	n.d.	n.d.	16	n.d.
Gallic acid	n.d.	0.9	n.d.	25
Galloyl glycosides	n.d.	n.d.	n.d.	19
Galloyl quinic acid	n.d.	n.d.	n.d.	310
Myricetin glycosides	n.d.	n.d.	n.d.	102
Quercetin glycosides	174	n.d.	n.d.	25
Kaemferol glycosides	16	n.d.	n.d.	n.d.
Flavonols (as equivalent rutin)	n.d.	35	n.d.	n.d.
Biflavonoids	1460	n.d.	n.d.	n.d.
Methylbiflavonoids	4	n.d.	n.d.	n.d.
Epigallocatechin-3-O-gallate (EGCG)	n.d.	211	n.d.	n.d.
Epigallocatechin (EGC)	n.d.	105	n.d.	n.d.
Epicatechin (EC)	n.d.	27	n.d.	n.d.
Epicatechin-3-O-gallate (ECG)	n.d.	36	n.d.	n.d.
Catechin	n.d.	4	n.d.	n.d.

n.d., not detected.

Table 3. Antimycotic activity of the *C. sinensis* extract and of ketoconazole towards yeast and yeast-like strains

Species	Strain	Diameter of inhibition area (mm)			
		<i>Camellia sinensis</i> extract (2500 µg/mL)		Ketoconazole (100 µg/mL)	
		A	B	A	B
<i>Candida albicans</i>	DBVPG 6133			56.2	60.8
<i>Candida albicans</i>	DBVPG 6157			51.6	73.6
<i>Candida glabrata</i>	DBVPG 7212	17.7	38.9		
<i>Candida glabrata</i>	DBVPG 3828	17.3	25.5		
<i>Pichia guilliermondii</i>	DBVPG 6140			54.6	64.5
<i>Candida parapsilosis</i>	DBVPG 6150			60.3	65.6
<i>Candida tropicalis</i>	DBVPG 3982				
<i>Candida zeylanoides</i>	DBVPG 6163			49.1	58.8
<i>Clavispora lusitaniae</i>	DBVPG 6142	12.9	12.0	54.2	67.7
<i>Clavispora lusitaniae</i>	DBVPG 6148	13.2	22.3	56.7	64.6
<i>Issatchenkia orientalis</i>	DBVPG 6782	16.5	17.5	29.6	31.1
<i>Kluyveromyces marxianus</i>	DBVPG 6141			56.2	72.4
<i>Saccharomyces cerevisiae</i>	DBVPG 6173	16.8	23.5		
<i>Saccharomyces cerevisiae</i>	DBVPG 6497	16.9	16.1		
<i>Saccharomyces cerevisiae</i>	DBVPG 6500	19.3	26.0		
<i>Yarrowia lipolitica</i>	DBVPG 6053			35.6	46.0
<i>Cryptococcus laurentii</i>	DBVPG 3883	17.8	17.3	39.7	45.2
<i>Cryptococcus laurentii</i>	DBVPG 4272	28.1	26.0		
<i>Cryptococcus laurentii</i>	DBVPG 6265	19.1	18.8	42.6	47.9
<i>Filobasidiella neoformans</i>	DBVPG 3428	18.8	22.0	44.8	56.6
<i>Filobasidiella neoformans</i>	DBVPG 6010	15.9	27.9	46.7	50.4
<i>Filobasidiella neoformans</i>	DBVPG 6225	16.0	17.4	36.8	50.8
<i>Filobasidiella neoformans</i>	DBVPG 6981	16.0	26.3	37.4	43.5
<i>Filobasidiella neoformans</i>	DBVPG 6982	17.4	28.0	38.4	53.4
<i>Prototheca wickerhamii</i>	DBVPG 8879	22.0	23.7		
<i>Prototheca zopfii</i>	DBVPG 8880				
<i>Prototheca zopfii</i>	DBVPG 8830				

A, test carried out on YNB + 2% glucose agar dishes; B, test carried out on YEPG agar dishes.

Table 4. Minimal inhibitory concentration (MIC) of the *C. sinensis* extract and of ketoconazole towards yeasts and yeast-like strains

Species	Strain	MIC (µg/mL)		
		<i>Camellia sinensis</i>		Ketoconazole
		Extract	Total polyphenols	
<i>Candida glabrata</i>	DBVPG 3828	1200	502	r
<i>Clavispora lusitaniae</i>	DBVPG 6148	700	293	7.5
<i>Issatchenkia orientalis</i>	DBVPG 6782	2400	1006	98.1
<i>Saccharomyces cerevisiae</i>	DBVPG 6173	2400	1006	r
<i>Cryptococcus laurentii</i>	DBVPG 6265	600	251	48.2
<i>Filobasidiella neoformans</i>	DBVPG 6010	4800	2011	48.1
<i>Prototheca wickerhamii</i>	DBVPG 8879	300	126	r

r, resistant to concentration of 100 µg/mL.

The growth of *Clavispora lusitaniae*, *Issatchenkia orientalis*, *Cryptococcus laurentii* and *Filobasidiella neoformans* strains was inhibited by *C. sinensis* extract and ketoconazole. In addition, some strains of *Prototheca wickerhamii*, *Saccharomyces cerevisiae*, *Cr. laurentii* and *Candida glabrata*, which were resistant to ketoconazole, were susceptible to *C. sinensis* extract while *Candida tropicalis* and *Prototheca zopfii* strains which are resistant to ketoconazole were also resistant to *C. sinensis* extract (Table 3).

The MICs of *C. sinensis* extract ranged from 300 to 4800 µg/mL (corresponding to 130–2010 µg/mL of total polyphenols) (Table 4). This study shows that the leaf

extract of *C. sinensis* possesses a broad antimycotic activity and, more importantly, is still active against yeast and yeast-like strains resistant to ketoconazole. As already mentioned, drug resistance is a real problem in the clinical, setting in agreement with recent literature reporting the occurrence of yeast strains able to grow in the presence of azole-derivatives (Marichal *et al.*, 1997; De Resende and De Resende, 1999; Bouchara *et al.*, 2000).

At very high concentration (25 000 µg/mL), *C. sinensis* extract caused a rapid decrease of viable cells of *Fil. neoformans* DBVPG 6010, whereas lower concentrations appeared to have an initially fungistatic phase

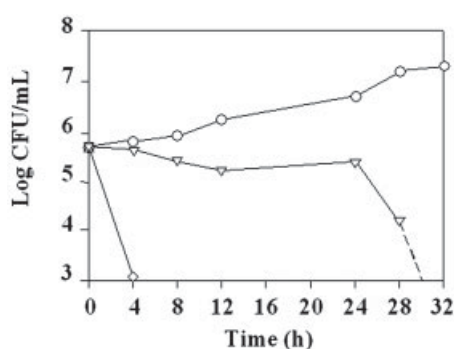


Figure 1. Assessment of fungicidal activity of the *C. sinensis* extracts towards *Fil. neoformans* DBVPG 6010. ○, control; ◇, 25 000 µg/mL; ▽, 12 500 µg/mL.

(24 h), rapidly followed by cell death (Fig. 1), the effect of the extract being dose-dependent (Fig. 2).

In order to identify the molecules involved in the antimycotic activity, the pure polyphenols present in *C. sinensis* extract composition (Table 2) were tested by ADWB. Among them only epicatechin-3-O-gallate (ECG) and epigallocatechin-3-O-gallate (EGCG) were able to inhibit the growth of *Fil. neoformans* DBVPG 6010 (data not shown).

Catechins and galloyl-derivatives of catechins are a well known class of compounds exhibiting a considerable antibacterial activity (Nisiyama and Kozaki, 1974; Ryu, 1982; Cheng-Chun *et al.*, 1999; Johns, 1999; Pieroni, 2000). Chou and Lin (1987) fractionated tea flush and found that the fraction exhibiting the strongest antimicrobial activity contained mainly EGC and ECG,

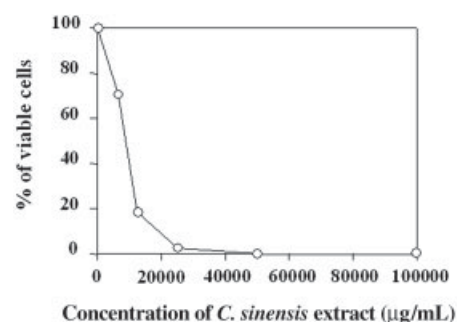


Figure 2. Dose-response effect of the *C. sinensis* extract towards *Fil. neoformans* DBVPG 6010.

whereas Hara *et al.* (1995) demonstrated that EGC, ECG and EGCG inhibit the growth of *Clostridium botulinum*. Interestingly, the green tea catechins were also able to inhibit adenovirus activity (Weber *et al.*, 2003). On the other hand, the antimycotic activity of these compounds, to our knowledge, has been very little investigated (Hirasawa and Takada, 2004). On the basis of the results of the present investigation, it is possible to point out that the broad activity of *C. sinensis* extract towards yeast and yeast-like strains may be ascribed apparently to the presence of ECG and EGCG.

Acknowledgements

The authors wish to thank Dr Valentina Caliani for her valuable help in the course of the work.

REFERENCES

- Arora DS, Ohlan D. 1997. *In vitro* studies of antifungal activity of tea (*Camellia sinensis*) and coffee (*Coffea arabica*) against wood rotting fungi. *J Basic Microbiol* **24**: 127–131.
- Atlas RM, Parks LC. 1993. *Handbook of Microbiological Media*. CRC Press: London.
- Ayoub SMH. 1989. Antimicrobial screening of Libyan medicinal plants. *Planta Med* **55**: 650–651.
- Bouchara JP, Zouhair R, Le Bodouil S *et al.* 2000. *In-vivo* selection of an azole resistant petite mutant of *Candida glabrata*. *J Med Microbiol* **49**: 977–984.
- Buzzini P, Martini A. 2000. Biodiversity of killer activity in yeasts isolated from the Brazilian rain forest. *Can J Microbiol* **46**: 607–611.
- Buzzini P, Martini A. 2001. Large-scale screening of selected *Candida maltosa*, *Debaryomyces hansenii* and *Pichia anomala* killer toxin activity against pathogenic yeasts. *Med Mycol* **39**: 479–482.
- Buzzini P, Pieroni A. 2003. Antimicrobial activity of extracts of *Clematis vitalba* towards pathogenic yeast and yeast-like microorganisms. *Fitoterapia* **74**: 347–400.
- Cassidy A, Hanley B, Lamuela-Raventos RM. 2000. Isoflavones, lignans and stilbenes: origins, metabolism and potential importance to human health. *J Sci Food Agric* **80**: 1044–1062.
- Chapman L, Johns T, Mahunnah RLA. 1997. Saponin-like *in vitro* characteristics of extracts from selected non-nutrient wild plant food additives used by Maasai in meat and milk based soups. *Ecol Food Nutr* **36**: 1–22.
- Cheng-Chun C, Lon-Leu L, King-Thom C. 1999. Antimicrobial activity of tea as affected by the degree of fermentation and manufacturing season. *Int J Food Microbiol* **48**: 125–130.
- Chou CC, Lin L. 1987. Antimicrobial activity of tea flush. *Proc World Congress of Food Technology*, Barcelona, Spain., 287–296.
- Cowan MM. 1999. Plant products as antimicrobial agents. *Clin Microbiol Rev* **12**: 564–582.
- De Resende JCP, De Resende MA. 1999. *In vitro* antifungal susceptibility of clinical isolates of *Candida* spp. from hospitalized patients. *Mycoses* **42**: 641–644.
- Etkin NL. 1996. Medicinal cuisines: diet and ethnopharmacology. *Int J Pharmacogn* **34**: 313–326.
- Hara YL, Luo SJ, Wickremasinghe RL *et al.* 1995. Use and benefit of tea. *Food Rev Int* **11**: 527–542.
- Hazen KC. 1995. New and emerging yeast pathogens. *Clin Microbiol Rev* **8**: 462–478.
- Hirasawa M, Takada K. 2004. Multiple effects of green tea catechin on the antifungal activity of antimycotics against *Candida albicans*. *J Antimicrob Chemother* **53**: 225–229.
- Johns T. 1999. Plant constituents and the nutrition and health of indigenous peoples. In *Ethnoecology. Situated Knowledge, Located Lives*, Nazarea VD (ed.). The University of Arizona Press: Tucson, 157–174.
- Kalowsky JA, Zanel GG, Klyn KA *et al.* 1997. Candidemia in Canadian tertiary care hospital from 1976–1996. *Diag Microbiol Infect Dis* **28**: 5–9.
- Krauze-Baranowska M, Cisowski W, Wiwart M *et al.* 1999. Antifungal biflavones from *Cupressocyparis leylandii*. *Planta Med* **65**: 572–573.
- Lin YM, Flavin MT, Schure R *et al.* 1999. Antiviral activities of biflavonoids. *Planta Med* **65**: 120–125.
- Mahajan V, Arora DS, Sabherwal U. 1991. Antibacterial activity of some tea samples. *Indian J Microbiol* **31**: 443–445.
- Marichal P, Vanden Bossche H, Odds FC *et al.* 1997. Molecular biological characterization of an azole-resistant *Candida glabrata* isolate. *Antimicrob Agents Chemother* **41**: 2229–2237.
- National Committee for Clinical Laboratory Standards. 1997.

- Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts*. Approved standard M27-A. National Committee for Clinical Laboratory Standards: Wayne PA.
- Nisiyama R, Kozaki M. 1974. Substances of green tea inhibiting the growth of lactic acid bacteria. *J Agric Chem* **48**: 83–89.
- Pieroni A. 2000. Medicinal plants and food medicines in the folk traditions of the upper Lucca Province, Italy. *J Ethnopharmacol* **70**: 235–273.
- Rex JH, Rinaldi MG, Pfaller MA. 1995. Resistance of *Candida* species to fluconazole. *Antimicrob Agents Chemother* **39**: 1–8.
- Riemersma RA, Rice-Evans CA, Tyrrell RM *et al.* 2001. Tea flavonoids and cardiovascular health. *Q J Med* **94**: 277–282.
- Romani A, Galardi C, Pinelli P *et al.* 2002a. HPLC quantification of flavonoids and biflavonoids in *Cupressaceae* leaves. *Chromatographia* **56**: 469–474.
- Romani A, Pinelli P, Galardi C *et al.* 2002b. Identification and quantification of galloyl derivatives, flavonoid glycosides and anthocyanins in leaves of *Pistacia lentiscus* L. *Phytochem Anal* **13**: 79–86.
- Romani A, Vignolini P, Galardi C *et al.* 2003. Polyphenolic content in different plants part of soy cultivar grown under natural conditions. *J Agric Food Chem* **51**: 5301–5306.
- Rusia K, Srivastava SK. 1988. Antimicrobial activity of some Indian medicinal plants. *Indian J Pharm Sci Jan–Feb*: 57–58.
- Ryu E. 1982. The inhibition of growth of selected bacteria by incorporating powdered tea in the medium. *Int J Zoon* **9**: 73–76.
- Sanglard D, Odds FC. 2002. Resistance of *Candida* species to antifungal agents: molecular mechanisms and clinical consequences. *Lancet Infect Dis* **2**: 73–75.
- Wang HK. 2000. The therapeutic potential of flavonoids. *Exp Opin Invest Drugs* **9**: 2103–2119.
- Warren NG, Hazen KC. 1998. *Candida*, *Cryptococcus* and other yeasts of medical importance. In *Manual of Clinical Microbiology*, Murray PR, Baron EJ, Pfaller MA, Tenover FC, Tenover RH, (eds). ASM Press: Washington DC, 1184–1198.
- Weber JM, Ruzindana-Umunyana A, Imbeault L *et al.* 2003. Inhibition of adenovirus infection and adenain by green tea catechins. *Antiviral Res* **58**: 167–173.