



Fine Regulation during Wound Healing by Mast Cells, A Physiological Role Not Yet Clarified

Stefano Bacci

Review

Research Unit of Histology and Embriology, Department of Biology, University of Florence, 50134 Florence, Italy; stefano.bacci@unifi.it

Abstract: Mast cells (MCs) are bone marrow-derived cells capable of secreting many active molecules, ranging from the mediators stored in specific granules, some of which have been known about for several decades (histamine, heparin), to small molecules produced immediately upon stimulation (membrane lipid derivatives, nitric oxide), to a host of constitutively secreted, multifunctional cytokines. With the aid of a wide array of mediators, the activated MCs control the key events of inflammation and therefore participate in the regulation of local immune response. On the basis of the structure, origin, principal subtypes, localization and function of these cells, their involvement in injury repair is therefore to be considered in acute and chronic conditions, respectively. The importance of MCs in regulating the healing processes is underscored by the proposed roles of a surplus or a deficit of their mediators in the formation of exuberant granulation tissue (such as keloids and hypertrophic scars), the delayed closure or dehiscence of wounds and the transition of acute to chronic inflammation.

Keywords: acute wounds; chronic wounds; mast cells; wound healing

1. Introduction

Mast cells (MCs) had long been elusive as to their functional role. The name itself tells us that they were first interpreted as nutrient storing cells, before being recognized as secretory cells. Later on and for a long time, emphasis on MCs secreting histamine and heparin and as effector cells of immediate type hypersensitivity had almost completely distracted from other possible roles of MCs in health and disease. The expanded knowledge on the structure, origin, and function of these cells has brought them to the front of stage of the injury response and repair processes through the release of histamine, glycosaminoglycans, enzymes, cytokines, arachidonic acid derivatives and nitric oxide and, perhaps, through direct, membrane molecule-mediated cell interactions.

2. Structure of Mast Cells

Von Recklinghausen (1863) probably observed what later came to be called MCs, however his description did not lead to their identification as a specific cell type.

Ehrlich (1878), identified MCs based on the metachromatic staining of their cytoplasmic granules in connective tissue. In other words, the granules are chromotropic. These cells, which are often found near vessels and nerves, are of various shapes (Figure 1a). The granules (Figure 1b) are delimited by a membrane and contain a paracrystalline matrix with lamellar or spiral and volute matrices. Preformed mediators stored in specific granules can be released from morphologically distinct secretion types (reviewed and modified by [1–6]).

Citation: Bacci, S. Fine regulation during wound healing by mast cells, a physiological role not yet clarified. *Int. J. Mol. Sci.* **2022**, *23*, 1820. https:// doi.org/10.3390/ijms23031820

Academic Editor: Aleksandra Klimczak

Received: 9 December 2021 Accepted: 4 February 2022 Published: 5 February 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/license s/by/4.0/).

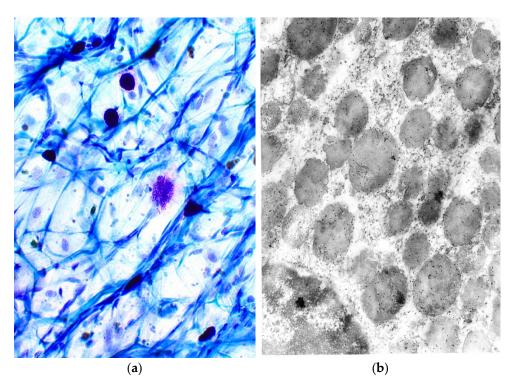


Figure 1. (a) Metachromatic properties of MC granules. Light microscopy, Toluidin blue, magnification × 40; (b) localization of tryptase in MC granules. Electron microscopy, magnification × 2500.

3. Origin and Survival of Mast Cells

Mature MCs are the progeny of multipotent hematopoietic stem cells that commit to the MC lineage already in the bone marrow. The progenitors are positive for CD13, C-Kit, and CD34 but negative for FceRI [7]. The secretion of TNF-alpha by other CD11b+ bone marrow cells is critical to the development and expansion of the MC lineage and some yet unidentified action exerted by bone marrow DC on MC precursors is needed to prime these precursors for appropriate homing in peripheral tissues, at least in mice [8–10]. Committed progenitors pass into the circulation to complete their maturation within peripheral tissues. The adhesion to endothelial cells is mediated by α -4 integrins, VCAM1 and E-selectin [11]. The migration of MC precursors into tissues is stimulated by SCF and by eotaxin; the latter interacts with the chemokine receptors CXCR2, CCR3, CXCR4 and CCR5. In the tissues, further proliferation and differentiation of MC precursors depend on the presence of the local growth factors and cytokines, SCF, IL3, IL4, IL6, IL9 and NGF [12,13]

The SCF and C-Kit signaling system is crucial for MC growth and development and the injection of SCF into the skin of humans results in local accumulation of MCs. MCs can quickly move within connective tissue and even transfer into epithelia and backwards. Histamine itself promotes MC migration [12,13].

NGF has been reported to be a survival factor for rat peritoneal MCs and for human cord blood-derived MCs in the presence of SCF. Interleukin3 induces mouse MC growth and enhances their development in response to SCF in vitro; in humans, IL3 receptor is expressed by MC progenitors and intestinal MCs but not by MCs from lung, uterus, kidney, tonsils and skin [14]. The survival, adhesion to fibronectin and cytokine production of human umbilical cord blood-derived MCs are stimulated by IL33; a possible role of adhesion receptors in MC survival has also been proposed. Human MCs express functional TRAIL receptors and SCF prevents their apoptosis induced by TRAIL receptor ligation; the survival of human mature MCs in tissues depends on the local production of SCF since its withdrawal results in apoptosis [6,9,15–17]

4. Mast Cell Heterogeneity

Mouse connective tissue MCs and mucosal MCs differ in their location, mediator content and responsiveness to activating stimuli. In humans, some subtypes of these cells were found: MCT were present in the lungs and intestinal mucosa and MCTC were found in the skin. Finally a population of MCc was also observed in humans; its features have not yet been fully elucidated. However is known that MC types can interchange with each other and this phenomena is determined by the microenvironment. The main differences between MCT and MCTC in humans are reported in Table 1 (reviewed and modified by [1,6,9,18–20]).

	МСт	МСтс.
Shape	Small, sparsely granulated	Large, densely granulated
Enzyme content of granules	Tryptase and others	Chymase, tryptase, and others
Response to compound 48/80	Proliferative	Secretory
Sensitivity to disodium-cromoglycate	None	Yes
Life span	Short	Long
Preferential location in humans	Lung, bowel	Skin, bowel
Antigens	CD50	CD32
Cytokines	GM-CSF, TGF-β	IL3
Complement receptor	None	CD88 (C5aR)

Table 1. Differences in some properties of MCT and MCTC.

Modified by Bacci, S., Bonelli, A., Romagnoli P. Mast cells in injury response. In Cell movement: New Research Trends. Abreu, T., Silva, G. Eds. Nova Science Publishers, Inc., Hauppage, NY, USA, 2009, pp. 81–121.

5. Mast Cell Relationships to Basophils

MCs and basophils are two distinct cell populations, which exhibit some common features including histamine and heparin secretion and share a common precursor up to an advanced stage of differentiation.

The main differences between MCs and basophils are summarized in Table 2 (reviewed and modified by [6,9,17,20–25].

Table 2. Differential features of basophils and MC in	n humans.
---	-----------

	Basophils	МС
Diameter	8-9 μm	13–14 μm
Granules	Few	Numerous
Compound exocytosis	Rare	Common
Tryptase in granules	No	Yes
Expression of C-Kit	No	Yes

Modified by Bacci, S., Bonelli, A., Romagnoli P. Mast cells in injury response. In cell movement: New Research Trends. Abreu, T., Silva, G. Eds. Nova Science Publishers, Inc., Hauppage, NY, USA, 2009, pp. 81–121.

6. Mast Cell Activation

The best-studied signal transduction pathways are those started by the activation of FccRI; in particular, early stimulation involves interaction with other Kinases such as LYN. LYN activates SYK, LAT as well as NTAL: the block of both LAT and NTAL pathways leads to abolition of calcium and degranulation upon FccRI stimulation. The mobilization of cytokines also starts from LAT with activations of factors including VAV56 (a tyrosine phosphorylation-regulated signal transduction molecule) and phosphorylation of MAPK. These molecules in turn, activate FOS and JUN transcription factors leading to cytokine gene activation. However, many of the pathways controlling

FccRI-induced MC activation have been described in detail (reviewed and modified by [6,9,17,26–28]).

7. The Secretory Products of Mast Cells

The granules of MCs contain glycosaminoglycans, neutral hydrolases, other enzymes and chemotactic factors which are secreted by the MCs by exocytosis under stimulation. Following the activation of the complement, MCs secrete numerous substances that promote vasodilation, the permeation of plasma proteins into the tissues, leukocyte diapedesis and the recruitment, differentiation and function of lymphocytes, macrophages and fibroblasts [1,6,9,29-32]. In the context of acquired immunity, the secretory products of MCs are released following antigen-antibody reactions. In addition to the substances contained in the chromotropic granules and known for a long time, MCs secrete various growth factors and cytokines. Most likely these molecules are not contained in chromotropic granules, except $TNF\alpha$ but are contained in small vesicles, secreted by exocytosis without being stored in the cytoplasm. Therefore, the regulation of cytokine secretion depends on that of the synthesis of these substances itself, with time intervals between the minute and the hour between the stimulation of the cell and the increase in secretion. MCs synthesize and release VIP and a somatostatin-like peptide. It has not yet been clarified whether the latter two substances are contained in chromotropic granules, or in other types of granules [1,6,9,26,29-36]. MCs also secrete products that are not contained in granules, but are directly synthesized under stimulation starting from membrane phospholipids; prostaglandins, leukotrienes, thromboxanes and plateletactivating factors. Finally, MCs secrete NO that modulates the release of histamine by the MCs themselves. However a list of the principal secretory products (in the tissue and in cell lines) is beyond the scope of this review. (For other details see the following references: [26-36]).

8. Mast Cells as Effector Cells in the Immune System

MCs participate in the processes of natural immunity, as they degranulate in response to stimuli of various types, including among other things the activation of complement through the alternative pathway, possess killer activities under certain conditions and are cytotoxic to helminths (reviewed and modified by [1,6,9,37]).

MCs can participate in the processes of acquired immunity, and the effector role they play in allergic reactions has long been known. Their secretory products determine vasodilation and influence the recruitment, differentiation and function of lymphocytes, macrophages, fibroblasts and MCs themselves. Therefore, MCs may also be important in the triggering of delayed-type hypersensitivity reactions and in the late stages of inflammation including fibrosis. The secretory products of MCs determine dilation and increase the permeability of capillary activation of coagulation, activation of fibrinolysis and a stimulus adhesion of leukocytes to the endothelium of blood vessels [Reviewed and modified by 1,6,9,37]. Furthermore, some MC secretory products stimulate the proliferation and secretory activity of fibroblasts and histamine can also affect other cell types involved in immune processes. The major effects of some mediators are summarized in Table 3.

 Table 3. Involvement of some mediators secreted by mast cells in the response of the immune system.

Mediators	Functions
Histamine	Activation of a group of suppressor T cells
Prostaglandin D2	Inhibition of the activity of helper T cells, stimulation of the differentiation and function of
	suppressor T cells, inhibition of IgE production
Leukotrienes	Similar function of prostaglandin D2 and inhibition of the differentiation of plasma cells.

VIP	Inhibition of the secretory and proliferative responses of at least some of the subgroups of T and B cells.
Heparin (low	Activation of macrophages to produce IL-1, which in turn affects both the macrophages
concentration)	themselves and the surrounding cells and the whole organism
	Modified by Bacci, S., Bonelli, A., Romagnoli P. Mast cells in injury response. In cell movement:

New Research Trends. Abreu, T., Silva, G. Eds. Nova Science Publishers, Inc, Hauppage, NY, 2009 pp 81–121. Bacci, S., Mastociti e cellule dendritiche della cute e delle mucose: analisi morfologiche e funzionali in condizioni normali e sperimentali. (Mast cells and dendritic cells of the skin and mucous membranes: morphological and functional analyses in normal and experimental conditions (Doctorate, PhD thesis, University of Florence, 1997).

9. The Wound Healing

The process takes place in distinct phases, but in some moments may overlap, preceded by a preliminary hemostatic phase. Below are the main events of the various phases that characterize this event.

Hemostatic phase: represents the local response to hemorrhage, caused by the rupture of blood vessels, through the action of thrombocytes and the activation of tissue coagulation factors. This phase is characterized by the formation of a clot, a structure consisting of a fibrin network in which the corpuscular elements of the blood which occupy the wound remain imprisoned. [38–46].

Inflammatory phase: in the case of the wound, inflammation provides for the elimination of the microbial agent, any foreign bodies and necrotic cells, but also the activation of those factors that are at the basis of the subsequent proliferative processes and, therefore, of the repair or replacement of damaged tissue. It involves vasodilation and plasma exudation and the proliferation of macrophages, mononuclear cells with phagocytic capacity, which together with neutrophil granulocytes clean the wound. The inflammatory reaction begins immediately after the trauma and lasts a few days, also persisting during the next phase. In this period the wound is edematous and strongly red [38–46].

Proliferative phase: this begins a few hours after the injurious event and has the purpose of replacing the clot with a solid, definitive structure. In fact, 24–72 h after the trauma, an important proliferation of fibroblasts guarantees the secretion of hyaluronic acid, an active component in the formation of collagen fibers. Fibroblasts already at the end of the first week represent almost all the cells present in the wound; their activity will continue for the time necessary for the collagen produced to fill the wound. At this point, having completed their task, around the third week, the fibroblasts are activated and acquire α -SM actin expression and become myofibroblasts. These myofibroblastic cells synthesize and deposit the ECM components that eventually replace the provisional matrix. Simultaneously with the others, the proliferation of cells of the basal layer of the epithelium also begins. [38–46].

Maturation phase: corresponds to that phase in which the wound, initially edematous and reddened, is permanently closed by a scar with very different characteristics: pale, smooth, inelastic, without skin appendages with reduced spraying and innervation. This phase lasts at least three weeks, but sometimes it also continues for months or years [38,40–46].

10. Mast Cells and Wound Repair

Weller et al., 2006 [47], studied experimentally induced skin wounds in MC-deficient KitW/KitW-v mice, normal Kit+/+ mice, and MC-reconstituted KitW/KitW-v mice. Wound closure was significantly impaired in the absence of MCs during the first 6 days of wound healing and histomorphometric analyses of MC degranulation at the wound edge revealed distance-dependent MC activation. In addition, KitW/KitW-v mice showed impaired extravasation and recruitment of neutrophils to the wounded areas. Notably, wound closure, extravasation, and neutrophil recruitment were found to be normal in

MC-reconstituted KitW/KitW-v mice. Besides, wound closure was reduced in mice treated with an H1-receptor antagonist but not after treatment with an H2-receptor antagonist or in the absence of TNF-alpha. Other authors have shown that the MC products histamine and serotonin exert mitogenic effects on murine epidermal keratinocytes in situ [48]. In vitro studies have demonstrated that MC can promote the conversion of fibroblasts to a myofibroblast phenotype [49], as well as stimulate fibroblast proliferation and migration [50] and that MCs, which accumulate at sites of neovascularization, can induce neoangiogenesis [51]. These findings undoubtedly indicate that MCs are involved in wound healing. In particular, Egozi et al. [52] demonstrated that MCs are able to control the key events of wound healing as an inflammatory response. However other authors evoked for MCs a role in the revascularization of damaged tissue, re-epithelialization, and deposition and subsequent remodeling of connective tissue [53,54]. Once activated by tissue injury, MCs release mediators which induce vasodilation and increase vascular permeability [47]. The endothelial cells, in turn, influence the functional state of MCs by releasing SCF, IL3 and thrombin which enhance migration, proliferation and local differentiation of the MCs. At the edge of wound, keratinocytes can secrete several cytokines and LL37, which influence MC recruitment and function [6].

MC numbers and the degranulation index increase at the border of a wound within a maximum of 1–3 h from trauma and decrease thereafter, becoming less than baseline values after 6 h, or more. The quick variation in MC numbers and location within connective tissue (15 min for trauma, in the skin) speaks against the recruitment of precursors and differentiation of new MCs as a relevant mechanism in this phase of the response to injury [55]. These cells increase again in number later on, to a maximum of 10 days, and decrease thereafter returning to control values after 21 days from wounding. The late increase correlates with the upregulation of MCP1 and with the production of TGF-beta, which is also a potent chemoattractant for MCs [56].

Oemichen et al. have confirmed that MCs degranulate at the border of skin wounds within 60 min, as estimated by enzyme histochemistry for a granule-bound esterase [57]. Molecules of MC origin, such as TNF-alpha, which has already been released in the first 15 min after wounding, and histamine, which is released even earlier, favor the adhesion of leukocytes to vessel by increasing the expression of several endothelial adhesion molecules [58]. Leukotrienes, proteases and cytokines, released by MC, represent chemotactic signals for neutrophils, basophils and eosinophils. Tryptase and cathepsin G also regulate endothelial–leukocyte interactions and leukocyte behavior at sites of inflammation. Chymase can induce eosinophils to express chemokines for neutrophils (reviewed by [6,59,60]). As the inflammatory process develops, MCs themselves can contribute directly to regulating these phases of injury response. The responses of fibroblasts may be regulated by TGF-beta, TNF-alpha and proteases [49].

Angiogenesis is stimulated by TGF-beta, VEGF, chymase and tryptase. In contrast, heparin may inhibit angiogenesis by interacting with, and inhibiting pro-angiogenetic factors [61–65]. In the late phases of repair, MC-derived growth factors and cytokines may influence the phenotype of the activated fibroblasts inducing the appearance of myofibroblasts which ensure the changeover from fibroplasia to contraction and final healing of the wound [6,49]. Concerning tissue remodeling, MCs can activate fibroblasts, promoting collagen synthesis: this effect may be partly due to tryptase, which has been shown to stimulate the synthesis of type I collagen in human dermal fibroblasts [66,67].

Besides skin, MCs produce and release potent proteolytic enzymes, such as matrix metalloproteinases by initiating the degradation of extracellular matrix. Finally, it has been demonstrated that cutaneous remodeling is modulated by skin MCs and MC-derived histamine. In fact, the inhibition of MC histamine synthesis in wounded rats has been shown to decrease the hydroxyproline content of granulation tissue, to delay epithelization, and to reduce wound-breaking strength [6,15,40,46,52,67].

The questions arises as to how MCs are stimulated to move into and within tissue. Possible sources of stimuli early upon injury are the extravasated platelets and autochthonous cells of the damaged tissue itself, in particular, endothelial cells, nerve fibers and the epidermis, as indicated by the ordered approach of MCs to this tissue upon selective photoinduced damage. The late, new increase in numbers is presumably dependent upon influx of precursors and their differentiation under stimuli provided by macrophages and possibly other cells in the site of inflammation. The final reduction to baseline numbers and activity may simply derive from the lack of further stimulation as the repair proceeds and macrophages themselves disappear, however, the possible existence of active mechanisms to switch off the inflammatory process, instead of a simple lack of stimulation, has yet to be addressed. Impairment of such a switch off, or persistent activation of stimulating mechanisms may be responsible for the persistence and protracted activation of high numbers of MCs, leading to fibrosis (for other details see [6,67]).

11. Mast Cells and Scars

MCs are likely candidates to play a role in the etiology of hypertrophic scar formation [68]. It has been reported that early cutaneous wounds express low levels of inflammation and can heal without a scar and can regenerate hair follicles. In contrast, wounds in the late fetal developmental stages have high levels of inflammation and heal with a fibrotic scar. In a mouse fetal repair model study where embryonic day E15 wounds healed without a scar and E18 wounds healed with a scar, it has been demonstrated that there were fewer MCs, which were less mature, and there was no degranulation upon wounding in the scarless E15 wounds compared to the fibrotic wounds produced at E18 [69–72]. In adult mouse skin, some experimental evidence has shown that MCs augment fibroblast activity, which is accountable for collagen deposition and remodeling in the scar formation/remodeling phase of repair. Additionally, other studies have suggested that MCs may affect collagen maturation and remodeling more than collagen production. While the majority of animal studies have suggested that MCs promote scar formation, some studies have suggested that MC-deficient mouse strains heal with similar granulation tissue and scar size compared to normal mice (reviewed by [71]). In addition, the inhibition of MC activity, with MC inhibitor disodium cromoglycate (DSCG), caused a significant decrease in scar width along with accelerated collagen re-organization. Despite the reduced scar width, DSCG treatment did not affect the breaking strength of the healed tissue. These results indicate that blockade of mast cell activation reduces scar formation and inflammation without further weakening the healed wound [72,73].

In man, the greatest increase in numbers of MCs at week 1 compared to uninjured skin with a gradual decrease from week 1 to week 8 has been demonstrated. These findings were similar to those of markers of angiogenesis as well as of CD8+cells; furthermore, macrophages slowly increased from uninjured skin and showed greatest levels at week 5 where levels reduced thereafter [71–73]. It has been demonstrated that MCs, after stimulation with substance P, activate fibroblasts through the release of histamine that is significantly elevated in the plasma of patients developing hypertrophic scars compared with age-matched normal volunteers. Since MCs are able to promote the proliferation of fibroblasts also by the release of TGF-beta1, TNF-alpha and IL-4, this indicates that MCs may play a role in hypertrophic scar formation via different mediators [71–73]

12. Mast cells and Chronic Wounds

Various pathophysiologies of wounds that do not follow the normal healing process and have a significant delay in healing receive the clinical designation of a chronic wound [74]. The incidence of chronic wounds is estimated to be about 1–2% in developed countries [75]. In particular in chronic wounds, MCs have a significantly different functional repertoire and location than in acute healing. It has been observed that MCs increase their number and degranulation index in chronic wounds other than expressing TNF α , as well as SCF and the receptor C-Kit [76–78] Signaling through C-Kit induces degranulation, promotes migration differentiation of new precursors, and prevents apoptosis, thereby potentially contributing to the chronic inflammatory microenvironment [76–78].

13. Neurogenic Stimuli and Mast Cells in Chronic Would Healing

Immune system activity can be modulated by the nervous system; this close correlation is also documented in healing wounds. Moreover, experimental observations suggest that neurogenic stimuli profoundly affect wound repair after injury and that delayed wound healing is observed in animal models after surgical resection of cutaneous nerves. All these observations clearly suggest that innervation and neuromediators play a pivotal physiological role in wound healing. Interactions between the nerves and other cells involved in wound healing, such as MCs, are crucial in the healing process and the expression in their cytoplasm of NGF and VIP clearly demonstrate this hypothesis and these mediators are able to interact with neurons and nerve fibers of the dermis, thus obtaining an improvement. The activation of nerve fibers could in turn be related to other phenomena in chronic wounds such as the increased secretion of extracellular matrix by fibroblasts as has been observed previously in the increase of TGF-beta and the response of cellular infiltrates [78–80]. Further work will undoubtedly be necessary to clarify these pathological disease states and potential mechanisms to aid in their resolution.

14. Conclusions

MCs can be proposed for a double role in injury response. Very early, they release different types of mediators activating the vascular phase of inflammation and the recruitment of leukocytes which provide for the cellular phase of inflammation itself. To these aims, MCs most probably concur with other ready-to-fire local control systems, i.e., platelets (which are activated immediately upon endothelial damage) and peripheral nerve fibers, in particular sensitive fibers which are involved in axo-axonal reflexes and secrete peptide mediators. In the skin, but not necessarily in other organs, MCs gather near the site of injury to perform their roles. In the intermediate phase of the response, MCs decrease in number, probably in part because they degranulate and so become undetectable by the usual histochemical methods, in part because they die in response to hyperstimulation and toxic substances (e.g., high concentrations of NO and other oxidants). The new, late increase in numbers and the late activity of MCs can concur to drive definitive tissue repair by stimulating angiogenesis and inducing fibroblasts to secrete extracellular matrix and to differentiate into myofibroblasts to contract the collagen matrix. MCs promote the proliferation of fibroblasts, endothelial cells, and keratinocytes during the proliferative phase of wound healing. In the mouse, histamine and serotonin (which is also secreted by rodent MCs) exert mitogenic effects on murine epidermal keratinocytes in situ and therefore influence re-epithelialization. In conclusion, if the news regarding the role of MCs in acute wounds begins to flow and, therefore, authorizes the reader to have ideas about the possible role of these cell types, the picture, however, turns out to be extremely confused and contradictory for the role assumed by these cells in chronic wounds. Concerning therapies, it would also be important to assess if histamine or TGF-beta induction, through other systemic or topical treatments, could be of use in chronic wound management. Therefore, the inducing of the TGF-beta or histamine hypothesized beneficial effects could be a further, intriguing option to face the urgent, unmet clinical needs in chronic wound management. Concerning the role of MCs in chronic wounds, from what has been reported in the review, this remains elusive and therefore should now be considered as a possible objective for this type of research [81].

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: I would like to extend my sincere thanks to the editorial team of International Journal of Molecular Science for continuous and efficient support during the development and publication of this manuscript.

Conflicts of Interest: The author declares no conflict of interest

Abbreviations

- MCs Mast cells
- FceRI High-affinity IgE receptor
- TNF Tumor necrosis factor
- VCAMVascular cell adhesion molecule
- SCF Stem cell factor
- NGF Nerve growth factor
- TRAIL TNF-related apoptosis inducing ligand
- MCT MCs containing only tryptase
- MCTC MCs containing both tryptase and chymase
- MCc MCs containing only chymase
- TGF Transforming growth factor
- CD Cluster of differentiation
- IL Interleukin
- LYN Lck/yes-related tyrosine kinase
- SYK Spleen associate tyrosine kinase
- LAT Linker for activation of T cells
- NTAL Non-T cell activation linker
- VIP Vaso active intestinal peptide
- NO Nitric oxide
- FGF Fibroblast growth factor
- MCP-1Monocyte chemoattractant protein 1
- TGF Transforming growth factor
- VEGF Vascular endothelial growth factor

References

- 1. Krystel-Whittemore, M.; Dileepan, K.N.; Wood, J.G. Mast Cell: A Multi-Functional Master Cell. *Front Immunol.* **2016**, *6*, 620. https://doi.org/10.3389/fimmu.2015.00620.
- 2. Ribatti, D. The Staining of Mast Cells: A Historical Overview. Int. Arch. Allergy Immunol. 2018, 176, 55-60. https://doi.org/10.1159/000487538.
- Galli, J.S.; Gaudenzio, N.; Tsai, M. Mast cells in inflammation and disease: Recent progress and ongoing concerns. *Annu. Rev. Immunol.* 2020, 38, 49–77. https://doi.org/10.1146/annurev-immunol-071719-094903.
- 4. Saccheri, P.; Travan, L.; Ribatti, D.; Crivellato, E. Mast cells, an evolutionary approach. *Ital. J. Anat. Embryol.* **2020**, *124*, 271–287. https://doi.org/10.13128/ijae-11657.
- 5. Fong, M.; Crane, J.S. Histology, Mast cells. In *StatPearls [Internet]*; StatPearls Publishing: Treasure Island, FL, USA, 2021. Available online: https://www.ncbi.nlm.nih.gov/books/NBK499904 (accessed on:: 04.02.2022).
- Bacci, S.; Bonelli, A.; Romagnoli, P. Mast cells in injury response. In *Cell Movement: New Research Trends*; Abreu, T., Silva, G., Eds.; Nova Science Publishers, New York, NY, USA, 2009; pp. 81–121.
- 7. Kirshenbaum, A.S.; Goff, J.P.; Semere, T.; Foster, B.; Scott, L.M.; Metcalfe, D.D. Demonstration that human mast cells arise from a progenitor cell population that is CD34(+), C-Kit(+), and expresses aminopeptidase N (CD13). *Blood* **1999**, *94*, 2333–2342.
- 8. Wright, H.V.; Bailey, D.; Kashyap, M.; Kepley, C.L.; Drutskaya, M.S.; Nedospasov, S.A.; Ryan, J.J. IL-3-mediated TNF production is necessary for mast cell development. *J. Immunol.* **2006**, *176*, 2114–2121.
- 9. Bacci, S.; Romagnoli, P. Drugs acting on mast cells function. A cell biological perspective. *Inflamm. Allergy Drug Targets* 2010, 9, 214–228.

- Alcaide, P.; Jones, T.G.; Lord, G.M.; Glimcher, L.H.; Hallgren, J.; Arinobu, Y.; Akashi, K.; Paterson, A.M.; Gurish, M.A.; Luscinskas, F.W. Dendritic cell expression of the transcription factor T-beta regulates mast cell progenitor homing to mucosal tissue. J. Exp. Med. 2007, 204, 431–439. https://doi.org/10.1084/jem.20060626.
- Gentek, R.; Ghigo, C.; Hoeffel, G.; Bulle, M.J.; Msallam, R.; Gautier, G.; Launay, P.; Chen, J.; Ginhoux, F.; Bajenoff, M. Hemogenic endothelial fate mapping reveals dual developmental origin of mast cells. *Immunity* 2018, 48, 1160–1171. https://doi.org/10.1016/j.immuni.2018.04.025.
- 12. Ochi, H.; Hirani, W.M.; Yuan, Q.; Friend, D.S.; Austen, K.F.; Boyce, J.A. T helper cell type 2 cytokine-mediated comitogenic responses and CCR3 expression during differentiation of human mast cells in vitro. *J. Exp. Med.* **1999**, *190*, 267–280.
- 13. Nakahata, T; Toru, H. Cytokines regulate development of human mast cells from hematopoietic progenitors. *Int. J. Hematol.* **2002**, *75*, 350–356.
- Kanbe, N; Kurosawa, M; Miyachi, Y; Kanbe, M; Saitoh, H; Matsuda, H. Nerve growth factor prevents apoptosis of cord bloodderived human cultured mast cells synergistically with stem cell factor. *Clin. Exp. Allergy* 2000, 30, 1113–1120.
- 15. Berent-Maoz, B.; Piliponsky, A.M.; Daigle, I.; Simon, H.U.; Levi-Schaffer, F. Human mast cells undergo TRAIL-induced apoptosis. J. Immunol. 2006, 176, 2272–2278.
- 16. Okayama, Y.; Kawakami; T Development, migration, and survival of mast cells. *Immunol. Res.* 2006, 34, 97–115. https://doi.org/10.1385/IR:34:2:97.
- 17. da Silva, E.Z.; Jamur, M.C.; Oliver, C. Mast cell function: A new vision of an old cell. J. Histochem. Cytochem. 2014, 62, 698–738. https://doi.org/10.1369/0022155414545334.
- 18. Crivellato, E.; Feltrami, C.A.; Mallardi, F.; Ribatti, D. The mast cell: An active participant or an innocent bystander? *Histol. Histopathol.* **2004**, *19*, 259–270. https://doi.org/10.14670/HH-19.259.
- Elieh Ali Komi, D.; Wöhrl, S.; Bielory, L. Mast Cell Biology at Molecular Level: A Comprehensive Review. *Clin. Rev. Allergy Immunol.* 2020, 58, 342–365. https://doi.org/10.1007/s12016-019-08769-2.
- Varricchi, G.; Pecoraro, A.; Loffredo, S.; Poto, R.; Rivellese, F.; Genovese, A.; Marone, G.; Spadaro, G. Heterogeneity of human mast cells with respect to MRGPR2 receptor expression and function. *Front. Cell. Neurosci.* 2019, 13, 299. https://doi.org/10.3389/fncel.2019.00299.
- Kocabas, C.N.; Yavuz, A.S.; Lipsky, P.E.; Metcalfe, D.D.; Akin, C. Analysis of the lineage relationship between mast cells and basophils using the C-Kit D816V mutation as a biologic signature. J. Allergy Clin. Immunol. 2005, 115, 1155–1161. https://doi.org/10.1016/j.jaci.2005.02.030.
- Crivellato, E.; Travan, L.; Ribatti, D. Mast cells and basophils: A potential link in promoting angiogenesis during allergic inflammation. *Int. Arch. Allergy Immunol.* 2010, 151, 89–97. https://doi.org/10.1159/000235998.
- 23. Voehringer, D. Protective and pathological roles of mast cells and basophils. *Nat. Rev. Immunol.* 2013, 13, 362–375. https://doi.org/10.1038/nri3427.
- 24. Kubo, M. Mast cells and basophils in allergic inflammation. *Curr. Opin. Immunol.* 2018, 54, 74–79. https://doi.org/10.1016/j.coi.2018.06.006.
- 25. Varricchi, G.; Raap, U.; Rivellese, F.; Marone, G.; Gibbs, B.F. Human mast cells and basophils-How are they similar how are they different? *Immunol. Rev.* 2018, 282, 8–34. https://doi.org/10.1111/imr.12627.
- 26. Moon, T.C.; Befus, A.D.; Kulka, M. Mast cell mediators: Their differential release and the secretory pathways involved. *Front. Immunol.* **2014**, *5*, 569. https://doi.org/10.3389/fimmu.2014.00569.
- 27. Yu, Y.; Blokhuis, B.R.; Garssen, J.; Redegeld, F.A. Non-IgE mediated mast cell activation. *Eur. J. Pharmavol.* 2016, 778, 33–43. https://doi.org/10.1016/j.ejphar.2015.07.017.
- Theoharides, T.C.; Tsilioni, I.; Ren, H. Recent advances in our understanding of mast cell activation—Or should it be mast cell mediator disorders? *Expert Rev. Clin. Immunol.* 2019, 15, 639–656. https://doi.org/10.1080/1744666X.2019.1596800.
- Castells, M.C. Mast cells mediators in allergic inflammation and mastocytosis. *Immunol. Allergy Clin. North Am.* 2006, 26, 465–485. https://doi.org/10.1016/j.iac.2006.05.005.
- González-de-Olano, D.; Álvarez-Twose, I. Mast Cells as Key Players in Allergy and Inflammation. J. Investig. Allergol. Clin. Immunol. 2018, 28, 365–378. https://doi.org/10.18176/jiaci.0327.
- Monument, M.J.; Hart, D.A.; Salo, P.T.; Befus, A.D.; Hildebrand, K.A. Neuroinflammatory Mechanisms of Connective Tissue Fibrosis: Targeting Neurogenic and Mast Cell Contributions. *Adv. Wound Care* 2015, 4, 137–151. https://doi.org/10.1089/wound.2013.0509.
- 32. Wernersson, S.; Pejler, G. Mast cell secretory granules: Armed for battle. *Nat. Rev. Immunol.* 2014, 14, 478–494. https://doi.org/10.1038/nri3690.
- Pastwińska, J.; Agier, J.; Dastych, J.; Brzezińska-Błaszczyk, E. Mast cells as the strength of the inflammatory process. *Pol. J. Pathol.* 2017, 68, 187–196. https://doi.org/10.5114/pjp.2017.71526.
- 34. Xu, H.; Bin, N.R.; Sugita, S. Diverse exocytic pathways for mast cell mediators. *Biochem. Soc. Trans.* 2018, 46, 235–247. https://doi.org/10.1042/BST20170450.
- Agier, J.; Pastwińska, J.; Brzezińska-Błaszczyk, E. An overview of mast cell pattern recognition receptors. *Inflamm. Res.* 2018, 67, 737–746. https://doi.org/10.1007/s00011-018-1164-5.
- 36. Castells, C.M.; Bankova, L. Mast Cells: Surface Receptors and Signal Transduction, 2021. Uptodate. Available online: https://www.uptodate.com/contents/mast-cells-surface-receptors-and-signal-transduction (accessed on: 04.02.2022).

- Varricchi, G.; Rossi, F.W.; Galdiero, M.R.; Granata, F.; Criscuolo, G.; Spadaro, G.; de Paulis, A.; Marone, G. Physiological Roles of Mast Cells: Collegium Internationale Allergologicum Update 2019. *Int. Arch. Allergy Immunol.* 2019, 179, 247–261. https://doi.org/10.1159/000500088.
- 38. Singer, A.J.; Clark, R.A. Cutaneous wound healing. New Eng. I. Med. 1999 341, 738-746. https://doi.org/10.1056/NEJM199909023411006.
- 39. Darby, I.A.; Laverdet, B.; Bonté; F; Desmoulière, A. Fibroblasts and myofibroblasts in wound healing. *Clin. Cosmet. Investig. Dermatol.* **2014**, *7*, 301–311. https://doi.org/10.2147/CCID.S50046.
- Gonzalez, A.C.; Costa, T.F.; Andrade, Z.A.; Medrado, A.R. Wound healing A literature review. An. Bras. De Dermatol. 2016, 91, 614–620. https://doi.org/10.1590/abd1806-4841.20164741.
- Cañedo-Dorantes, L; Cañedo-Ayala, M. Skin Acute Wound Healing: A Comprehensive Review. Int. J. Inflam. 2019, 2019,3706315. https://https://doi.org/10.1155/2019/3706315.
- Rodrigues, M.; Kosaric, N.; Bonham, C.A.; Gurtner, G.C. Wound Healing: A Cellular Perspective. *Physiol. Rev.* 2019, 99, 665–706. https://doi.org/10.1152/physrev.00067.2017.
- Visha, M.G.; Karunagaran, M. A review on wound healing. Int. J. Clinicopathol. Correl. 2019, 3, 50–59. https://doi.org/10.4103/ijcpc.ijcpc_13_19.
- 44. Bacci, S. Cutaneous wound healing: Cellular mechanisms and therapies (an update). Med. Res. Arch. 2019, 7, 12.
- 45. Tottoli, E.M.; Dorati, R.; Genta, I.; Chiesa, E.; Pisani, S.; Conti, B. Skin Wound Healing Process and New Emerging Technologies for Skin Wound Care and Regeneration. *Pharmaceutics* **2020**, *12*, 735–765. https://doi.org/10.3390/pharmaceutics12080735.
- Wilkinson, H.N.; Hardman, M.J. Wound healing: Cellular mechanisms and pathological outcomes. *Open Biol.* 2020, 10, 200223–200236. https://doi.org/10.1098/rsob.200223.
- 47. Weller, K.; Foitzik, K.; Paus, R.; Syska, W.; Maurer, M. Mast cells are required for normal healing of skin wounds in mice. *FASEB J.* **2006**, *20*, 2366–2368. https://doi.org/10.1096/fj.06-5837fje.
- Maurer, M.; Opitz, M.; Henz, B.M.; Paus, R. The mast cell products histamine and serotonin stimulate and TNF-alpha inhibits the proliferation of murine epidermal keratinocytes in situ. *J. Dermatol. Sci.* 1997, 16, 79–84. https://doi.org/10.1016/s0923-1811(97)00043-1.
- Gailit, J.; Marchese, M.J.; Kew, R.R.; Gruber, B.L. The differentiation and function of myofibroblasts is regulated by mast cell mediators. J. Investig. Dermatol. 2001, 117, 1113–1119. https://doi.org/10.1046/j.1523-1747.2001.15211.x.
- 50. Levi-Schaffer, F.; Kupietzky, A. Mast cells enhance migration and proliferation of fibroblasts into an in vitro wound. *Exp. Cell. Res.* **1990**, *188*, 42–49. https://doi.org/10.1016/0014-4827(90)90275-f.
- 51. Norrby, K.; Jakobsson, A.; Sörbo, J. Mast-cell-mediated angiogenesis: A novel experimental model using the rat mesentery. *Virchows Arch. B* **1986**, *52*, 195–206. 10.1007/BF02889963.
- 52. Egozi, E.I.; Ferreira, A.M.; Burns, A.L.; Gamelli, R.L.; Dipietro, L.A. Mast cells modulate the inflammatory but not the proliferative response in healing wounds. *Wound Repair Regen*. 2003, *11*, 46–54. https://doi.org/10.1046/j.1524-475x.2003.11108.x.
- 53. Artuc, M.; Hermes, B.; Steckelings, U.M.; Grützkau, A.; Henz, B.M. Mast cells and their mediators in cutaneous wound healing--active participants or innocent bystanders? *Exp. Dermatol.* **1999**, *8*, 1–16.
- 54. Baum, C.L.; Arpey, C.J. Normal cutaneous wound healing: Clinical correlation with cellular and molecular events. *Dermatol. Surg.* **2005**, *31*, 674–686.
- Bonelli, A.; Bacci, S.; Norelli, G.A. Affinity cytochemistry analysis of mast cells in skin lesions: A possible tool to assess the timing of lesions after death. *Int. J. Leg. Med.* 2003,117, 331–334. https://doi.org/10.1007/s00414-003-0396-1.
- Trautmann, A.; Toksoy, A.; Engelhardt, E.; Bröcker, E.B.; Gillitzer, R. Mast cell involvement in normal human skin wound healing; expression of monocyte chemoattractant protein-1 is correlated with recruitment of mast cells which synthesize interleukin-4 in vivo. J. Pathol. 2000, 190, 100–106. https://doi.org/10.1002/(SICI)1096-9896(200001)190:1<100::AID-PATH496>3.0.CO;2-Q.
- Oehmichen, M.; Gronki, T.; Meissner, C.; Anlauf, M.; Schwark, T. Mast cell reactivity at the margin of human skin wounds: An early cell marker of wound survival? *Forensic Sci. Int.* 2009, 191, 1–5. https://doi.org/10.1016/j.forsciint.2009.05.020.
- Bacci, S.; Romagnoli, P.; Norelli, G.A.; Forestieri, A.L.; Bonelli, A. Early increase in TNF-alpha-containing mast cells in skin lesions. *Int. J. Leg. Med.* 2006, 120, 138–142. https://doi.org/10.1007/s00414-005-0030-5.
- Steinhoff, M.; Buddenkotte, J.; Shpacovitch, V.; Rattenholl, A.; Moormann, C.; Vergnolle, N.; Luger, T.A.; Hollenberg, M.D. Proteinase-activated receptors: Transducers of proteinase-mediated signaling in inflammation and immune response. *Endocr. Rev.* 2005, 26, 1–43. https://doi.org/10.1210/er.2003-0025.
- Terakawa, M.; Tomimori, Y.; Goto, M.; Fukuda, Y. Mast cell chymase induces expression of chemokines for neutrophils in eosinophilic EoL-1 cells and mouse peritonitis eosinophils. *Eur. J. Pharmacol.* 2006 538, 175–181. https://doi.org/10.1016/j.ejphar.2006.03.053.
- 61. Abdel-Majid, R.M.; Marshall, J.S. Prostaglandin E2 induces degranulation-independent production of vascular endothelial growth factor by human mast cells. *J. Immunol.* **2004**, *172*, 1227–1236. https://doi.org/10.4049/jimmunol.172.2.1227.
- Muramatsu, M.; Katada, J.; Hattori, M.; Hayashi, I.; Majima, M. Chymase mediates mast cell-induced angiogenesis in hamster sponge granulomas. *Eur. J. Pharmacol.* 2000, *18*, 181–191. https://doi.org/10.1016/s0014-2999(00)00350-2.
- Doggrell, S.A.; Wanstall, J.C. Vascular chymase: Pathophysiological role and therapeutic potential of inhibition. *Cardiovasc. Res.* 2004, 61, 653–662. https://doi.org/10.1016/j.cardiores.2003.11.029.

- 64. Somasundaram, P.; Ren, G.; Nagar, H.; Kraemer, D.; Mendoza, L.; Michael, L.H.; Caughey, G.H.; Entman, M.L.; Frangogiannis, N.G. Mast cell tryptase may modulate endothelial cell phenotype in healing myocardial infarcts. *J. Pathol.* 2005, 205, 102–111. https://doi.org/10.1002/path.1690.
- 65. Presta, M.; Leali, D.; Stabile, H.; Ronca, R.; Camozza, M.; Coco, L.; Moroni, E.; Liekens, S.; Rusnati, M. Heparin derivatives as angiogenesis inhibitors. *Curr. Pharm. Des.* 2003, *9*, 553–566. https://doi.org/10.2174/1381612033391379.
- 66. Abe, M; Kurosawa, M; Ishikawa, O; Miyachi, Y; Kido, H. Mast cell tryptase stimulates both human dermal fibroblast proliferation and type I collagen production. *Clin. Exp. Allergy* **1998**, *28*, 1509–1517. https://doi.org/10.1046/j.1365-2222.1998.00360.x.
- 67. Komi, D.; Khomtchouk, K.; Santa Maria, P.L. A review of the contribution of mast cells in wound healing: Involved molecular and cellular mechanisms. *Clin. Rev. Allergy Immunol.* 2020, *58*, 298–312. https://doi.org/10.1007/s12016-019-08729-w.
- van der Veer, W.M.; Bloemen, M.C.; Ulrich, M.M.; Molema, G.; van Zuijlen, P.P.; Middelkoop, E.; Niessen, F.B. Potential cellular and molecular causes of hypertrophic scar formation. *Burns* 2009, *35*, 15–29. https://doi.org/10.1016/j.burns.2008.06.020.
- 69. Wulff, B.C.; Parent, A.E.; Meleski, M.A.; DiPietro, L.A.; Schrementi, M.E.; Wilgus, T.A. Mast cells contribute to scar formation during fetal wound healing. *J. Investig. Dermatol.* **2012**, *132*, 458–465. https://doi.org/10.1038/jid.2011.324.
- 70. Ud-Din, S.; Wilgus, T.A.; Bayat, A. Mast cells in skin scarring: A review of animal and human research. *Front. Immunol.* **2020**, *11*, 552205. https://doi.org/10.3389/fimmu.2020.552205.
- 71. Wilgus, T.A.; Ud-Din, S.; Bayat, A. A Review of the Evidence for and against a role for mast cells in cutaneous scarring and fibrosis. *Int. J. Molec. Sci.* 2020, *21*, 9673. https://doi.org/10.3390/ijms21249673.
- 72. Chen, L.; Schrementi, M.E.; Ranzer, M.J.; Wilgus, T.A.; DiPietro, L.A. Blockade of mast cell activation reduces cutaneous scar formation. *PLoS ONE* **2014**, 22; e85226. https://doi.org/10.1371/journal.pone.0085226.
- 73. Wang, Z.C.; Zhao, W.Y.; Cao, Y.; Liu, Y.Q.; Sun, Q.; Shi, P.; Cai, J.Q.; Shen, X.Z.; Tan, W.Q. The Roles of Inflammation in keloid and hypertrophic scars. *Front. Immunol.* **2020**, *11*, 603187. https://doi.org/10.3389/fimmu.2020.
- Kyaw, B.M.; Järbrink, K.; Martinengo, L.; Car, J.; Harding, K.; Schmidtchen, A. Need for Improved Definition of "Chronic Wounds" in Clinical Studies. Acta Derm. Venereol. 2018, 98, 157–158. https://doi.org/10.2340/00015555-2786.
- 75. Gottrup, F. A specialized wound-healing center concept: Importance of a multidisciplinary department structure and surgical treatment facilities in the treatment of chronic wounds. *Am. J. Surg.* **2004**, 187, 8S–43S. https://doi.org/10.1016/S0002-9610(03)00303-9.
- Huttunen, M.; Aalto, M.L.; Harvima, R.J.; Horsmanheimo, M.; Harvima, I.T. Alterations in mast cells showing tryptase and chymase activity in epithelializating and chronic wounds. *Exp. Dermatol.* 2000, *9*, 258–265. https://doi.org/10.1034/j.1600-0625.2000.009004258.x.
- Huttunen, M.; Naukkarinen, A.; Horsmanheimo, M.; Harvima, I.T. Transient production of stem cell factor in dermal cells but increasing expression of Kit receptor in mast cells during normal wound healing. *Arch. Dermatol. Res.* 2002, 294, 324–330. https://doi.org/10.1007/s00403-002-0331-1.
- 78. Corsi, A.; Lecci, P.P.; Bacci, S.; Cappugi, P.; Pimpinelli, N. Early activation of fibroblasts during PDT treatment in leg ulcers. *G. Ital. Dermatol. Venereol.* **2016**, 151, 223–229.
- Grandi, V.; Bacci, S.; Corsi, A.; Sessa, M.; Puliti, E.; Murciano, N.; Scavone, F.; Cappugi, P.; Pimpinelli, N. ALA-PDT exerts beneficial effects on chronic venous ulcers by inducing changes in inflammatory microenvironment, especially through increased TGF-beta release: A pilot clinical and translational study. *Photodiagnosis Photodyn. Ther.* 2018, 21, 252–256. https://doi.org/10.1016/j.pdpdt.2017.12.012.
- Grandi, V.; Paroli, G.; Puliti, E.; Bacci, S.; Pimpinelli, N. Single ALA-PDT irradiation induces increase in mast cells degranulation and neuropeptide acute response in chronic venous ulcers: A pilot study. *Photodiagnosis Photodyn. Ther* 2021, 34, 102222. https://doi.org/10.1016/j.pdpdt.2021.102222.
- 81. Bacci, S. Cellular Mechanisms and Therapies in Wound Healing: Looking toward the Future. *Biomed.* **2021**, *9*, 1611, doi:10.3390/biomedicines9111611.