

Contents lists available at ScienceDirect

Science of the Total Environment

journal homepage: www.elsevier.com/locate/scitotenv



Ingestion of chitosan-starch blends: Effect on the survival of supralittoral amphipods

Alberto Ugolini^{a,*}, Alessandro Russo^a, Jessica Costa^b, Alessandra Cincinelli^c, Tania Martellini^c, Luca Conti^c, Duccio Cavalieri^a, Luca Mercatelli^d, Rebecca Pogni^b

^a Dipartimento di Biologia, Università di Firenze, Italy

^b Dipartimento di Biotecnologie, Chimica e Farmacia, Università di Siena, Italy

^c Dipartimento di Chimica "Ugo Schiff", Università di Firenze, Italy

^d Istituto Nazionale di Ottica - CNR Firenze, Italy

HIGHLIGHTS

G R A P H I C A L A B S T R A C T

- Biopolymers blends are likely to be released in the environment.
- Sandhoppers were fed with chitosanstarch mixtures at different percentages.
- Chitosan sheets were analysed before and after ingestion by ATR-FTIR.
- Sandhoppers fed with chitosan did not survive.



ARTICLE INFO

Editor: Damià Barceló

Keywords: Chitosan-starch blends NMR FTIR Sandhoppers Talitrus saltator

ABSTRACT

Sandy beach ecosystems are particularly affected by plastic pollution. Supralittoral amphipods are important components of the food web in sandy beaches and their ability to ingest microplastics and bioplastics has been assessed. Chitosan, a polysaccharide obtained by deacetylation of chitin, the second most abundant polymer in the world, represents an interesting component to produce novel bioplastics in combination with other biopolymers like starch. Here, the possibility of ingesting chitosan-starch blends and the possible effects on the amphipod *Talitrus saltator* were investigated. Groups of adult individuals were fed with sheets containing mixtures of chitosan and starch in different percentages for 7 and 14 days. The results showed that chitosan ingestion is dependent on the percentage of starch present in the mixture. Moreover, FTIR analyses of both sheets and faecal pellets after consumption show that chitosan is not digested. Furthermore, the survival rate of amphipods fed with a mixture of chitosan and starch decreases after one week compared to the control groups (100 % starch and paper), and drops drastically to 0 % after two weeks the experimental observations evidenced that chitosan is avoided as food resource and its consumption significantly affects the survival capacity of *T. saltator*. It is

* Corresponding author at: Dipartimento di Biologia, Via Romana 17, 50125 Firenze, Italy. *E-mail address: alberto.ugolini@unifi.it* (A. Ugolini).

https://doi.org/10.1016/j.scitotenv.2024.175302

Received 20 May 2024; Received in revised form 2 August 2024; Accepted 3 August 2024 Available online 5 August 2024 0048-9697/© 2024 Elsevier B.V. All rights are reserved, including those for text and data mining, AI training, and similar technologies.

1. Introduction

Plastic pollution poses a threat to ecosystems worldwide (*e.g.* see Derraik, 2002; Browne et al., 2010, 2011; Andrady, 2011; Galgani et al., 2019), particularly sandy beach ecosystems (Scopetani et al., 2018; Horn et al., 2019; Serra-Gonçalves et al., 2019; Xu et al., 2020; Abelouah et al., 2022; Rose et al., 2024). The sandy shores are subject to accumulation of plant and animal material of marine and terrestrial origin. In this type of ecosystem, the stranded material is used by detritivorous, scavengers, and herbivorous organisms and degraded by bacteria (Griffiths and Stenton-Dozey, 1981; Griffiths et al., 1983; Porri et al., 2011; Nakamura et al., 2022). Among the beached materials, plastic and microplastic are often present in considerable quantities (Thompson et al., 2004; Kaberi et al., 2013; Lots et al., 2017; Martellini et al., 2018) and are ingested by many invertebrates (*e.g.* see Xu et al., 2020; Costa et al., 2023).

The amount of plastic waste entering the ocean is still uncertain, but studies (*i.e.* Jambeck et al., 2015; Zhu, 2021) estimated it at 4–23 million tonnes per year and is predicted to increase in the next years. Moreover, Baghi et al. (2022) reported that plastic production has nearly doubled in the last decade and is foreseen to rise further reaching 33 billion tonnes by 2050. Due to an increased awareness of the environmental impacts from conventional plastics, bioplastics are increasingly being proposed as a good environmentally friendly alternative (Averous et al., 2001; Bastioli, 2001; Hottle et al., 2013; Arikan and Ozsoy, 2015; Atiwesh et al., 2021; Melchor-Martínez et al., 2022).

One of the EU directives requires all plastic packaging placed on the market should be reusable or economically recycled by 2030 (Calabro and Grosso, 2018; European Commission, 2018; European Bioplastic, 2020; Venâncio et al., 2022). As a result, many companies have produced and promoted "green" plastics, which have been positively received by consumers by associating bioplastic, biodegradable, compostable, or degradable with fast decomposition properties without environmental traces (Goel et al., 2021). The production of bioplastic has increased in recent years, reaching according to European Bioplastics, approximately 2.1 million tonnes in 2019, with a predicted growth to 2.4 million tonnes by 2024 (European Bioplastics, 2021; Baghi et al., 2022). Not all bioplastics are biodegradable (Albertsson et al., 2020; European Bioplastic, 2022) in open environment and, even if they are, the biodegradability standards are only valid under specific and controlled conditions of humidity, temperature, and microorganism concentration (e.g. Tong et al., 2022); thus, their improper disposal should not be underestimated. Therefore, the replacement of plastics by bioplastics may lead citizens to misbehave about waste disposal (Dilkes-Hoffman et al., 2019a, 2019b). Recently, the growing production of bioplastics and their abandonment in the marine and coastal environments has led to increased interest in the assessment of effects on aquatic organisms (see Teuten et al., 2009 for a review). Currently, chitosan might be a material of great interest for the bioplastic industry, being one of the most abundant linear polysaccharides on our planet. It is a partially N-deacetylated derivative of chitin, a biopolymer second only to cellulose in terms of abundance in nature. Chitosan is used for various industrial applications and as a biodegradable antimicrobial food packaging material competing with traditional non-biodegradable plastic-based materials. It is used in various sectors, such as biomedical, agriculture and water treatment. It is also used as a component in the cosmetic, textile, electronic, paper and food industries, and in some disposable products (for reviews see Yilmaz Atay, 2019; Zhang et al., 2021; Roman-Doval et al., 2023, Costa et al., 2024). Chitosan may have a lower environmental impact than traditional petroleum-based plastics as it is biodegradable and compostable (Leceta et al., 2013), however,

even in this case, plasticizers and cross-linkers are used in some chitosan-containing products to improve their properties (Priyadarshi and Rhim, 2020). Furthermore, chitosan and its derivatives have attracted particular attention regarding their antimicrobial and antifungal activity (Kendra and Hadwiger, 1984; Muzzarelli et al., 1990; Tsai and Su, 1999; Li et al., 2002; Qi et al., 2004; Kong et al., 2010; Yilmaz Atay and Çelik, 2017; Aktuganov et al., 2018; Kidibule et al., 2021; Guarnieri et al., 2022).

Thus, chitosan appears to be a potential future replacement for many of the bioplastics currently in use. However, the negative effects of ingesting macro and microplastics on many marine organisms are well documented by an extensive literature on this topic (e.g. see Cole et al., 2011 and Guzzetti et al., 2018 for reviews) but despite the fact that marine sandy shores are among the main places of deposition of various types of plastics, only a few studies have investigated the effects of bioplastic ingestion by supralittoral arthropods (Tosetto et al., 2016; Hodgson et al., 2018; Carrasco et al., 2019; Shruti and Kutralam-Muniasamy, 2019; Martellini et al., 2023). Among these, it is known that many amphipod species are of considerable importance in the degradation of beached material and constitute a food resource for many invertebrates and birds, including migratory species. Furthermore, talitrid amphipods being herbivores, scavengers and grazers (Palluault, 1954; Lopez et al., 1977; Adin and Riera, 2003; Colombini et al., 2003, 2015; Olabarria et al., 2009; Porri et al., 2011; Abdelrhman et al., 2017) can absorb various types of pollutants; in fact, supralittoral amphipods, like Talitrus. saltator (Montagu) are good indicators of trace metals (Rainbow et al., 1999; Rainbow, 2002; Marsden and Rainbow, 2004; Ungherese and Ugolini, 2009; Ungherese et al., 2010a,b; Ugolini et al., 2004, 2005; Ugolini and Ungherese, 2012; Rochman et al., 2013; Jelassi et al., 2021), polybrominated diphenyl ethers (PBDEs) and polycyclic aromatic hydrocarbons (PAHs) (Ugolini and Ungherese, 2012; Ugolini et al., 2012; Ungherese et al., 2016) and it is demonstrated that they can ingest microplastics mixed with food (Ugolini et al., 2013; Hodgson et al., 2018; Scopetani et al., 2018; Iannilli et al., 2018; Carrasco et al., 2019).

The important role that amphipods play in the supralittoral zone of sandy coasts in the trophic chain of the detritus, consisting of the fragmentation and modification of stranded organic material, is well known (Palluault, 1954; Griffiths et al., 1983). Furthermore, amphipods constitute an important source of nutrition for various species of riparian birds, including migratory ones (Dauvin, 2024).

It seemed useful to consider the potential use of chitosan-starch blends, as components to produce new biocomposites, as a food source and the possible effects of their ingestion on the survival rate of supralittoral amphipods. The role of this species in marine coastal ecology as a bio-indicator for various organic and inorganic contaminants has been widely described, while, to the best of our knowledge, the effects that the intake of bioplastics of different compositions may have on it remain rather unexplored. With the aim of filling this gap, this study explores the effect, under controlled conditions (temperature, photoperiod), of chitosan/starch blends in different percentages and in the absence of a bacterial film on the survival of *T. saltator*.

2. Materials and methods

Adult specimens of *T. saltator*, as it is one of the most common species of the supralittoral of sandy coasts present from the Mediterranean to the European coasts of the Atlantic Ocean, North Sea and Baltic Sea, were used for the experiments. A work-flow of the experimental procedure is reported in Supplementary Information (Fig. S1). The animals (about 200 individuals) were collected one by one using entomological aspirators in the spring-summer of 2022 and 2023 in the Migliarino San Rossore Massaciuccoli Regional Natural Park (Pisa, Fiume Morto Vecchio locality, 43°44′55" N; 10°16′31″ E). The amphipods were transported in boxes with wet sand to the laboratory in Florence within 3 h from the collection and placed in plastic tanks (17 × 11 × 13 cm). Each tank was divided in half by a 3 cm high plastic partition glued to the bottom so that the artificial sand (Amtra deko) in one half could be moistened with seawater, while the other remained dry. The temperature was regulated at 25 ± 2 °C, the artificial lighting cycle (L:D = 12:12) was in phase with the natural one.

In each tank, 10 individuals fasted for 2 days were placed before the start of the experiment. A total of 30 or 40 individuals were tested for each experimental treatment. All the food sources used were made up of 5×5 cm sheets of 1) absorbent paper and dry food for aquarium fish (Sera Vipan, Germany), standard food normally used to keep sandhoppers in captivity even for a long time (controls); 2) 100 % starch; 3) 100 %-chitosan; 4) 75 %-chitosan mixed with 25 %-starch; 5) 50 %-chitosan mixed with 50 %-starch; 6) 25 %-chitosan mixed with 75 %-starch. No additives, like plasticizers, were added to the samples.

Chitosan mixed with starch sheets were developed using the solvent casting method (Gupta et al., 2022). 100 %-Starch sheets were prepared dissolving 400 mg of corn starch (Sigma Aldrich) in 50 ml of hot distillated water at 80 °C, with continuous stirring for 15 min. The resulting solution was then poured into a Petri dish (10 \times 10 cm) and dried. Once the solvent evaporated, a 100 % starch sheet was formed. For 100 %-chitosan sheets, 400 mg of chitosan (Sigma Aldrich - low molecular weight) was solubilized in 50 ml of 2 % acetic acid at 60 °C, stirred magnetically for 10 min, and then poured into a Petri dish to dry. To prepare the 75 %-chitosan - 25 %-starch sheets, the polymers were solubilized separately. Firstly, 100 mg of starch powder was dissolved in 15 ml of hot distillated water at 80 °C under magnetic stirrer for 15 min. At the same time, 300 mg of chitosan was solubilized in 35 ml of 2 %acetic acid at 60 °C with stirring for 10 min. Subsequently, the starch solution was added to chitosan solution, thoroughly mixed to obtain a homogeneous solution, and poured into a Petri dish until it is dried. Similar procedures were followed for 50 %-chitosan - 50 %-starch sheet and 25 %-chitosan - 75 %-starch sheet, with changes in amount of powder and solvent. For the preparation of the first sheet, two solutions were prepared with 200 mg of powder dissolved in 25 ml of solvent, while for the second one, 100 mg of chitosan in 15 ml of acetic acid and 300 mg of starch in 35 ml of water were used (Li et al., 2013). The resulting sheets had a thickness of 0.02 mm.

For each treatment 7 days after the introduction of the sheets into the tanks (one sheet of one type per tank placed on the dry sand), the surface not eaten by the amphipods was measured in relation to the total sheet surface. This was possible thanks to a specially written software in the MatLab environment (The MathWork, Inc.) capable of detecting the white (uneaten surface) and black (eaten surface) pixels that compose the photographic image in the absence of halftones, of each sheet at the end of the experiment (Fig. 1, see also Martellini et al., 2023). Furthermore, for each treatment, the number of individuals still alive 7 and 14 days after the start of the experiment was considered.

2.1. Chemical analyses

To investigate the possible degradation of chitosan following intestinal passage, feces from sandhoppers exposed to 50 %-chitosan-50 %-starch and 100 %-starch were collected within two days of deposition. As a control, analyses were also conducted on 50 %-chitosan – 50 %-starch, 100 %-starch, and 100 %-chitosan samples taken from unused sheets. These analyses were carried-out by an IRAffinity-1S by SHI-MADZU equipped with the ATR sampling accessory (MIRacleTM PIKE Technologies) and GC–MS analysis (GC 7890 A and MS 5975C Agilent Technologies), in the 4000–500 cm⁻¹ range. Additionally, the faecal pellets obtained from the amphipods were subjected to FTIR analysis.



Fig. 1. Photographs of the consumption of polymers by *T. saltator* after 7 days. Only the most representative images, chosen *ad hoc*, of all the photos of the sheets used are shown. The uneaten surface is the white one. 100 %-starch (St100), paper, 25 %-chitosan mixed with 75 %-starch (C25), 50 %-chitosan mixed with 50 %-starc- (C50), 75 %-chitosan mixed with 25 %-starch (C75), and 100 %-chitosan (C100).

2.2. Statistical analysis

Statistical analyses were performed using SPSS (IBM). Comparisons between the different treatments relating to consumption were carried out using the Mann-Whitney *U* test while for comparisons between treatments relating to survival the G test was used (Siegel and Castellan Jr., 1988; Zar, 1999). The limit of statistical significance was set at $P \leq 0.05$.

3. Results

3.1. Consumption

After 7 days in the tanks (Fig. 2) it was evident that the 100 %-chitosan sheet was not eaten by the sandhoppers as the sheet appears practically intact. The non-consumed chitosan area was significantly larger than that found for all other treatments (P = 0.032, Mann Whitney U test, it was the worst value obtained among all comparisons). Among the comparisons between the uneaten parts relating to sheets containing chitosan and starch in different percentages, the only statistically significant one was between 75 %-chitosan and 25 %-chitosan, the latter being preferred by sandhoppers (P = 0.021, Mann Whitney U test). Therefore, 25 %, 50 %, and 75 %-chitosan seemed more palatable than the polymer with the highest percentage of chitosan. The comparison between paper and 100 %-starch does not reach statistical significance (U = 18, P = 0.365) whereas all comparisons between sheets containing chitosan in various percentages vs 100 %-starch and vs paper are significant (U = 7, P = 0.05 in the worst case: 50 %-chitosan vs paper).

3.2. Survival rate

A high survival rate is observable after 7 days (Fig. 3A) both in animals fed with paper (26/30, 90 %) and in those fed with 100 %-starch (27/30, 87 %) (Fig. 3, G = 0.151, df = 1, P—NS, G test). A high survival rate (21/30, 70 %) was also recorded in the presence of 100 %-chitosan, in fact the comparisons between survival with 100 %-chitosan *vs* 100 %-starch and *vs* paper were not fully statistically significant, G = 3.728, df = 1, 0.05 < P < 0.1, G test, in the best case) while the comparisons between 100 %-chitosan *vs* 75 %-chitosan (G = 5.210, df = 1, P < 0.05, G test) and reached significance chitosan –100 % *vs* 50 %-chitosan (G =



Fig. 2. Box-plot diagrams of the uneaten area after 7 days of the sheets of paper, 100 %-starch (St100), paper, 25 %-chitosan mixed with 75 %-starch (C25), 50 %-chitosan mixed with 50 %-starch (C50), 75 %-chitosan mixed with 25 %-starch (C75), and 100 %-chitosan (C100). The asteriscs indicate the probability level (P < 0.05) of the comparisons among groups (Mann Whitney U test). See text for further information and more details about statistical analysis (Section 3.1)



Fig. 3. Survival rate of *T. saltator* after 7 and 14 days (A and B, respectively) in the presence of paper sheets, 100 %-starch (St100), paper, 25 %-chitosan mixed with 75 %-starch (C25), 50 %-chitosan mixed with 50 %-starch (C50), 75 %-chitosan mixed with 25 %-starch (C75), and 100 %-chitosan (C100). The numbers above the bars indicate the number of tested individuals. The asteriscs indicate the probability level of the comparisons among groups (G test) (* = P < 0.05, ** = P < 0.02, *** = P < 0.01, ~ means 0.05 < P < 0.1). See text for further information and more details about statistical analysis (see Section 3.2).

7.247, df = 1, P < 0.01, G test). The comparison between 100 %-chitosan vs 25 %-chitosan was not statistically significant (G = 0.643, df = 1, P=NS, G test) The comparisons between paper or 100 %-starch and the number of survivors exposed to sheets containing different percentages of chitosan and starch were all statistically significant (the worst level was related to the comparison between 100 %-starch vs 25 %-chitosan, G = 5.461, df = 1, 0.01 < P < 0.02).

After 14 days (Fig. 3B), it could be noted that the survival in the presence of 100 %-chitosan was 0 %. The same was observed for 75 %-chitosan and 50 %-chitosan. Even for 25 %-chitosan the survival was low (3 %). In the comparison among the survivals recorded in the presence of the sheets with the different percentages of chitosan, full statistical significance was never reached; the best level was achieved in the comparison between 100 %-chitosan *vs* 25 %-chitosan (G = 3.699, df = 1, 0.05 < P < 0.1, G test). Even the comparison between paper and 100 %-starch did not reach statistical significance (G = 0.314, df = 1, 0.05 < P < 0.1, G test).

P=NS, G test) and a high survival of *T. saltator* was evident (paper = 90 %, starch = 95 %). Comparisons between 7 and 14 days showed no significant differences in the number of sandhoppers surviving in the presence of paper (G = 0.231, df = 1, P=NS, G test) or 100 %-starch (G = 0.910, df = 1, P=NS, G test). However, comparisons made between the number of survivors after 7 and 14 days within each treatment with sheets containing different percentages of chitosan were all significant: the 7 vs 14 days comparison with 25 %-chitosan was the worst result (G = 17.318, df = 1, P < 0.001, G test).

3.3. Chemical analyses

In Fig. 4A are reported the ATR-FTIR spectra for 30 %-chitosan compared to 100 %-chitosan and 100 %-starch, which show the typical absorption bands of these two polysaccarides (Hebeish et al., 2009; Drabczyk et al., 2020). Although, the spectra of the two polysaccharides



Fig. 4. A, ATR-FTIR spectra of 100 %-chitosan, 100 %-starch, and 50 %-chitosan-with the main absorption bands; B, comparison between spectra obtained for chitosan-50 % and its corresponding faecal pellets. The stars indicate the diagnostic absorptions of starch while in the insets is shown a focus on the 1450–990 cm⁻¹ region highlighting the main differences between the spectra of chitosan -50 % (feeding sheet) and faecal pellets.

100 % could seem very similar, significant differences can be evidenced in their FTIR profiles. Observing the adsorption region between 1151 and 999 cm⁻¹, associated to the asymmetric stretching of the C-O-C bridge and the C—O stretching (Wang and Xie, 2010), can be evidenced that the C—O stretching of starch-100 % is significantly shifted towards lower frequencies, of *c.ca* 29 cm⁻¹, if compared to that of 100 %-chitosan. This is of particular interest in the comparative analysis between the FTIR spectra of 50 %-chitosan and faecal pellets (see Fig. 4B, inset).

As expected, the FTIR spectrum of 50 %-chitosan (light grey, Fig. 4A) contains all the signals separately displayed by its components, thus confirming the presence of both polysaccharides in the mixed sample.

The comparison between the FTIR spectra of 50 %-chitosan and of its corresponding faecal pellets is reported in Fig. 4B. As shown, in the profile of faecal pellets can be still appreciated the absorption bands of both chitosan and starch, thus hinting at their simultaneous presence in the sample. Interesting information is then achieved by analysis of the representative fingerprint region of spectra. More, in this region the spectrum of faecal pellets exhibits the typical triplet of absorptions of

chitosan, as it can be easily appreciated from the inset of Fig. 4B, where it is visible the last C—O stretching resonating at 1028 cm⁻¹, instead of at 999 cm⁻¹ as in 50 %-chitosan. This can be assumed as diagnostic of a predominant contribution of chitosan in the infrared spectrum of faecal pellets indicating a probable digestion of the starch component of the polymeric matrix. Outside this region, the differences found at 1648 and 1558 cm⁻¹ among the two spectra are less reliable to gain information regarding the composition of faecal pellets as, on one side, vibrations of chitosan in this region have been generally reported to significantly vary depending on the particular chitosan fabrication (Kwon and Jeong, 2020), and, importantly, the absorption of starch at ~1642 cm⁻¹ is strongly affected by the highly variable crystallinity of commonly commercialized starches (Kizil et al., 2002).

Therefore, despite a quantitative estimation cannot be made based on the sole IR data, taken together these data suggest a preferential digestion of the starch-component of the polymer, in line with the predominant presence of the chitosan component in the faecal pellets witnessed by infrared analysis.

4. Discussion

The main achievement of our research is that the ingestion of chitosan significantly affects the survival capacity of *T. saltator* and seems that chitosan is avoided as a food resource.

The high survival rates after both 7 and 14 days in the starch-100 %and paper (i.e. cellulose) treatments show that these elements may constitute an important trophic resource for supralittoral amphipods. These findings align with current knowledge on the feeding habits of supralittoral talitrids, although some species-specific differences are known (Mengoni et al., 2013; Abdelrhman et al., 2017). In fact, T. saltator, grazer and scavenger feeds on stranded detritus of both animal and vegetable origin. In the categories with different percentages of chitosan the survival rates to the seventh day vary greatly. With chitosan-25 % there is a survival rate of 60 %, with chitosan-50 % and 75 % the survival drops to 37 % and 42 %, respectively but, surprisingly, in the presence of chitosan-100 % survival increases (70 %). However, after 14 days, while with paper or 100 % starch survival remains practically unchanged compared to 7 days, survival in the presence of chitosan drops to 0 % regardless of the percentage contained in the food sheet (Fig. 3).

Despite the reduced surface area consumed (Fig. 2), the survival rate after 7 days in 100 %-chitosan appears so high could be due to the use of temporary food sources, such as feces or dead individuals. However, these very limited food sources do not seem sufficient to cover nutritional needs for 14 days.

In accordance with the above, the most consumed sheets were paper and 100 %-starch. Among the sheets containing chitosan the highest consumption was observed for 25 %-chitosan; this is probably due to the higher percentage of starch making it more palatable, while for 100 %-chitosan the consumption was almost absent. It should be also noted that sandhoppers fed with 100 %-starch and paper survive in greater numbers than individuals fed with sheets with various percentages of chitosan for which higher mortality rates are found. This was further evidenced by the FTIR analysis of faecal pellets from individuals fed with different feeding sheets (Fig. 4). Furthermore, the difference in survival could also be attributed to alterations in the digestive system due to chitosan ingestion. In fact, following ingestion, chitosan tends to swell, creating a physical bulk that can prevent the correct processing of food within the digestive system and inducing a false perception of satiety; moreover, chitosan is also known for reducing dietary fat absorption in the intestine (Cheung et al., 2015). It has also been demonstrated that in Tubifex tubifex worms chitosan induced oxidative stress (Mosleh et al., 2007). Furthermore, chitosan possesses antimicrobial properties, being effective against the growth of some bacteria and fungi (Kendra and Hadwiger, 1984; Muzzarelli et al., 1990; Roller and Covill, 1999; Tsai and Su, 1999; Li et al., 2002; Qi et al., 2004; Carlson et al.,

2008a, 2008b; Xing et al., 2015; Kidibule et al., 2021; Dou et al., 2024) and once ingested by *T. saltator* chitosan could likely influence its digestive capabilities by altering or making inefficient the intestinal microbiome (Mengoni et al., 2013; Abdelrhman et al., 2017).

5. Conclusions

The ingestion of chitosan mixed with starch significantly affects the survival capacity of *T. saltator* within 14 days from the ingestion. Therefore, these results, suggest that the use of high percentage of chitosan to produce novel biocomposites, can be more dangerous for littoral fauna than other single components to produce starch-based bioplastics, as recently demonstrated on the same species (see Scopetani et al., 2018; Martellini et al., 2023). Beyond the evidence that chitosan is not a nutrional source for *T. saltator* a possible detrimental effect, associated to the antibacterial properties of this polymeric component, can be also envisaged.

Supplementary data to this article can be found online at https://doi.org/10.1016/j.scitotenv.2024.175302.

Funding

The research was funded by the University of Florence (local funds RICATEN) assigned to A.U, DC, TM, LC, AC.

Declaration of generative AI in scientific writing

No AI or AI-assisted tools were used during the preparation of our manuscript.

CRediT authorship contribution statement

Alberto Ugolini: Writing – review & editing, Writing – original draft, Validation, Supervision, Methodology, Investigation, Funding acquisition, Formal analysis, Data curation, Conceptualization. Alessandro Russo: Writing – review & editing, Investigation, Data curation. Jessica Costa: Methodology, Investigation. Alessandra Cincinelli: Writing – review & editing, Validation, Methodology, Investigation, Funding acquisition. Tania Martellini: Writing – review & editing, Methodology, Investigation, Funding acquisition. Luca Conti: Writing – original draft, Investigation, Funding acquisition. Duccio Cavalieri: Writing – review & editing, Visualization, Funding acquisition. Luca Mercatelli: Writing – original draft, Software, Methodology, Investigation. Rebecca Pogni: Writing – review & editing, Visualization, Validation, Methodology, Conceptualization.

Declaration of competing interest

The authors declare that they have no conflicting interests that influenced this research.

Data availability

Data will be made available on request.

Acknowledgements

Thanks are due to the Ente Parco Regionale di Migliarino, San Rossore, Massaciuccoli (Pisa, Italy) for authorization to amphipods collection.

References

- Abelouah, M.R., Ben-Haddad, M., Rangel-Buitrago, N., Hajji, S., El Alem, N., Ait Alla, A., 2022. Microplastics pollution along the central Atlantic coastline of Morocco. Mar. Pollut. Bull. 174, 113190.
- Adin, R., Riera, P., 2003. Preferential food source utilization among stranded macroalgae by *Talitrus saltator* (amphipod, Talitridae): a stable isotopes study in the northern coast of Brittany (France). Estuar. Coast. Shelf Sci. 56, 91–98.
- Aktuganov, G.E., Safina, V.R., Galimzianova, N.F., Kuz'mina, L. Yu, Gilvanova, E.A., Boyko, T.F., Melent'ev, A.I., 2018. Chitosan resistance of bacteria and micromycetes differing in ability to produce extracellular chitinases and chitosanases. Microbiology 87, 716–724.
- Albertsson, A.C., Bødtker, G., Boldizar, A., Filatova, T., Prieto Jimenez, M.A., Loos, K., 2020. Biodegradability of plastics in the open environment. Science Advice for Policy by European Academies. Evidence Review Report 8. https://doi.org/ 10.26356/biodegradabilityplastics.

Andrady, A.L., 2011. Microplastics in the marine environment. Mar. Pollut. Bull. 62, 1596–1605.

- Arikan, E.B., Ozsoy, H.D., 2015. A review: investigation of bioplastics. J. Civil Eng. Architect. 9, 188–192. https://doi.org/10.17265/1934-7359/2015.02.007.
- Atiwesh, G., Mikhael, A., Parrish, C.C., Banoub, J., Le, T.A.T., 2021. Environmental impact of bioplastic use: a review. Heliyon 7, e07918. https://doi.org/10.1016/j. heliyon.2021.e07918.
- Averous, L., Fringant, C., Moro, L., 2001. Starch-based biodegradable materials suitable for thermoforming packaging. Starch - Stärke 53, 368–371. https://doi.org/ 10.1002/1521-379X (200108) 53:83.0.CO;2-W.
- Baghi, F., Gharsallaoui, A., Dumas, E., Ghnimi, S., 2022. Advancements in biodegradable active films for food packaging: effects of nano/microcapsule incorporation. Foods 11, 760. https://doi.org/10.3390/foods11050760.
- Bastioli, C., 2001. Global status of the production of biobased packaging materials. Starch - Sträke 53, 351–355. https://doi.org/10.1002/1521-379X(200108)53:83.0. CO;2-R.
- Browne, M.A., Galloway, T.S., Thompson, R.C., 2010. Spatial patterns of plastic debris along estuarine shorelines. Environ. Sci. Technol. 49, 3404–3409.
- Browne, M.A., Crump, P., Niven, S.J., Teuten, E., Tonkin, A., Galloway, T.S., Thompson, R.C., 2011. Accumulation of microplastic on shorelines woldwide: sources and sinks. Environ. Sci. Technol. 45, 9175–9179.
- Calabrò, P.S., Grosso, M., 2018. Bioplastics and waste management. Waste Manag. 78, 800–801. https://doi.org/10.1016/j.wasman.2018.06.054.
- Carlson, R., Taffs, R., Davison, W., Stewart, P., 2008a. Anti-biofilm properties of chitosan-coated surfaces. J. Biomat. Sci. Polymer Edition 19, 1035–1046. https:// doi.org/10.1163/156856208784909372.
- Carlson, R.P., Taffs, R., Davison, W.M., Stewart, P.S., 2008b. Anti-biofilm properties of chitosan-coated surfaces. J. Biomat. Sci. Polymer Edition 19, 1035–1046. https:// doi.org/10.1163/156856208784909372.
- Carrasco, A., Pulgar, J., Quintanilla-Ahumada, D., Perez-Venegas, D., Quijón, P.A., Duarte, C., 2019. The influence of microplastics pollution on the feeding behavior of a prominent sandy beach amphipod, *Orchestoidea tuberculata* (Nicolet, 1849). Mar. Pollut. Bull. 145, 23–27. https://doi.org/10.1016/j.marpolbul.2019.05.018.
- Cheung, R.C.F., Ng, T.B., Wong, J.H., Chan, W.Y., 2015. Chitosan: an update on potential biomedical and pharmaceutical applications. Mar. Drugs 13 (8), 5156–5186.
- Cole, M., Lindaque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants in the marine environment: a review. Mar. Pollut. Bull. 62, 2588–2597.
- Colombini, I., Chelazzi, L., Gibson, R., Atkinson, R., 2003. Influence of marine allochthonous input on sandy beach communities. Oceanogr. Marine Biol. Annu. Rev. 41, 115–159.

Colombini, I., Fallaci, M., Chelazzi, L., 2015. Ecological strategies of Macarorchestia remyi compared to two sympatric talitrids of a Tyrrhenian beach. Acta Oecol. 67, 49–58.

- Costa, L.L., da Costa, I.D., da Silva Oliveira, A., Zalmon, I.R., 2023. Microplastic ecology: testing the influence of ecological traits and urbanization in microplastic ingestion by sandy beach fauna. Est. Coast. Shelf Sci. 290, 108406 https://doi.org/10.1016/j. ecss.2023.108406.
- Costa, J., Baratto, M.C., Spinelli, D., Leone, G., Magnani, A., Pogni, R., 2024. A novel bioadhesive based on chitosan-polydopamine-xanthan gum for glass, cardboard and textile commodities. Polymers 16, 1806. https://doi.org/10.3390/polym16131806.
- Dauvin, J.-C., 2024. Overview of predation by birds, cephalopods, fish and marine mammals on marine benthic amphipods. J. Mar. Sci. Eng. 12, 403. https://doi.org/ 10.3390/jmse12030403.
- Derraik, J.G.B., 2002. The pollution of the marine environment by plastic debris: a review. Mar. Pollut. Bull. 44, 842–852.
- Dilkes-Hoffman, L., Ashworth, P., Laycock, B., Pratt, S., Lant, P., 2019a. Public attitudes towards bioplastics – knowledge, perception and end-of-life management. Resources Conserv. Recycl. 151, 104479 https://doi.org/10.1016/j.resconrec.2019.104479.
- Dilkes-Hoffman, L., Lant, P.A., Laycock, B., Pratt, S., 2019b. The rate of biodegradation of PHA bioplastics in the marine environment: a meta-study. Mar. Pollut. Bull. 142, 15–24. https://doi.org/10.1016/j.marpolbul.2019.03.020.
- Dou, X., Fan, N., Yang, J., Zhang, Z., Wu, B., Wei, X., Shi, S., Zhang, W., Feng, Y., 2024. Research progress on chitosan and its derivatives in the fields of corrosion inhibition and antimicrobial activity. Environ. Sci. Pollut. Res. 31, 30353–30369. https://doi. org/10.1007/s11356-024-33351-5.
- Drabczyk, A., Kudłacik-Kramarczyk, S., Głab, M., Kedzierska, M., Jaromin, A., Mierzwiński, D., Tyliszczak, B., 2020. Physicochemical investigations of chitosanbased hydrogels containing *Aloe vera* designed for biomedical use. Materials 13, 3073. https://doi.org/10.3390/ma13143073.
- European Bioplastic, 2020. New Market data 2019: bioplastics industry shows dynamic growth. https://www.european-bioplastics.org/.
- European Bioplastic, 2021. EUBIO_Admin. Market. Eur. Bioplastics E.V., available online. https://www.european-bioplastics.org/market/.

Abdelrhman, K.F., Bacci, G., Marras, B., Nistri, A., Schintu, M., Ugolini, A., Mengoni, A., 2017. Exploring the bacterial gut microbiota of supralittoral talitrid amphipods. Res. Microbiol. 168, 74–84.

A. Ugolini et al.

European Bioplastic, 2022. What are bioplastics? https://docs.european-bioplastics.org/ publications/fs/EuBP_FS_What_are_bioplastics.pdf. https://www.european-bioplas tics.org/.

European Commission, 2018. Packaging Waste Directive (EU) 2018/852. European Commission Brussels.

- Galgani, L., Beiras, R., Galgani, F., Pani, C., Borja, A., 2019. Editorial: impacts of marine litter. Front. Mar. Sci. 6, 208.
- Goel, V., Luthra, P., Kapur, G.S., Ramakumar, S.S.V., 2021. Biodegradable/bio-plastics: myths and realities. J. Polymers Environ. 29, 3079–3104. https://doi.org/10.1007/ s10924-021-02099-1.
- Griffiths, C.L., Stenton-Dozey, J.M.E., 1981. The fauna and rate of degradation of stranded kelp. Estuar. Coast. Shelf Sci. 12, 645–653. https://doi.org/10.1016/ S0302-3524(81)80062-X.
- Griffiths, C.L., Stenton-Dozey, J.M.E., Koop, K., 1983. Kelp wrack and the flow of energy through a sandy beach ecosystem. In: Sandy beaches as ecosystems: based on the proceedings of the first international symposium on sandy beaches, held in Port Elizabeth, South Africa. Springer Netherlands, pp. 547–556.
- Guarnieri, A., Triunfo, M., Scieuzo, C., Ianniciello, D., Tafi, E., Hahn, T., Zibek, S., Salvia, R., De Bonis, A., Falabella, P., 2022. Antimicrobial properties of chitosan from different developmental stages of the bioconverter insect *Hermetia illucens*. Sci. Reports 12, 8084.
- Gupta, V., Biswas, D., Roy, S.A., 2022. Comprehensive review of biodegradable polymerbased films and coatings and their food packaging applications. Materials 15, 5899. https://doi.org/10.3390/ma15175899.
- Guzzetti, E., Sureda, A., Tejada, S., Faggio, C., 2018. Microplastic in marine organism: environmental and toxicological effects. Environ. Toxicol. Pharmacol. 64, 164–171. https://doi.org/10.1016/j.etap.2018.10.009.
- Hebeish, A., Aly, A.A., El-Shafei, A., Zaghloul, S., 2009. Synthesis and characterization of cationized starches for application in flocculation, finishing and sizing. Egypt. J. Chem. 60, 97–109. https://doi.org/10.1002/star.200700627.
- Hodgson, D.J., Bréchon, A.L., Thompson, R.C., 2018. Ingestion and fragmentation of plastic carrier bags by the amphipod *Orchestia gammarellus*: effects of plastic type and fouling load. Mar. Pollut. Bull. 127, 154–159. https://doi.org/10.1016/j. marpolbul.2017.11.057.
- Horn, D., Miller, M., Andrson, S., Steele, C., 2019. Microplastics are ubiquitous on California beaches and enter the coastal food web through consumption by Pacific mole crabs. Mar. Pollut. Bull. 139, 231–237.
- Hottle, T.A., Bilec, M.M., Landis, A.E., 2013. Sustainability assessments of bio-based polymers. Polymer Degrad. Stabil. 98, 1898–1907. https://doi.org/10.1016/j. polymdegradstab. 2013.06.016.
- Iannilli, V., di Gennaro, A., Lecce, F., Sighicelli, M., Falconieri, M., Pietrelli, L., Poeta, G., Battisti, C., 2018. Microplastics in *Talitrus saltator* (Crustacea, Amphipoda): new evidence of ingestion from natural contexts. Environ. Sci. Pollut. Res. 25, 28725–28729. https://doi.org/10.1007/s11356-018-2932-z.
- Jambeck, J.R., Geyer, R., Wilcox, C., Siegler, T.R., Perryman, M., Andrady, A., Narayan, R., Law, K.L., 2015. Plastic waste inputs from land into the ocean. Science 13 (347), 768–771. https://doi.org/10.1126/science.1260352.
- Jelassi, R., Khemaissia, H., Ghemari, C., Raimond, M., Souty-Grosset, C., Nasri-Ammar, K., 2021. Responses of Orchestia montagui (Amphipoda, Talitridae) to copper and zinc mixture. In: Kisbi, M., et al. (Eds.), Recent advances in environmental science from euro-mediterranean and surrounding regions, 2nd ed., Environ. Sci. Eng.
- Kaberi, H., Tsangaris, C., Zeri, C., Mousdisd, G., Papadopoulos, A., Streftaris, N., 2013. Microplastics along the shoreline of a Greek island (kea isl., Aegean Sea): Types and densities in relation to beach orientation, characteristics and proximity to sources. In: In: 4th International Conference on Environmental Management, Engineering, Planning and Economics (CEMEPE) and SECOTOX Conference, Mykonos Island, Greece, pp. 197–202.
- Kendra, D.F., Hadwiger, L.A., 1984. Characterization of the smallest chitosan oligomer that is maximally antifungal to *fusarium solani* and elicits pisatin formation in *Pisum sativum*. Exp. Mycol. 8, 276–281. https://doi.org/10.1016/0147-5975(84)90013-6.
- Kidibule, P.E., Costa, J., Atrei, A., Plou, F.J., Fernandez-Lobato, M., Pogni, R., 2021. Production and characterization of chitooligosaccharides by the fungal chitinase Chit42 immobilized on magnetic nanoparticles and chitosan beads: selectivity, specificity and improved operational utility. RSC Adv. 11, 5529. https://doi.org/ 10.1039/d0ra10409d.
- Kizil, R., Irudayaraj, J., Seetharaman, K., 2002. Characterization of irradiated starches by using FT-Raman and FTIR spectroscopy. J. Agric. Food Chem. 50, 3912–3918. https://doi.org/10.1021/jf011652p.
- Kong, M., Chen, X.G., Xing, K., Park, H.J., 2010. Antimicrobial properties of chitosan and mode of action: a state-of-the-art review. Int. J. Food Microbiol. 144, 51–63.
- Kwon, W., Jeong, E., 2020. Detoxification properties of guanidinylated chitosan against chemical warfare agents and its application to military protective clothing. Polymers 12, 1461. https://doi.org/10.3390/polym12071461.
- Leceta, I., Guerrero, P., Cabezudo, S., Caba, K.D.L., 2013. Environmental assessment of chitosan-based films. J. Cleaner Product. 41, 312–318. https://doi.org/10.1016/j. jclepro.2012.09.049.
- Li, Z., Zhuang, X., Liu, X., Guan, Y., Yao, K., 2002. Study on antibacterial Ocarboxymethylated chitosan/cellulose blend film from LiCl/N, N-dimethylacetamide solution. Polymer 43, 1541–1547. https://doi.org/10.1016/S0032-3861(01)00699-1
- Li, H., Gao, X., Wang, Y., Zhang, X., Tong, Z., 2013. Comparison of chitosan/starch composite film properties before and after cross-linking. Int. J. Biol. Macromol. 52, 275–279. https://doi.org/10.1016/j.ijbiomac.2012.10.016.

- Lopez, G., Levinton, J., Slobodkin, L., 1977. The effect of grazing by the detritivore Orchestia grillus on Spartina litter and its associated microbial community. Oecologia 30, 111–127.
- Lots, F.A.E., Behrens, P., Vijver, M.G., Horton, A.A., Bosker, T., 2017. A large-scale investigation of microplastic contamination: abundance and characteristics of microplastics in European beach sediment. Mar. Pollut. Bull. 123, 219–226.
- Marsden, I.D., Rainbow, P.S., 2004. Does the accumulation of trace metals in crustaceans affect their ecology e the amphipod example? J. Exp. Mar. Biol. Ecol. 300, 373–408.
- Martellini, T., Guerranti, C., Scopetani, C., Ugolini, A., Chelazzi, D., Cincinelli, A., 2018. A snapshot of microplastics in the coastal areas of the Mediterranean Sea. Trends Analytical Chem. 109, 173–179. https://doi.org/10.1016/j.trac.2018.09.028.
- Martellini, T., Russo, A., Cincinelli, A., Santini, S., Lofrumberto, C., Baini, M., Ciattini, S., Conti, L., Mostardini, F., Mercatelli, L., Ugolini, A., 2023. Bioplastics on marine sandy shores: effects on the key species *Talitrus saltator* (Montagu, 1808). Sci. Total Environ. 876, 162811 https://doi.org/10.1016/j.scitotenv.2023.162811.
- Melchor-Martínez, E.M., Macías-Garbett, R., Alvarado-Ramírez, L., Araújo, R.G., Sosa-Hernández, J.E., Ramírez-Gamboa, D., Parra-Arroyo, L., Alvarez, A.G., Monteverde, R.P.B., Cazares, K.A.S., Reyes-Mayer, A., Yáñez Lino, M., Iqbal, H.M.N., Parra-Saldívar, R., 2022. Towards a circular economy of plastics: an evaluation of the systematic transition to a new generation of bioplastics. Polymers 14, 1203. https://doi.org/10.3390/polym14061203.
- Mengoni, A., Focardi, A., Bacci, G., Ugolini, A., 2013. High genetic diversity and variability of bacterial communities associated with the sandhopper *Talitrus saltator* (Montagu) (Crustacea, Amphipoda). Estuar. Coast. Shelf Sci. 131, 75–82.
- Mosleh, Y. Y., Paris-Palacios, S., Ahmed, M.T., Mahmoud, F.M., Osman, M.A., Biagianti-Risbourg, S., 2007. Effects of chitosan on oxidative stress and metallothioneins in aquatic worm Tubifex tubifex (Oligochaeta, Tubificidae). Chemosphere 67, 167–175.
- Muzzarelli, R., Tarsi, R., Filippini, O., Giovanetti, E., Biagini, G., Varaldo, P., 1990. Antimicrobial properties of N-carboxybutyl chitosan. Antimicrob. Agents Chemother. 34, 2019–2023. https://doi.org/10.1128/AAC.34.10.2019.
- Nakamura, S., Yumioka, J., Kachi, S., Baba, Y., Kawai, S., 2022. Bacterial and fungal gut microbiota of supralittoral talitrid amphipods feeding on brown macroalgae and paper. PloS One 17, e0279834.
- Olabarria, C., Incera, M., Garrido, J., Rodil, I.F., Rossi, F., 2009. Intraspecific diet shift in *Talitrus saltator* inhabiting exposed sandy beaches. Estuar. Coast. Shelf Sci. 84, 282–288.
- Palluault, M., 1954. Notes ecologiques sur le *Talitrus saltator*. Arch. Zool. Experim. Géner. 91, 105–129.
- Porri, F., Hill, J., McQuaid, C., 2011. Associations in ephemeral systems: the lack of trophic relationships between sandhoppers and beach wrack. Mar. Ecol. Progr. Ser. 426, 253–262. https://doi.org/10.3354/meps08951.
- Priyadarshi, R., Rhim, J., 2020. Chitosan-based biodegradable functional films for food packaging applications. Innov. Food Sci. Emerg. Technol. 62, 102346 https://doi. org/10.1016/j.ifset.2020.102346.
- Qi, L., Xu, Z., Jiang, X., Hu, C., Zou, X., 2004. Preparation and antibacterial activity of chitosan nanoparticles. Carbohydr. Res. 339, 2693–2700. https://doi.org/10.1016/j. carres.2004.09.007.
- Rainbow, P.S., 2002. Trace metal concentrations in aquatic invertebrates: why and so what? Environ. Pollut. 120, 497–507. https://doi.org/10.1016/S0269-7491(02) 00238-5.
- Rainbow, P.S., Amiard-Triquet, C., Amiard, J., Smith, B., Best, S., Nassiri, Y., Langston, W., 1999. Trace metal uptake rates in crustaceans (amphipods and crabs) from coastal sites in NW Europe differentially enriched with trace metals. Mar. Ecol. Progr. Series 183, 189–203. https://doi.org/10.3354/meps183189.
- Rochman, C.M., Hoh, E., Hentschel, B.T., Kaye, S., 2013. Long-term field measurement of sorption of organic contaminants to five types of plastic pellets: implications for plastic marine debris. Environ. Sci. Technol. 47, 1646–1654. https://doi.org/ 10.1021/es303700s.
- Roller, S., Covill, N., 1999. The antifungal properties of chitosan in laboratory media and apple juice. Int. J. Food Microbiol. 47, 67–77. https://doi.org/10.1016/s0168-1605 (99)00006-9.
- Roman-Doval, R., Torres-Arellanes, S., Tenorio-Barajas, A., Gomez-Sanchez, A., Valencia-Lezcano, A.A., 2023. Chitosan: properties and its application in agriculture in context of molecular weight. Polymers 15, 2867. https://doi.org/10.3390/ polym15132867.
- Rose, D.L.G., Hudson, M.D., Bray, S., Gaca, P., 2024. Assessment of the estuarine shoreline microplastics and mesoplastics of the river Itchen, Southampton (UK) for contaminants and for their interaction with invertebrate fauna. Environ. Sci. Pollut. Res. 31, 6437–6459. https://doi.org/10.1007/s11356-023-31396-6.
- Scopetani, C., Cincinelli, A., Martellini, T., Lombardini, E., Ciofini, A., Fortunati, A., Pasquali, V., Ciattini, S., Ugolini, A., 2018. Ingested microplastic as a two-way transporter for PBDEs in *Talitrus saltator*. Environ. Res. 167, 411–417. https://doi. org/10.1016/j.envres.2018.07.030.

Serra-Gonçalves, C., Lavers, J.L., Bond, A.L., 2019. Global review of beach debris monitoring and fututre recommendations. Environ. Sci. Technol. 53, 1258–12167.

- Shruti, V.C., Kutralam-Muniasamy, G., 2019. Bioplastics: missing link in the era of microplastics. Sci. Total Environ. 697, 134139 https://doi.org/10.1016/j.scitotenv. 2019.134139.
- Siegel, S., Castellan Jr., N.J., 1988. Nonparametric Statistics for the Behavioral Sciences, 2nd edition. McGrawHill, New York. https://doi.org/10.1177/ 014662168901300212.
- Teuten, E.L., Saquing, J.M., Knappe, D.R.U., Barlaz, M.A., Jonsson, S., Björn, A., Rowland, S.J., Thompson, R.C., Galloway, T.S., Yamashita, R., Ochi, D., Watanuki, Y., Moore, C., Viet, P.H., Tana, T.S., Prudente, M., Boonyatumanond, R., Zakaria, M.P., Akkhavong, K., Ogata, Y., Hirai, H., Iwasa, S., Mizukawa, K.,

A. Ugolini et al.

Hagino, Y., Imamura, A., Saha, M., Takada, H., 2009. Transport and release of chemicals from plastic to the environment and to wildlife. Philos. Trans R. Soc. B: Biol. Sci. 364, 2027–2045.

- Thompson, R.C., Olsen, Y., Mitchell, R.P., Davis, A., Rowland, S.J., John, A.W.G., McGonigle, D., Russel, A.E., 2004. Lost at sea: where is all the plastic? Science 304, 838.
- Tong, H., Zhong, X., Duan, Z., Yi, X., Cheng, F., Xu, W., Yang, X., 2022. Micro- and nanoplastics released from biodegradable and conventional plastics during degradation: formation, aging factors, and toxicity. Sci. Total Environ. 833, 155275 https://doi.org/10.1016/j.scitotenv.2022.155275.
- Tosetto, L., Brown, C., Williamson, J.E., 2016. Microplastics on beaches: ingestion and behavioural consequences for beachhoppers. Mar. Biol. 163, 199. https://doi.org/ 10.1007/s00227-016-2973-0.
- Tsai, G.J., Su, W.H., 1999. Antibacterial activity of shrimp chitosan against *Escherichia coli*. J. Food Protect. 62, 239–243. https://doi.org/10.4315/0362-028x-62.3.239.

Ugolini, A., Ungherese, G., 2012. Sandhoppers as bioindicators of anthropogenic influence on Mediterranean sandy beaches. In: Life in the Mediterranean Sea: A Look at Habitat Changes, Environmental Science, Engineering and Technology Ser. Nova Science Publ, New York, pp. 413–443.

- Ugolini, A., Borghini, F., Calosi, P., Bazzicalupo, M., Chelazzi, G., Focardi, S., 2004. Mediterranean *Talitrus saltator* (Crustacea, Amphipoda) as a biomonitor of heavy metals contamination. Mar. Pollut. Bull. 48, 526–532. https://doi.org/10.1016/j. marpolbul.2003.10.002.
- Ugolini, A., Borghini, F., Focardi, S., Chelazzi, G., 2005. Heavy metals accumulation in two syntopic sandhopper species: *Talitrus saltator* (Montagu) and *Talorchestia ugolinii* Bellan Santini and Ruffo. Mar. Pollut. Bull. 50, 1328–1334. https://doi.org/ 10.1016/j.marpolbul.2005.04.041.
- Ugolini, A., Perra, G., Focardi, S., Somigli, S., Martellini, T., Cincinelli, A., 2012. Sandhopper *Talitrus saltator* (Montagu) as a bioindicator of contamination by polycyclic aromatic hydrocarbons. Bull. Environ. Contam. Toxicol. 89, 1272–1276.
- Ugolini, A., Ungherese, G., Ciofini, M., Lapucci, A., Camaiti, M., 2013. Microplastic debris in sandhoppers. Estuar. Coast. Shelf Sci. 129, 19–22. https://doi.org/ 10.1016/j.ecss.2013.05.026.
- Ungherese, G., Ugolini, A., 2009. Sandhopper solar orientation as a behavioural biomarker of trace metals contamination. Environ. Pollut. 157, 1360–1364. https:// doi.org/10.1016/j.envpol.2008.11.038.

- Ungherese, G., Baroni, D., Focardi, S., Ugolini, A., 2010a. Trace metal contamination of Tuscan and eastern Corsican coastal supralitoral zones: the sandhopper *Talitrus saltator* (Montagu) as a biomonitor. Ecotoxicol. Environ. Safety 73, 1919–1924. https://doi.org/10.1016/j.ecoenv.2010.06.021.
- Ungherese, G., Mengoni, A., Somigli, S., Baroni, D., Focardi, S., Ugolini, A., 2010b. Relationship between heavy metals pollution and genetic diversity in Mediterranean populations of the sandhopper *Talitrus saltator* (Montagu) (Crustacea, Amphipoda). Environ. Pollut. 158, 1638–1643. https://doi.org/10.1016/j.envpol.2009.12.007.
- Ungherese, G., Cincinelli, A., Martellini, T., Ugolini, A., 2016. Biomonitoring of PCBs contamination in the supralittoral environment using the sandhopper *Talitrus saltator* (Montagu). Chem. Ecol. 32, 301–311. https://doi.org/10.1080/ 02757540.2015.1135908.
- Venâncio, C., Lopes, I., Oliveira, M., 2022. Bioplastics: known effects and potential consequences to marine and estuarine ecosystem services. Chemosphere 136810. https://doi.org/10.1016/j.chemosphere.2022.136810.

Wang, Y., Xie, W., 2010. Synthesis of cationic starch with a high degree of substitution in an ionic liquid. Carbohydr. Polym. 80, 1172–1177. https://doi.org/10.1016/j. carbool.2010.01.042.

- Xing, K., Zhu, X., Peng, X., Qin, S., 2015. Chitosan antimicrobial and eliciting properties for pest control in agriculture: a review. Agron. Sustain. Develop. 35, 569–588.
- Xu, X., Wong, C.Y., Tam, N.F.Y., Lo, H.S., Cheung, S.G., 2020. Microplastics in invertebrates on soft shores in Hong Kong: influence of habitat, taxa, and feeding mode. Sci. Total Environ. 715, 136999.
- Yilmaz Atay, H., 2019. Antibacterial activity of chitosan-based systems. In: Functional Chitosan, Jana, S., Jana, S. Eds, Cap. 15. Springer, Singapore, pp. 457–489. https:// doi.org/10.1007/978-981-15-0263-7_15.
- Yilmaz Atay, H., Çelik, E., 2017. Investigations of antibacterial activity of chitosan in the polymeric composite coatings. Progr. Organic Coat. 102, B, 194–200.
- Zar, H.J., 1999. Biostatistical Analysis, 4th edition. Prentice Hall, Upper Saddle River. Zhang, X., Yuan, J., Li, F., Xiang, J., 2021. Chitin synthesis and degradation in crustaceans: a genomic view and application. Mar. Drugs 19, 153. https://doi.org/ 10.3390/md19030153.
- Zhu, X., 2021. The plastic cycle an unknown branch of the carbon cycle. Front. Mar. Sci. 7, 609243 https://doi.org/10.3389/fmars.2020.609243.