

Genetic causes of heart failure with preserved ejection fraction: emerging pharmacological treatments

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Received 31 May 2022; revised 13 November 2022; accepted 26 November 2022



Graphical Abstract



Genetic causes of HFpEF: distinct vs. shared mechanisms with 'mainstream' HFpEF, and specific treatments. While the pathophysiology and phenotypes of specific aetiologies are unique, several disease features overlap with mainstream HFpEF. Therefore, each of these conditions may offer important insights and identify potential therapeutic targets which may translate from rare to more prevalent forms of disease. ASO, antisense oligonucleotide; ATTR, transthyretin amyloidosis; siRNA, small-interfering RNA; TTR, transthyretin.

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⁺ C.R. sadly passed away in October 2022. This work is dedicating to his lasting scientific, academic, and human legacy.

Abstract

Heart failure with preserved ejection fraction (HFpEF) is a major driver of cardiac morbidity and mortality in developed countries, due to ageing populations and the increasing prevalence of comorbidities. While heart failure with reduced ejection fraction is dominated by left ventricular impairment, HFpEF results from a complex interplay of cardiac remodelling, peripheral circulation, and concomitant features including age, hypertension, obesity, and diabetes. In an important subset, however, HFpEF is subtended by specific diseases of the myocardium that are genetically determined, have distinct pathophysiology, and are increasingly amenable to targeted, innovative treatments. While each of these conditions is rare, they collectively represent a relevant subset within HFpEF cohorts, and their prompt recognition has major consequences for clinical practice, as access to dedicated, disease-specific treatments may radically change the quality of life and outcome. Furthermore, response to standard heart failure treatment will generally be modest for these individuals, whose inclusion in registries and trials may dilute the perceived efficacy of treatments targeting mainstream HFpEF. Finally, a better understanding of the molecular underpinnings of monogenic myocardial disease may help identify therapeutic targets and develop innovative treatments for selected HFpEF phenotypes of broader epidemiological relevance. The field of genetic cardiomyopathies is undergoing rapid transformation due to recent, groundbreaking advances in drug development, and deserves greater awareness within the heart failure community. The present review addressed existing and developing therapies for genetic causes of HFpEF, including hypertrophic cardiomyopathy, cardiac amyloidosis, and storage diseases, discussing their potential impact on management and their broader implications for our understanding of HFpEF at large.

Keywords

Heart failure with preserved ejection fraction (HFpEF) • Hypertrophic cardiomyopathy • Fabry disease • Danon disease • Cardiac amyloidosis • Gene therapy • Myosin inhibitors • Tafamidis • Drug development

Introduction

Heart failure with preserved ejection fraction (HFpEF) is a broad term encompassing a spectrum of conditions with diverse pathophysiology, currently defined as heart failure (HF) with left ventricular ejection fraction (LVEF) > 50%, in the absence of prior evidence of systolic dysfunction.¹⁻⁴ The complementary definition of HF with normal LVEF and elevated left ventricular (LV) filling pressure at rest or during exercise is possibly more useful conceptually.^{5,6} In addition, based on the haemodynamic response to exercise, HFpEF can be diagnosed in the presence of an end-expiratory pulmonary artery wedge pressure (PAWP) \geq 25 mmHg at peak exercise, a PAWP/cardiac output slope >2 mmHg/L/min, or a respiratory-averaged mean pulmonary artery pressure (PAP) > 30 mmHg at peak exercise with a total pulmonary resistance >3 Wood Units and PAWP \geq 20 mmHg.⁵

The evolving understanding of HFpEF has led to the identification of different clinical phenotypes with only partially overlapping pathophysiology. While the classic model of heart failure with reduced ejection fraction (HFrEF) is largely a *cardiocentric* condition dominated by functional LV impairment, 'mainstream' HFpEF results from a complex interaction of the heart, the peripheral circulation, and the environment, with major roles played by age, female gender, and comorbidities such as obesity, diabetes, hypertension, and inflammation.^{3,4} These differences are reflected in outcomes: HFrEF patients die predominantly of worsening HF and sudden cardiac death, while in HFpEF, non-cardiac mortality due to dysfunction of multiple organ systems plays an important role.⁷

In a peculiar subset of patients, however, HFpEF is subtended by genetic conditions that have distinct pathophysiology and are increasingly amenable to targeted treatments^{3,4} (*Graphical Abstract* and *Table 1*). These conditions belong to the realm of primary myocardial disease i.e. cardiomyopathies—with clinical profiles and outcomes that are largely independent of extra-cardiac factors: as such, they rather follow the *cardiocentric* paradigm of HF generally associated with systolic dysfunction. Despite sharing several features of mainstream HFpEF, such as hypertrophy, interstitial fibrosis, myocardial inflammation, microvascular dysfunction, and disordered autophagy and mitochondria, their phenotypes are unique.^{3,6,8-10} These aetiologies present in individuals that are generally younger (with the exception of cardiac amyloidosis), have little or none of the classic co-morbidities associated with HFpEF, and show no predilection for the female gender. Rather, a male preponderance is observed, particularly in X-linked conditions such as Fabry disease (FD).⁸ A long history of neglected complaints and a suggestive pedigree are common. Importantly, these patients exhibit peculiar electrocardiogram (ECG) and multimodality imaging phenotypes (Figure 1) and may present with distinct extra-cardiac red flags such as carpal tunnel syndrome in cardiac amyloidosis or angiokeratomas and proteinuria in FD. For all, diastolic dysfunction represents the main cause of HF symptoms, with systolic dysfunction occurring only in selected subsets following progressive myocardial replacement fibrosis and myocyte loss.⁸ Marked atrial remodelling and dysfunction are frequent complications, triggering and perpetuating atrial fibrillation, further worsening HF symptoms. Cardioembolic risk is generally high irrespective of CHA₂DS₂-VASc score, and ischaemic events may occur even in patients who are in sinus rhythm and adequately treated with anticoagulants.¹⁰ Systemic manifestations can exacerbate HF symptoms, such as involvement of the peripheral nervous system with chronotropic incompetence in FD, or muscular involvement in glycogenosis, and mortality is largely due to refractory HF or arrhythmic events.^{10,11}

While each of these conditions may be uncommon, they collectively represent a relevant subset of HFpEF cohorts, in whom prompt recognition has major consequences for the clinical practice: while the response to standard treatment will generally be modest for these individuals, access to dedicated therapies may radically change their quality of life and outcome.^{8,11,12} On the other hand, by enhancing our understanding of cardiac pathophysiology, genetic cardiomyopathies are shedding light on novel therapeutic targets that may be relevant to the broader HFpEF world.^{3,4} Once considered a niche of limited epidemiological interest, the field of genetic cardiomyopathies is undergoing rapid transformation due to recent, groundbreaking advances in drug development.¹² We here review the existing and developing

	НСМ	ATTRv amyloidosis	Fabry	Danon
Prevalence of heart failure	35%	50%-80%	20%	20%
LVEF at presentation	75%	40%–50%	70%	20%
Progressive LVEF dysfunction	t in 10% with end-stage evolution	↓ in advanced phases	↓ in a small subgroup in advanced disease	οZ
Left ventricular volumes	= →	→	II	$\stackrel{\rightarrow}{\rightarrow}$
Longitudinal systolic function	→	††	→	\rightarrow
Longitudinal strain	↓ (no apical sparing)	↓ (with apical sparing)	Units (inferolateral impairment)	U (possible apical sparing)
Chamber stiffness	¢	11	←	ţ
Parietal stiffness	Ļ		\	$\downarrow\downarrow$
Myocardial relaxation	П	→	→	2
Stroke volume	Π	<pre>tt and fixed</pre>	II	\rightarrow
Inotropic reserve	П	††	2	2
NT-proBNP	Ļ	111	÷	ż
LVH pattern and ancillary features	Asymmetric LVH, LVOTO, elongated mitral leaflets, abnormal morphology and insertion of papillary muscles	Thickening of AV valves, interatrial septum and RV free wall. Granular-sparkling appearance of myocardial tissue	Disproportionate hypertrophy of papillary muscles, 'Binary sign'	Massive concentric LVH
Cardiac magnetic resonance findings	Midwall/transmural LGE in the most hypertrophic segments. Mildly increased native T1 and ECV	Diffuse subendocardial or transmural LGE, increased native T1 and ECV	Midwall LGE in basal inferolateral segment. Reduced native T1, normal ECV	Extensive LGE with mid-septum sparing. Reduced native T1, normal ECV
Typical ECG features	High QRS voltages, q waves	Normal-Jow* QRS voltages, AV blocks, q waves	High QRS voltages, short PQ interval	High QRS voltages, pre-excitation, AV blocks
LVEF, left ventricular ejection fraction;	LVH, left ventricular hypertrophy; LVOTO, left ventricul	ar outflow tract obstruction; AV, atrioventricular; RV, right	ventricle; LGE, late gadolinium enhancemen	;; ECV, extra-cellular volume.

 Table 1
 Functional profile of genetic cardiomyopathies associated with heart failure with preserved ejection fraction

therapies for the principal genetic causes of HFpEF and discuss their potential implications for a deeper understanding of mechanisms leading to cardiac dysfunction and HF.

Hypertrophic cardiomyopathy

Hypertrophic cardiomyopathy (HCM) is the most common genetic cardiomyopathy, with a prevalence of 1:500 in the general population, often caused by sarcomere protein gene mutations.^{10,13,14} The phenotype is a complex blend including asymmetric LV hypertrophy not explained by abnormal loading conditions, mitral valve abnormalities, microvascular remodelling, and myocardial disarray. At the molecular level, sarcomere gene mutations directly or indirectly cause a hypercontractile phenotype resulting in downstream consequences ranging from cardiomyocyte calcium overload and electrophysiological remodelling to mitochondrial dysfunction, energy depletion, myocardial ischaemia, and replacement fibrosis.^{15,16} Clinically, the disease is characterized by arrhythmic propensity and limiting symptoms of varying severity, either in the context of hyperdynamic LV contraction promoting dynamic outflow obstruction or-in advanced disease-of LV dysfunction associated with extensive myocardial fibrosis.^{10,13,14} Notably, the natural history of HCM may be influenced by environmental factors such as obesity, partly mediated by secondary worsening of the intrinsic microvascular abnormalities typical of the disease, as well as obesity-related inflammation.¹⁷ The role of the environment on phenotypic expression and the course of HCM, however, remains largely unresolved.¹⁸

To date, treatment of obstructive HCM rests on beta-blocker or verapamil, with the addition of disopyramide as a negative inotrope.¹⁶ Patients with persistent symptoms require invasive options including surgical myectomy and alcohol septal ablation.^{10,13} When HF develops due to LV fibrosis and dysfunction, culminating in the so-called end-stage phase, standard HF therapy including cardiac resynchronization shows modest efficacy.¹³ To date, none of the classic HF treatments, including renin–angiotensin–aldosterone system inhibitors and spironolactone, have been shown to alter disease progression or improve outcome. Until recently, HCM was an orphan condition in urgent need of drugs targeting its core pathophysiological mechanisms.¹⁶

Our current understanding of HCM pathogenesis originates from patients and experimental animals carrying cardiac beta myosin heavy chain gene (MHY7) mutations. These are gain-of-function variants leading to enhanced myosin activation and an increase in the number of heads involved in cardiac contraction at any given cycle.^{15,19} All pathophysiological and clinical manifestations of HCM are now believed to derive from this molecular 'original sin', or along similar pathways in HCM patients with variants in other sarcomeric genes, such as MYBPC, which also result, albeit indirectly, in myosin over-activation.¹⁵ Following seminal work from James Spudich's group, this realization led to the concept of myosin inhibition as a potential therapeutic strategy for HCM, representing the mirror image of myosin stimulation with omecamtiv mecarbil in HFrEF.^{20,21} The first-in-class cardiac myosin inhibitor, mavacamten, showed promising results in a mouse model of MHY7-associated HCM, in which the drug markedly attenuated the functional and structural phenotype, including disarray and interstitial fibrosis²² (Figure 2). The drug, therefore, entered human experimentation, focusing on patients with symptomatic obstructive HCM. This choice was based on the well-established relation between LV hypercontractility, dynamic obstruction, and limiting symptoms, as well as on the consistent and measurable symptomatic improvement following gradient relief.¹³

The landmark Phase 3 EXPLORER-HCM trial was successfully completed in 2020,²³ and led to Food and Drug Administration (FDA) approval of mavacamten in April 2022. The study randomized 251 patients with symptomatic obstructive HCM to mavacamten or placebo, on a background therapy including beta-blockers or calcium antagonists.²³ The primary endpoint was a composite assessing variation in oxygen consumption at cardiopulmonary exercise testing and perceived symptom burden. After a 30-week treatment period, patients were considered to have achieved the endpoint if their peak oxygen consumption (pVO₂) increased \geq 1.5 mL/kg/min (compared with baseline) with an improvement of at least one New York Heart Association (NYHA) class or ≥3.0 mL/kg/min with no worsening of NYHA class. Overall, 37% of patients in the mavacamten arm reached the primary endpoint compared with 17% in the placebo arm (P = 0.0005). In addition, there was a consistent benefit of mavacamten across all secondary endpoints, including pVO₂ increase vs. baseline (1.4 vs. -0.1 mL/kg/min), NYHA functional class (\geq 1 NYHA class improvement in 65% vs. 31% in the placebo arm), relief of post-exercise LV outflow gradient (an almost 50 mmHg reduction, compared with no change in the placebo arm), symptomatic status and quality of life (13.6 point improvement in Kansas City Cardiomyopathy Questionnaire, compared with 4.2 in the placebo arm; P < 0.001 for all comparisons).²³

Clinical and haemodynamic improvement with mavacamten was independent of age, gender, and genetic status, and was associated with favourable biomarker profile, as shown by a marked reduction in serum N-terminal pro-B-type natriuretic peptide (NT-proBNP) and highsensitivity troponin I. Mavacamten was generally well tolerated, and its safety profile was not different from placebo. Notably, the average reduction in LVEF in the mavacamten arm was only 4%, and most patients remained well above a normal threshold of systolic function. Only seven patients experienced a temporary reduction in LVEF to values <50%; three had protocol-driven temporary treatment discontinuation during the 30-week treatment, while in four, the event occurred at end-of-treatment visit. In all patients, LVEF recovered to normal values following washout, except one who experienced only partial recovery (LVEF 50%) due to a procedural complication following atrial fibrillation ablation.²³ A recent interim analysis based on the patients enrolled in the EXPLORER-HCM Long-Term Extension study has shown sustained benefit and safety of mavacamten beyond 1 year.²⁴ In the recently presented VALOR-HCM trial (A study to evaluate mavacamten in adults with symptomatic obstructive HCM who are eligible for septal reduction therapy, NCT04349072), mavacamten was effective in reducing the need for invasive septal reduction therapies in obstructive patients referred to surgical centres due to severe symptoms.²⁵

Another myosin inhibitor, aficamten, is currently undergoing Phase 3 experimentation in the international, multicentre, randomized Sequoia HCM trial, also focusing on symptomatic obstructive HCM patients. The study has been designed on the wake of the successful Phase II randomized placebo-controlled sequential cohort REDWOOD-HCM trial, showing excellent symptomatic gradient relief and symptomatic improvement in patients with obstructive HCM.²¹ Aficamten has a similar mechanism of action to mavacamten, although the binding site and half-life are different.²⁶

As myosin inhibitors start to penetrate clinical practice, relevant gaps in knowledge remain, calling for prudent use in the real world. Such caution is well reflected in the titration algorithm recommended in the mavacamten prescription label (https://www.accessdata.fda.gov/drugsatfda_ docs/label/2022/214998s000lbl.pdf). Despite their excellent safety

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Figure 1 Electrocardiogram and imaging features of specific causes of heart failure with preserved ejection fraction. (A-E) Sarcomeric obstructive hypertrophic cardiomyopathy in a 45-year-old female. (A) Electrocardiogram presents high QRS voltages and diffuse repolarization abnormalities with diphasic-negative T waves in precordial leads V4–V6. (B) Echocardiography shows significant hypertrophy of basal and mid-septum favouring left ventricular outflow tract obstruction (not shown). (C) Strain imaging shows reduced longitudinal strain velocities in the basal-medium segments of the interventricular septum and anterior wall. (D) At cardiac magnetic resonance, a transmural area of late gadolinium enhancement of the midanterior interventricular septum is present; septal native T1 mapping values are within the normal range in (E) (1000 ms; normal values 960-1040 ms). (F-J) Transthyretin amyloidosis cardiac amyloidosis in a 79-year-old male. (F) Electrocardiogram shows sinus rhythm with left atrial enlargement and low-normal QRS voltages in V4–V6. (G) Echocardiography shows concentric left ventricular hypertrophy with granular-sparkling appearance of the myocardium, thickening of interatrial septum and atrioventricular valves, and left atrial enlargement. (H) Strain rate imaging is characterized by a significant diffuse reduction of longitudinal strain velocities with apical sparing. (1) At cardiac magnetic resonance diffuse LGE more evident in subendocardial myocardial layers is evident, with significantly increased septal T1 mapping values (/) (1100 ms; normal values 960–1040 ms). (K–O) Fabry disease in a 51-year-old male. (K) ECG shows sinus rhythm with a short PQ interval (118 ms), high QRS voltages, and diffuse repolarization abnormalities with negative T waves in V4–V6 leads. (L) Echocardiography shows concentric hypertrophy with thickening of right ventricular free wall and septal endocardium. (M) At strain rate, imaging significantly reduced longitudinal strain velocities in inferolateral and anterolateral mid-basal segments are present. (N) Cardiac magnetic resonance shows midwall LGE in the lateral basal segments with reduced septal native T1 mapping values (890 ms; normal values 960–1040 ms). Pseudo-normalized native T1 mapping values (980 ms; normal values 960–1040 ms) are present in the lateral segment corresponding to LGE area (O).

record so far, there are no data regarding the long-term safety and efficacy of myosin inhibitors, and particular caution is required in the identification of slow metabolizers, at risk of excessing reduction in LVEF during titration or in the presence of intervening modifiers such as atrial fibrillation. Furthermore, whether myosin inhibitors may benefit symptomatic patients with non-obstructive HCM remains unresolved.²⁷ The most important question, however, is whether myosin inhibitors may fulfil the promise of exerting in patients the disease-modifying impact shown in experimental HCM models,^{22,28} and thus prevent long-term myocardial energy depletion, fibrosis, and LV remodelling, ultimately impacting outcome (*Figure 2*). Because of the slow progression and low event rates typical of HCM, answers to this question will require long exposure times. However, based on the solid scientific rationale and encouraging initial results, hopes are justified for the first class of drugs specifically addressing the core disease mechanism of HCM.

Hereditary transthyretin amyloidosis

Cardiac amyloidosis is characterized by extra-cellular deposition of misfolded proteins in the heart with the pathognomonic histological property of green birefringence when a tissue specimen is examined under cross-polarized light after staining with Congo red.^{29–33} While more than 30 known proteins are capable of aggregating as amyloid *in vivo*, only nine accumulate in the myocardium producing significant cardiac disease, and more than 98% of currently diagnosed cardiac amyloidosis result from fibrils composed of either monoclonal immunoglobulin light chains (AL) or transthyretin (ATTR), either in its genetically altered (i.e. hereditary—ATTRv) or wild-type (ATTRwt) form.³² Two main mechanisms of tissue and organ damage occur in amyloidosis: chronic infiltration and acute proteotoxic effect of circulating precursors and



Figure 2 Myosin inhibitors: mechanism of action and clinical benefits. Molecular mechanism of action of myosin inhibitors, with demonstrated structural effects in animal models and clinical benefits in clinical trials.

non-amyloid aggregates. The latter is particularly evident in AL but also occurs in ATTRv and ATTRwt.³⁰ Myocardial infiltration by rigid, spaceoccupying amyloid fibrils leads to increased stiffness and dysfunction of atrial and ventricular myocardium. Infiltration is generally diffuse but can be mainly subendocardial or with patchy areas of transmural involvement; involvement of the valves, pericardium, and conduction system is common.³⁰ Cardiac amyloidosis is classically considered a form of restrictive cardiomyopathy. However, a restrictive haemodynamic profile is limited to the advanced phases of the disease, in which a progressive increase in parietal and chamber stiffness leads to an upward and leftward shift in the end-diastolic pressure-volume relationship with concomitant declines in stroke volume, cardiac output, and, frequently, arterial blood pressure.²⁹ Very high NT-proBNP values result, which represent an important clinical red flag.³² Parallel declines in stroke volume and end-diastolic volume explain why LVEF is often preserved during the course of the disease, despite the fact that myocardial contraction is reduced—as shown by reduced LV longitudinal function and strain.²⁹ When a restrictive physiology has fully developed, the duration of diastole and increased filling pressures have a limited impact on ventricular filling. As a result, stroke volume is virtually fixed and cardiac output becomes crucially dependent on heart rate.^{29,33}

Previously considered a rare disease, cardiac amyloidosis is considered today rather underdiagnosed than rare. In systematic screening studies on HFpEF, up to 12% of patients are found to have ATTR amyloidosis.^{30,31} The clinical manifestations are heterogeneous and range from an exclusive or dominant neurological phenotype, with motor sensory polyneuropathy and autonomic disturbances, to a pure cardiologic phenotype, with HFpEF as the most frequent manifestation.^{30,33} Organ involvement is strictly driven by the underlying genetic variant: 1122L, T60A, and I68L are classic examples of 'cardiac' mutations.³¹ Notably, between 3% and 4% of the African-American population in the USA harbour I122L, albeit penetrance is low.³⁴

Treatment of cardiac amyloidosis has long been limited to the treatment and prevention of complications including HF, arrhythmias, conduction disturbances, and thromboembolism. Recent advances, however, have shifted the focus towards halting or delaying amyloid deposition by specific disease-modifying drugs, ^{31,33,35} which are becoming increasingly available for ATTRv (as well as ATTRwt) amyloidosis. Conversely, treatment of AL amyloidosis largely rests on therapies addressing the underlying haematological condition. ^{33,36}

Novel therapies act at different steps of the ATTR amyloidogenic process: some have proven effective in stabilizing circulating TTR molecules by preventing its dissociation or cleavage into amyloidogenic fragments, others in reducing the production of mutated TTR (an approach previously treated by liver transplantation) with gene silencers. Other investigational compounds include agents directed at removing amyloid fibrils, potentially reverting tissue infiltration³³ (Figure 3). Tafamidis, a kinetic TTR stabilizer, binds to the unoccupied thyroxine-binding sites of tetrameric TTR and prevents the amyloidogenic cascade.^{37,38} Tafamidis was the first drug tested in a randomized, controlled trial, and is now approved by European Medicines Agency (EMA) and FDA for ATTR cardiac amyloidosis patients who have reasonably expected survival. The ATTR-ACT study demonstrated a 30% reduction in all-cause mortality and in cardiovascular-related hospitalization and a slower decline in quality of life after 30 months of treatment in ATTRv or ATTRwt cardiomyopathy patients, compared with placebo.³⁹ ATTR-ACT can be considered the 'three first times' trial: (i) the first time a medical treatment has been shown to reduce mortality and HF hospitalization in HFpEF; (ii) the first time a medical treatment has been shown to reduce mortality and morbidity in TTR-related amyloidosis; (iii) the first time an HF drug has proven effective on a hard endpoint by acting centrally on the myocardium, rather than peripherally or by neurohormonal modulation.³⁹ Focusing on a similar therapeutic aim, AG10 is a highly selective, small-molecule TTR stabilizer. Phase I and II studies showed a good toxicity profile and stabilization of both mutant and wild-type TTR.⁴⁰ A Phase 3 efficacy and safety study to evaluate AG10 compared with placebo in subjects with symptomatic ATTR cardiomyopathy is ongoing (ClinicalTrials.gov Identifier: NCT03860935).



Figure 3 Novel treatments for transthyretin amyloidosis cardiac amyloidosis. Currently approved and forthcoming treatments for transthyretin cardiac amyloidosis represented according to their targets in the pathogenetic process of amyloidosis from transthyretin synthesis to cardiac infiltration.

Gene silencing by small-interfering RNA (siRNA) and antisense oligonucleotide (ASO) technologies causes a drastic reduction of TTR production by the liver. The siRNA patisiran and the ASO inotersen have been approved for the treatment of patients with ATTRv-related polyneuropathy, irrespective of cardiac involvement.^{35,41} Patisiran, currently undergoing Phase 3 experimentation (APOLLO-B; NCT03997383), is administered by intravenous infusion once every 3 weeks and is generally well tolerated. Other gene silencers for ATTR cardiac amyloidosis currently under investigation in Phase 3 trials include vutrisiran (HELIOS B; NCT04153149) and eplontersen (CARDIO-TTRansform; NCT04136171). Finally, the most recently devised strategy is the blockade of TTR production by genome editing by CRISPR–Cas9 technology: preliminary findings of a Phase 1 study (NCT04601051) are encouraging.⁴¹

Storage diseases

Storage diseases represent a group of inherited errors of metabolism characterized by the accumulation of intracellular material in various organs and tissues, including the myocardium. Those presenting with cardiomyopathy include Fabry, Danon, and PRKAG2-related disease,^{42–44} often referred to as HCM phenocopies, due to overlapping morphological and functional features such as cardiac wall thickening, diastolic dysfunction, left atrial enlargement, microvascular ischaemia, and myocardial fibrosis. However, they only exceptionally develop LV outflow tract obstruction and often show distinct cardiac and extra-cardiac red flags that may help suspect a rare disease and exclude sarcomeric HCM.^{8,42}

Fabry disease

Fabry disease is an X-linked disorder caused by pathogenic variants in the GLA gene, leading to deficient activity of the enzyme

 α -galactosidase A. Such deficit leads to progressive intralysosomal storage of globotriaosylceramide (Gb3) in affected tissues, including heart, vessels, kidneys, and peripheral nervous system.^{11,45} Systemic manifestations, mainly occurring in males, include small fibres neuropathic pain, hypo-anhidrosis with heat and cold intolerance, angiokeratomas, gastrointestinal disturbances, and cornea verticillata. Renal involvement with albuminuria occurs early in young male patients while cardiomyopathy and premature cerebrovascular accidents usually manifest in the third or fourth decade.⁴⁵ Some missense variants, including p.N215S and p.F113L, are associated with residual α -galactosidase A activity, leading to a late-onset phenotype predominantly affecting the heart.¹¹ Heterozygous female patients usually present at an older age and with milder clinical manifestations compared with males. However, in the case of skewed x chromosome inactivation, female patients may also develop severe phenotypes and present a severe clinical course.⁴⁶ Cardiac involvement in FD is characterized by progressive thickening of cardiac walls mimicking sarcomeric disease (Figure 1): in consecutive cohorts, FD accounts for up to 1% of patients with an initial diagnosis of HCM.^{11,47} Mitral and aortic valve regurgitation are common due to leaflet infiltration; LV dysfunction may ensue due to progressive fibrosis and myocyte loss. Atrioventricular block, atrial fibrillation, and ventricular arrhythmias manifest late during the course of the cardiomyopathy.¹¹

Available treatments for FD include enzyme-replacement therapies (ERTs) (agalsidase-alfa and agalsidase-beta) and the oral pharmacological chaperone (migalastat) (*Graphical Abstract*).^{11,45} Collectively, these therapies have radically improved the outcome of FD patients, largely due to their positive effects on the progression of renal disease. However, early start of treatment is critical in order to obtain satisfactory results.^{45,48,49} Furthermore, long-term studies and registry data demonstrate that ERT may halt or slow the progression of cardiac disease, reducing the rate of cardiovascular events.⁵⁰ Regression of mild LV hypertrophy has been occasionally reported, with some evidence that LV hypertrophy may be prevented by early treatment.⁴⁹ In ERT-naïve patients with initial signs of cardiomyopathy, 1 year of ERT stabilized LV mass index and attenuated reduction in T1, an early magnetic resonance imaging (MRI) marker of disease.⁵¹

Chaperone molecules such as migalastat are iminosugars that bind to the catalytic domain of α -Gal A promoting its proper folding and trafficking to the lysosome thus increasing enzymatic activity.⁵² While ERT is indicated in all patients with FD, migalastat can be administered only to patients with amenable variants (i.e. mutations that are responsive to the drug in a dedicated *in vitro* assay, as listed in https://galafold.com/hcp/ amenability). In clinical trials and open-label extension studies, the efficacy of migalastat is comparable with ERT in reducing the overall progression of the disease. In addition, migalastat treatment is associated with a sustained decrease of LV mass index, suggesting distinct diseasemodification properties at the myocardial level.^{11,52}

New therapies for FD include second-generation ERTs, substrate reduction therapies, and gene and mRNA therapies. Pegunigalsidase-alpha is a pegylated form of α -Gal A produced in a PlantCell Ex system, characterized by a much longer circulatory half-life and increased heart and kidney uptake compared with currently available ERTs.⁵³ In the Phase III BRIDGE trial (NCT03018730), patients switched from agalsidase-alfa to pegunigalsidase-alfa showed slower progression of renal failure (estimated glomerular filtration rate slope improved from -5.1 to 0.23 mL/min/ 1.73 m²/year in both males and females).⁵³ Other Phase III clinical trials (NCT02795676; NCT03018730. NCT03180840) evaluating pegunigalsidase-alfa treatment are ongoing. Substrate reduction therapy is also being pursued, by oral iminosugars inhibiting glycosphingolipid synthesis and lowering the cellular content of Gb3. Two agents, venglustat and lucerastat, are currently under investigation in Phase II and III clinical trials, respectively.⁵⁴ In addition, cardiotropic vectors specifically targeting myocardial tissue have been developed for gene therapy of FD. Results from Phase I/II STAAR trial with the adenoviral vector the isaralgagene-civaparvovec were promising, leading to increased α-Gal A activity in four patients, ranging from 2- to 15-fold above normal mean values. In one patient, with sustained 15-fold enzymatic activity at Week 52, ERT was withdrawn (https://clinicaltrials.gov/ct2/show/NCT04046224). Adult male patients with classic FD enrolled in the 4D-310 trial presented a sustained increase of α-Gal A activity and significant decrease in lyso-Gb3 levels, together with promising cardiac improvement in terms of T1 mapping increase at MRI (https://ir.4dmoleculartherapeutics.com/static-files/ bb460939-74bf-44bb-a0e0-caf85c0ac58c). Finally, initial experience with human α -Gal A mRNA encapsulated with lipid nanoparticles, administered to mice and non-human primates, has led to a significant increase in α -Gal A levels in the liver, heart, and kidney.⁵⁵

Cardiac glycogenoses

Danon disease is an X-linked dominant lysosomal storage disorder, caused by a deficiency of the lysosome-associated membrane protein-2 (LAMP2), a crucial promoter of lysosomal biogenesis. Such deficit leads to the accumulation of autophagic vacuoles containing amorphic material and glycogen deposition in cardiac and skeletal myocytes.^{42,56,57} The prevalence of Danon disease among patients with HCM phenotype is 4%–6% in paediatric cohorts and 0.7%–4.0% in adults.^{56–58} Males present during childhood with severe systemic involvement including ocular, hepatic, cognitive, and musculoskeletal manifestations.^{42,56} Danon cardiomyopathy is severe, characterized by rapidly progressive, massive biventricular hypertrophy, leading to early death or

transplant.^{42,56} In females, conversely, cardiac involvement is milder, due to the lyonization process, and may present in the absence of systemic manifestations around the third or fourth decade of life, often indistinguishable from sarcomeric HCM. The ECG classically shows a short PR interval, pre-excitation, and high QRS voltages with deep T-wave inversion.^{59,60} Atrioventricular conduction abnormalities have been observed in up to 15% of patients.^{60,61} A gene therapy programme is currently being developed for Danon disease using a recombinant adeno-associated virus serotype 9 (AAV9) containing the human lysosome-associated membrane protein 2 isoform B (LAMP2B) transgene (RP-A501). In a Phase I trial, a single intravenous dose was generally well tolerated and led to cardiac LAMP2B gene expression associated with preliminary evidence of cardiac and extracardiac benefits.⁶²

Finally, PRKAG2-related cardiomyopathy is a rare disease accounting for 0.23%-1% of consecutive patients with an initial diagnosis of HCM.^{43,63} Pathogenic variants in the gamma-2 regulatory subunit of AMP-activated protein kinase (PRKAG2) cause cardiomyopathy through an anomalous activation of adenosine monophosphate (AMP) kinase, leading to increased glucose uptake and cytosolic accumulation of glycogen metabolism by-products (polyglucan and amylopectin).⁶³ The disease, transmitted with autosomal dominant inheritance pattern, is characterized by severe HCM with LV pre-excitation and a high prevalence of atrioventricular blocks and ventricular tachyarrhythmias.⁴³ Although specific pharmacological treatments are lacking, a correct diagnosis is essential for management, as these patients often present with syncope due to atrioventricular block, but may also develop life-threatening ventricular tachyarrhythmias. Aggressive monitoring of arrhythmias and timely device implantation is warranted even at an early age. Conversely, ablation of accessory pathways should be considered only in selected, symptomatic patients, due to frequent recurrences and the risk of advanced atrioventricular block. 43,64

Implications for the broader heart failure with preserved ejection fraction scenario

Randomized trials involving HFpEF patients have been notoriously challenging, particularly when LVEF exceeds 60%–65%,⁶⁵ and only recently have large randomized outcome studies advanced the field, as spironolactone and sacubitril/valsartan have shown benefit for patients in the lower end of the HFpEF LVEF spectrum, and sodium-glucose cotransporter 2 (SGLT2) inhibitors have shown definite improvement on survival.^{66,67} One explanation for the disappointing outcome of many HFpEF trials includes the lack of clear therapeutic rationale for using drugs such as angiotensin receptor blockers, in contrast to their use in HFrEF, where neurohormonal activation is a clear driver of prognosis.⁶⁷ Furthermore, the HFpEF syndrome seems to represent a far more heterogeneous group than HFrEF, due to the 'contamination' by subsets of patients not likely to respond to the interventions. It has been estimated that up to 15%–20% of patients enrolled in HFpEF outcome trials may have a more specific disease causing the syndrome.⁶⁸ In trials enrolling thousands of patients at hundreds of multinational sites, little attention had been paid over the years to identifying such patients prior to enrollment. However, the availability of specific therapies has now created a highly compelling rationale to identify potential outliers, not only during clinical practice but also during the screening period of clinical trials. In the near future, artificial intelligence and machine



Figure 4 Dissecting the complexities of the heart failure with preserved ejection fraction spectrum: the cardiomyopathy mindset. Flowchart illustrating the approach to the differential diagnosis of potentially treatable Heart failure with preserved ejection fraction causes, through a cardiomyopathic mindset. For each diagnostic step (in vertical), red flags suggesting a specific cardiomyopathy (in horizontal) underlying the initial heart failure with preserved ejection fraction presentation are reported. LV, left ventricle; ECV, extra-cellular volume; LGE, late gadolinium enhancement; CK, creatine kinase; AST, aspartate transaminase; ALT, alanine transaminase; AV, atrioventricular; LVH, left ventricular hypertrophy; GLS, global longitudinal strain; ATTRv, hereditary transthyretin amyloidosis; LAMP2, lysosomal-associated membrane protein 2.

learning techniques may provide the best solution to define HFpEF subsets and differentiate monogenic diseases within a number of broader phenotypes,⁶⁹ as well as highlight common mechanisms across subsets.⁷⁰ In contemporary practice, the 'red flag' concept continues to represent the most effective approach alerting clinicians to certain clinical signs requiring further evaluation with targeted imaging, biomarkers, and genetic tests.⁸ Often referred to as the *cardiomyopathy mindset (Figure 4*), such an approach can be extended to the evaluation of HFpEF, in order to reach a timely suspicion of specific aetiologies based on dedicated algorithms.⁷¹ As the general awareness for rare cardiac diseases increases in the medical community, earlier recognition, and improvements in the management of these patients will inevitably follow.⁸

Notably, the emergence of treatments targeted at genetic causes of HFpEF has other, far-reaching implications, by providing a unique insight into cardiac pathophysiology and identifying novel targets of potential relevance for the HF community. Treatments that have been developed for a well-defined niche of HFpEF may turn out to be beneficial to wider groups of patients sharing similar mechanisms of disease—as witnessed in the last two decades in the field of oncology.⁷² A case in point is represented by HFpEF with hypercontractile phenotype. Recent evidence shows that, among adults without cardiovascular disease, LVEF in the supranormal range is associated with a higher risk of adverse cardiovascular outcomes. In a large MRI-based study, participants with LVEF above the 75th percentile had an increased risk of adverse events independent of their baseline risk profile and other potential confounders.⁷³ Consistently, a combined analysis of the UK Biobank cohort and the BioMe cohort (using MRI and echocardiography-based assessment of LVEF, respectively) demonstrated an increased risk of mortality and higher burden of HF symptoms and natriuretic peptide elevation among individuals with LVEF >70%.⁷⁴ Intriguingly, higher LVEF was associated with a greater risk of events only among those with low stroke volume

index—a combination reminiscent of initial/mild HCM phenotypes.⁷⁵ Because sarcomere genetic variants are present in 1:200 in the general population, it is tempting to postulate that a number of these individuals may harbour unrecognized HCM.⁷⁶ In the remainder, late-onset, hypertrophic/hypercontractile phenotypes are now thought to result from the interplay of ageing, polygenic predisposition, and acquired modifications occurring both in cardiomyocytes (e.g. titin hypophosphorylation) and the extra-cellular matrix.⁷⁷ Although cardiomyocytes are known to exhibit a number of age-related modifications, including hypertrophy and changes in contraction and relaxation,⁷⁸ there is still limited understanding of how ageing may promote HF. Among other insults, ageing cardiomyocytes are exposed to somatic mutations, i.e. genomic changes that escape DNA repair mechanisms over time. Very recent evidence suggests that cardiomyocytes accumulate age-related single nucleotide variants at rates higher than neurons and lymphocytes, but similar to hepatocytes.⁷⁹ In organs characterized by active metabolism, such as the heart, cells more often tend to develop somatic mutations and become polyploid, possibly in order to endure oxidative stress by increasing cell size and metabolic production. However, polyploidization may also promote alterations in the transcriptome and metabolome and might ultimately result in dysfunction, with HFpEF as the final result.⁷⁹

Rosch et al.⁸⁰ recently provided a thorough characterization of HFpEF phenotypes including haemodynamic and histological features. Intriguingly, they found that patients with LVEF in the low-normal range (50%–60%) showed features overlapping with HFrEF, including larger cardiac dimensions and higher levels of myocardial fibrosis. Conversely, patients with supranormal LVEF (>60%) had smaller but thicker ventricles, less fibrosis, and increased preload and afterload sensitivity in the context of enhanced diastolic and systolic stiffness. Such behaviour is consistent with the observation that drugs that are successful in HFrEF, fail to show benefit in HFpEF with supranormal LVEF,⁸¹ suggesting that this is a different patient subset altogether. In such individuals, it is tempting to speculate that myosin inhibitors may play a role by countering the core pathophysiological and clinical abnormalities, irrespective of their cause (genetic vs. acquired). Many gaps in knowledge remain. Promisingly, however, just as many years ago the empiric treatment recommendations for 'diastolic HF' were informed by the approach to HCM, today a favourable Phase 2 trial signal in non-obstructive HCM²⁷ has led to a pilot trial of mavacamtem in patients with HFpEF.⁸²

Storage diseases are providing invaluable insight regarding the role of deranged autophagy and myocardial inflammation in inherited and acquired cardiac diseases. Autophagy regulation is central in normal and diseased myocardium, and changes in autophagy activation are implicated in several cardiac conditions including cardiomyopathies and HF.^{83,84} Lysosomal storage disorders represent a unique model to improve our knowledge of lysosomal function and develop new therapeutic strategies targeting autophagy regulation.⁸⁵ In addition to well-established autophagy inducers like rapamycin and other agents regulating the mTOR signalling pathways, there is increasing evidence that drugs effective in the management of HFpEF, such as SGLT2 inhibitors, and other cardiovascular drugs including beta-adrenergic blockers, calcium channel blockers, and amiodarone, exert part of their therapeutic actions by influencing myocyte autophagy.^{86,87} Interestingly, while small non-coding RNAs are among the main regulators of autophagic processes, autophagy itself may represent a key step in the mechanism of action of therapeutic ASO.^{88,89} Furthermore, autophagy is interlinked with unfolded protein response, an intracellular process triggered by the accumulation and aggregation of unfolded proteins in the endoplasmic reticulum lumen.^{90,91} A persistent activation of this process may lead to secondary protein metabolism alterations, perturbation of autophagy, and tissue inflammation, ultimately promoting cell dysfunction and death.⁹⁰ While the accumulation of misfolded or abnormally degraded proteic material is a common feature in genetic disorders specifically associated with abnormal contractile or cytoskeletal protein synthesis, unfolded protein response has been increasingly associated with the most prevalent acquired cardiovascular conditions, including hypertensive heart disease, ischaemic heart disease, and HF.⁹¹ In a recent experimental HFpEF model,⁹² nitric oxide synthase induction was shown to cause deficient activity of unfolded protein response effectors potentially associated with further interstitial accumulation of proteins. Today, targeting the unfolded protein response represents a promising therapeutic approach for a broad spectrum of conditions including cancer and neurodegenerative disorders, as well as cardiac disease.⁹¹

Conclusions

A conceptually important subset of HFpEF is due to genetic causes amenable to treatment. While their epidemiological relevance may seem small compared with mainstream HFpEF, the clinical implications of a timely diagnosis for individual patients and their families cannot be overemphasized. Furthermore, the theoretical implications of such disease paradigms for broader sections of the HFpEF population are just beginning to emerge. As the field rapidly evolves, the concept that deep phenotyping of HFpEF patients will enable personalized therapies targeting different underlying mechanisms is emerging powerfully. It is hoped that from the privileged observation site of monogenic disease, novel solutions will surface for the whole HF community. The concept of treating mechanisms, rather than clinical diagnosis, is already a reality in the world of oncology and may represent the near future for cardiologists.

Acknowledgements

The *Graphical Abstract* and *Figure 4* have been partially created by M.P. with officially licensed Biorender application.

Funding

There were no sources of funding for this paper.

Conflict of interest: I.O. is a consultant for Bristol Myers Squibb, Cytokinetics, Sanofi Genzyme, Amicus, Menarini International, Rocket Pharma, and Tenaya and has received research grants from BMS, Cytokinetics, Sanofi Genzyme, Shire Takeda, Amicus, Menarini International, Boston Scientific, Menarini International, and Bayer. J.E.U. serves as the Chair of a DMC for Cytokinetics and is a consultant to Cardurion, Bayer, Reprieve, Sequana, and Alleviant Medical. M.P. has received advisory board honoraria Myers Squibb and speaker fees from Sanofi Genzyme, Shire Takeda, Amicus Therapeutics, Pfizer, and Bristol Myers Squibb. C.R. has served as a speaker and has received consulting fees from Pfizer, Alnylam, and Eidos and has received research grants from Pfizer.

Data availability

Not applicable.

References

- McDonagh TA, Metra M, Adamo M, Gardner RS, Baumbach A, Böhm M, et al. 2021 ESC guidelines for the diagnosis and treatment of acute and chronic heart failure: developed by the task force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) with the special contribution of the Heart Failure Association (HFA) of the ESC. Eur Heart J 2021;42:3599–3726. https://doi.org/ 10.1093/eurheartj/ehab368
- Heidenreich PA, Bozkurt B, Aguilar D, Allen LA, Byun JJ, Colvin MM, et al. 2022 AHA/ ACC/HFSA guideline for the management of heart failure: a report of the American College of Cardiology/American Heart Association Joint Committee on clinical practice guidelines. *Circulation* 2022;**145**:e895–e1032. https://doi.org/10.1161/CIR. 000000000001063
- Gevaert AB, Kataria R, Zannad F, Sauer AJ, Damman K, Sharma K, et al. Heart failure with preserved ejection fraction: recent concepts in diagnosis, mechanisms and management. Heart 2022;108:1342–1350. https://doi.org/10.1136/heartjnl-2021-319605
- Lam CSP, Voors AA, de Boer RA, Solomon SD, van Veldhuisen DJ. Heart failure with preserved ejection fraction: from mechanisms to therapies. *Eur Heart J* 2018;39: 2780–2792. https://doi.org/10.1093/eurheartij/ehy301
- Baratto C, Caravita S, Soranna D, Faini A, Dewachter C, Zambon A, et al. Current limitations of invasive exercise hemodynamics for the diagnosis of heart failure with preserved ejection fraction. *Circ Heart Fail* 2021;**14**:e007555. https://doi.org/10.1161/ CIRCHEARTFAILURE.120.007555
- Pieske B, Tschöpe C, de Boer RA, Fraser AG, Anker SD, Donal E, et al. How to diagnose heart failure with preserved ejection fraction: the HFA-PEFF diagnostic algorithm: a consensus recommendation from the Heart Failure Association (HFA) of the European Society of Cardiology (ESC). Eur Heart J 2019;40:3297–3317. https://doi.org/10.1093/ eurheartj/ehz641
- Vaduganathan M, Patel RB, Michel A, Shah SJ, Senni M, Gheorghiade M, et al. Mode of death in heart failure with preserved ejection fraction. J Am Coll Cardiol 2017;69: 556–569. https://doi.org/10.1016/j.jacc.2016.10.078
- Rapezzi C, Arbustini E, Caforio AL, Charron P, Gimeno-Blanes J, Helio T, et al. Diagnostic work-up in cardiomyopathies: bridging the gap between clinical phenotypes and final diagnosis. A position statement from the ESC Working Group on Myocardial and Pericardial Diseases. *Eur Heart J* 2013;**34**:1448–1458. https://doi.org/10.1093/ eurheartj/ehs397
- Smiseth OA, Morris DA, Cardim N, Cikes M, Delgado V, Donal E, et al. Multimodality imaging in patients with heart failure and preserved ejection fraction: an expert consensus document of the European Association of Cardiovascular Imaging. Eur Heart J Cardiovasc Imaging 2022;23:e34–e61. https://doi.org/10.1093/ehjci/jeab154
- Elliott PM, Anastasakis A, Borger MA, Borggrefe M, Cecchi F, Charron P, et al. 2014 ESC guidelines on diagnosis and management of hypertrophic cardiomyopathy: the task force for the diagnosis and management of hypertrophic cardiomyopathy of the European Society of Cardiology (ESC). Eur Heart J 2014;35:2733–2779. https://doi. org/10.1093/eurhearti/ehu284
- Pieroni M, Moon JC, Arbustini E, Barriales-Villa R, Camporeale A, Vujkovac AC, et al. Cardiac involvement in Fabry disease: JACC review topic of the week. J Am Coll Cardiol 2021;77:922–936. https://doi.org/10.1016/j.jacc.2020.12.024

- Helms AS, Thompson AD, Day SM. Translation of new and emerging therapies for genetic cardiomyopathies. JACC Basic Transl Sci 2021;7:70–83. https://doi.org/10.1016/j. jacbts.2021.07.012
- Ommen SR, Mital S, Burke MA, Day SM, Deswal A, Elliott P, et al. 2020 AHA/ACC guideline for the diagnosis and treatment of patients with hypertrophic cardiomyopathy: executive summary: a report of the American College of Cardiology/American Heart Association Joint Committee on clinical practice guidelines. *Circulation* 2020; 142:e533–e557. https://doi.org/10.1161/CIR.00000000000938
- Maron BJ, Desai MY, Nishimura RA, Spirito P, Rakowski H, Towbin JA, et al. Diagnosis and evaluation of hypertrophic cardiomyopathy: JACC state-of-the-art review. J Am Coll Cardiol 2022;79:372–389. https://doi.org/10.1016/j.jacc.2021.12.002
- Trivedi DV, Adhikari AS, Sarkar SS, Ruppel KM, Spudich JA. Hypertrophic cardiomyopathy and the myosin mesa: viewing an old disease in a new light. *Biophys Rev* 2018;**10**: 27–48. https://doi.org/10.1007/s12551-017-0274-6
- Ammirati E, Contri R, Coppini R, Cecchi F, Frigerio M, Olivotto I. Pharmacological treatment of hypertrophic cardiomyopathy: current practice and novel perspectives. Eur J Heart Fail 2016;18:1106–1118. https://doi.org/10.1002/ejhf.541
- Fumagalli C, Maurizi N, Day SM, Ashley EA, Michels M, Colan SD, et al. Association of obesity with adverse long-term outcomes in hypertrophic cardiomyopathy. JAMA Cardiol 2020;5:65–72. https://doi.org/10.1001/jamacardio.2019.4268
- Finocchiaro G, Magavern E, Sinagra G, Ashley E, Papadakis M, Tome-Esteban M, et al. Impact of demographic features, lifestyle, and comorbidities on the clinical expression of hypertrophic cardiomyopathy. J Am Heart Assoc 2017;6:e007161. https://doi.org/ 10.1161/JAHA.117.007161
- Spudich JA. Three perspectives on the molecular basis of hypercontractility caused by hypertrophic cardiomyopathy mutations. *Pflugers Arch Eur J Physiol* 2019;**471**:701–717. https://doi.org/10.1007/s00424-019-02259-2
- Teerlink JR, Diaz R, Felker GM, McMurray JJV, Metra M, Solomon SD, et al. Cardiac myosin activation with omecamtiv mecarbil in systolic heart failure. N Engl J Med 2021;384: 105–116. https://doi.org/10.1056/NEJMoa2025797
- Morelli C, Ingrasciotta G, Jacoby D, Masri A, Olivotto I. Sarcomere protein modulation: the new frontier in cardiovascular medicine and beyond. *Eur J Intern Med* 2022;**102**:1–7. https://doi.org/10.1016/j.ejim.2022.04.020
- Green EM, Wakimoto H, Anderson RL, Evanchik MJ, Gorham JM, Harrison BC, et al. Heart disease: a small-molecule inhibitor of sarcomere contractility suppresses hypertrophic cardiomyopathy in mice. Science 2016;351:617–621. https://doi.org/10.1126/ science.aad3456
- Olivotto I, Oreziak A, Barriales-Villa R, Abraham TP, Masri A, Garcia-Pavia P, et al. Mavacamten for treatment of symptomatic obstructive hypertrophic cardiomyopathy (EXPLORER-HCM): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2020;396:759–769. https://doi.org/10.1016/S0140-6736(20)31792-X
- Rader F, Choudhury L, Saberi S, Fermin D, Wheeler MT, Abraham TP, et al. Long-term safety of mavacamten in patients with obstructive hypertrophic cardiomyopathy: interim results of the Mava-Long Term Extension (LTE) study. J Am Coll Cardiol 2021; 77:532. https://doi.org/10.1016/S0735-1097(21)01891-X
- Desay MY, Owens A, Geske JB, Wolski K, Naidu SS, Smedira NG, et al. Myosin inhibition in patients with obstructive hypertrophic cardiomyopathy referred for septal reduction therapy. J Am Coll Cardiol 2022;80:95–108.
- Chuang C, Collibee S, Ashcraft L, Wang W, Vander Wal M, Wang X, et al. Discovery of aficamten (CK-274), a next-generation cardiac myosin inhibitor for the treatment of hypertrophic cardiomyopathy. J Med Chem 2021;64:14142–14152. https://doi.org/10. 1021/acs.jmedchem.1c01290
- Ho CY, Mealiffe ME, Bach RG, Bhattacharya M, Choudhury L, Edelberg JM, et al. Evaluation of mavacamten in symptomatic patients with nonobstructive hypertrophic cardiomyopathy. J Am Coll Cardiol 2020;75:2649–2660. https://doi.org/10.1016/j.jacc. 2020.03.064
- Lehman SJ, Crocini C, Leinwand LA. Targeting the sarcomere in inherited cardiomyopathies. Nat Rev Cardiol 2022;19:353–363. https://doi.org/10.1038/s41569-022-00682-0
- Maurer MS, Elliott P, Comenzo R, Semigran M, Rapezzi C. Addressing common questions encountered in the diagnosis and management of cardiac amyloidosis. *Circulation* 2017;**135**:1357–1377. https://doi.org/10.1161/CIRCULATIONAHA.116. 024438
- Ruberg FL, Grogan M, Hanna M, Kelly JW, Maurer MS. Transthyretin amyloid cardiomyopathy: JACC state-of-the-art review. J Am Coll Cardiol 2019;73:2872–91. https://doi. org/10.1016/j.jacc.2019.04.003
- Aimo A, Merlo M, Porcari A, Georgiopoulos G, Pagura L, Vergaro G, et al. Redefining the epidemiology of cardiac amyloidosis. A systematic review and meta-analysis of screening studies. Eur J Heart Fail 2022. https://doi.org/10.1002/ejhf.2532
- Benson MD, Buxbaum JN, Eisenberg DS, Merlini G, Saraiva MJM, Sekijima Y, et al. Amyloid nomenclature 2018: recommendations by the International Society of Amyloidosis (ISA) nomenclature committee. *Amyloid* 2018;25:215–219. https://doi. org/10.1080/13506129.2018.1549825
- Garcia-Pavia P, Rapezzi C, Adler Y, Arad M, Basso C, Brucato A, et al. Diagnosis and treatment of cardiac amyloidosis: a position statement of the ESC working group on myocardial and pericardial diseases. Eur Heart J 2021;42:1554–1568. https://doi.org/ 10.1093/eurheartj/ehab072

- Quarta CC, Buxbaum JN, Shah AM, Falk RH, Claggett B, Kitzman DW, et al. The amyloidogenic V122I transthyretin variant in elderly black Americans. N Engl J Med 2015; 372:21–29. https://doi.org/10.1056/NEJMoa1404852
- Aimo A, Castiglione V, Rapezzi C, Franzini M, Panichella G, Vergaro G, et al. RNA-targeting and gene editing therapies for transthyretin amyloidosis. Nat Rev Cardiol 2022;19:655–667. https://doi.org/10.1038/s41569-022-00683-z
- Palladini G, Merlini G. What is new in diagnosis and management of light chain amyloidosis? Blood 2016;**128**:159–168. https://doi.org/10.1182/blood-2016-01-629790
- Hammarstrom P, Schneider F, Kelly JW. Trans-suppression of misfolding in an amyloid disease. Science 2001;293:2459–2462. https://doi.org/10.1126/science.1062245
- Coelho T, Merlini G, Bulawa CE, Fleming JA, Judge DP, Kelly JW, et al. Mechanism of action and clinical application of tafamidis in hereditary transthyretin amyloidosis. *Neurol Ther* 2016;5:1–25. https://doi.org/10.1007/s40120-016-0040-x
- Maurer MS, Schwartz JH, Gundapaneni B, Elliott PM, Merlini G, Waddington-Cruz M, et al. Tafamidis treatment for patients with transthyretin amyloid cardiomyopathy. N Engl J Med 2018;379:1007–1016. https://doi.org/10.1056/NEJMoa1805689
- Judge DP, Heitner SB, Falk RH, Maurer MS, Shah SJ, Witteles RM, et al. Transthyretin stabilization by AG10 in symptomatic transthyretin amyloid cardiomyopathy. J Am Coll Cardiol 2019;74:285–295. https://doi.org/10.1016/j.jacc.2019.03.012
- Gillmore JD, Gane E, Taubel J, Kao J, Fontana M, Maitland ML, et al. CRISPR-Cas9 in vivo gene editing for transthyretin amyloidosis. N Engl J Med 2021;385:493–502. https://doi. org/10.1056/NEJMoa2107454
- Pereira NL, Grogan M, Dec GW. Spectrum of restrictive and infiltrative cardiomyopathies: part 1 of a 2-part series. J Am Coll Cardiol 2018;71:1130–1148. https://doi.org/ 10.1016/j.jacc.2018.01.016
- Lopez-Sainz A, Dominguez F, Lopes LR, Ochoa JP, Barriales-Villa R, Climent V, et al. Clinical features and natural history of PRKAG2 variant cardiac glycogenosis. J Am Coll Cardiol 2020;**76**:186–197. https://doi.org/10.1016/j.jacc.2020.05.029
- Lotan D, Salazar-Mendiguchía J, Mogensen J, Rathore F, Anastasakis A, Kaski J, et al. Clinical profile of cardiac involvement in Danon disease: a multicenter European registry. Circ Genom Precis Med 2020;13:e003117. https://doi.org/10.1161/CIRCGEN.120. 003117
- Ortiz A, Germain DP, Desnick RJ, Politei J, Mauer M, Burlina A, et al. Fabry disease revisited: management and treatment recommendations for adult patients. Mol Genet Metab 2018;123:416–427. https://doi.org/10.1016/j.ymgme.2018.02.014
- Echevarria L, Benistan K, Toussaint A, Dubourg O, Hagege AA, Eladari D, et al. X-chromosome inactivation in female patients with Fabry disease. *Clin Genet* 2016; 89:44–54. https://doi.org/10.1111/cge.12613
- Doheny D, Srinivasan R, Pagant S, Chen B, Yasuda M, Desnick RJ, et al. Fabry disease: prevalence of affected males and heterozygotes with pathogenic GLA mutations identified by screening renal, cardiac and stroke clinics, 1995–2017. J Med Genet 2018;55: 261–268. https://doi.org/10.1136/jmedgenet-2017-105080
- Linhart A, Germain DP, Olivotto I, Akhtar MM, Anastasakis A, Hughes D, et al. An expert consensus document on the management of cardiovascular manifestations of Fabry disease. *Eur J Heart Fail* 2020;**22**:1076–1096. https://doi.org/10.1002/ejhf.1960
- Germain DP, Elliott PM, Falissard B, Fomin VV, Hilz MJ, Jovanovic A, et al. The effect of enzyme replacement therapy on clinical outcomes in male patients with Fabry disease: a systematic literature review by a European panel of experts. *Mol Genet Metab Rep* 2019; 19:100454. https://doi.org/10.1016/j.ymgmr.2019.100454
- Ortiz A, Abiose A, Bichet DG, Cabrera G, Charrow J, Germain DP, et al. Time to treatment benefit for adult patients with Fabry disease receiving agalsidase β: data from the Fabry registry. J Med Genet 2016;53:495–502. https://doi.org/10.1136/jmedgenet-2015-103486
- Nordin S, Kozor R, Vijapurapu R, Augusto JB, Knott KD, Captur G, et al. Myocardial storage, inflammation, and cardiac phenotype in Fabry disease after one year of enzyme replacement therapy. *Circ Cardiovasc Imaging* 2019;**12**:e009430. https://doi.org/10.1161/ CIRCIMAGING.119.009430
- Germain DP, Hughes DA, Nicholls K, Bichet DG, Giugliani R, Wilcox WR, et al. Treatment of Fabry's disease with the pharmacologic chaperone migalastat. N Engl J Med 2016;375:545–555. https://doi.org/10.1056/NEJMoa1510198
- 53. Linhart A, Dostalova G, Nicholls K, West M, Tøndel C, Jovanovic A, et al. Switching from agalsidase alfa to pegunigalsidase alfa for treating Fabry disease—one year of treatment: data from BRIDGE—a phase III open label study. *Mol Genet Metab* 2020;**129**:S98–S99. https://doi.org/10.1016/j.ymgme.2019.11.249
- Peterschmitt MJ, Crawford NPS, Gaemers SJM, Ji AJ, Sharma J, Pham TT. Pharmacokinetics, pharmacodynamics, safety, and tolerability of oral venglustat in healthy volunteers. *Clin Pharmacol Drug Dev* 2021;**10**:86–98. https://doi.org/10.1002/ cpdd.865
- 55. Zhu X, Yin L, Theisen M, Zhuo J, Siddiqui S, Levy B, et al. Systemic mRNA therapy for the treatment of Fabry disease: preclinical studies in wild-type mice, Fabry mouse model, and wild-type non-human primates. Am J Hum Genet 2019;**104**:625–637. https://doi.org/10.1016/j.ajhg.2019.02.003
- Maron BJ, Roberts WC, Arad M, Haas TS, Spirito P, Wright GB, et al. Clinical outcome and phenotypic expression in LAMP2 cardiomyopathy. JAMA 2009;301:1253–1259. https://doi.org/10.1001/jama.2009.371

- Yang Z, McMahon CJ, Smith LR, Bersola J, Adesina AM, Breinholt JP, et al. Danon disease as an underrecognized cause of hypertrophic cardiomyopathy in children. *Circulation* 2005;**112**:1612–1617. https://doi.org/10.1161/CIRCULATIONAHA.105.546481
- Fu L, Luo S, Cai S, Hong W, Guo Y, Wu J, et al. Identification of LAMP2 mutations in early-onset Danon disease with hypertrophic cardiomyopathy by targeted nextgeneration sequencing. Am J Cardiol 2016;**118**:888–894. https://doi.org/10.1016/j. amjcard.2016.06.037
- Konrad T, Sonnenschein S, Schmidt FP, Mollnau H, Bock K, Ocete BQ, et al. Cardiac arrhythmias in patients with Danon disease. Europace 2017;19:1204–1210. https://doi. org/10.1093/europace/euw215
- Darden D, Hsu JC, Tzou WS, von Alvensleben JC, Brooks M, Hoffmayer KS, et al. Fasciculoventricular and atrioventricular accessory pathways in patients with Danon disease and preexcitation: a multicenter experience. *Heart Rhythm* 2021;**18**: 1194–1202. https://doi.org/10.1016/j.hrthm.2021.03.024
- Liu Y, Wang F, Chen X, Liang Y, Deng H, Liao H, et al. Fasciculoventricular pathways responsible for ventricular preexcitation in patients with Danon disease. Circ Arrhythm Electrophysiol 2018;11:e006704. https://doi.org/10.1161/CIRCEP.118.006704
- Greenberg B, Eshraghian E, Battiprolu P, Ricks D, Yarabe P, Schwartz J, et al. Abstract 10727: results from first-in-human clinical trial of RP-A501 (AAV9:LAMP2B) gene therapy treatment for Danon disease. *Circulation* 2021;**144**:A10727. https://doi.org/10. 1161/circ.144.suppl_1.10727
- Arad M, Benson DW, Perez-Atayde AR, McKenna WJ, Sparks EA, Kanter RJ, et al. Constitutively active AMP kinase mutations cause glycogen storage disease mimicking hypertrophic cardiomyopathy. J Clin Invest 2002;109:357–362. https://doi.org/10. 1172/JCl0214571
- 64. Thevenon J, Laurent G, Ader F, Laforêt P, Klug D, Duva Pentiah A, et al. High prevalence of arrhythmic and myocardial complications in patients with cardiac glycogenosis due to PRKAG2 mutations. Europace 2017;19:651–659. https://doi.org/10.1093/europace/ euw067
- Udelson JE, Stevenson LW. The future of heart failure diagnosis, therapy, and management. *Circulation* 2016;**133**:2671–2686. https://doi.org/10.1161/CIRCULATIONAHA. 116.023518
- Solomon SD, McMurray JJV, Anand IS, Ge J, Lam CSP, Maggioni AP, et al. Angiotensin– neprilysin inhibition in heart failure with preserved ejection fraction. N Engl J Med 2019; 381:1609–1620. https://doi.org/10.1056/NEJMoa1908655
- Braunwald E. Gliflozins in the management of cardiovascular disease. N Engl J Med 2022; 386:2024–2034. https://doi.org/10.1056/NEJMra2115011
- Hahn VS, Yanek LR, Vaishnav J, Ying W, Vaidya D, Lee YZJ, et al. Endomyocardial biopsy characterization of heart failure with preserved ejection fraction and prevalence of cardiac amyloidosis. JACC Heart Fail 2020;8:712–724. https://doi.org/10.1016/j.jchf.2020.04. 007
- Paulus WJ, Zile MR. From systemic inflammation to myocardial fibrosis: the heart failure with preserved ejection fraction paradigm revisited. *Circ Res* 2021;**128**:1451–1467. https://doi.org/10.1161/CIRCRESAHA.121.318159
- Sabbah MS, Fayyaz AU, de Denus S, Felker GM, Borlaug BA, Dasari S, et al. Obese-inflammatory phenotypes in heart failure with preserved ejection fraction. *Circ Heart Fail* 2020;**13**:e006414. https://doi.org/10.1161/CIRCHEARTFAILURE.119. 006414
- Oghina S, Bougouin W, Bézard M, Kharoubi M, Komajda M, Cohen-Solal A, et al. The impact of patients with cardiac amyloidosis in HFpEF trials. JACC Heart Fail 2021;9: 169–178. https://doi.org/10.1016/j.jchf.2020.12.005
- Krause DS, Van Etten RA. Tyrosine kinases as targets for cancer therapy. N Engl J Med 2005;353:172–187. https://doi.org/10.1056/NEJMra044389
- 73. Shah S, Segar MVV, Kondamudi N, Ayers C, Chandra A, Matulevicius S, et al. Supranormal left ventricular ejection fraction, stroke volume, and cardiovascular risk: findings from population-based cohort studies. JACC Heart Fail 2022;10:583–594. https://doi.org/10.1016/j.jchf.2022.05.007
- 74. Forrest IS, Rocheleau G, Bafna S, Argulian E, Narula J, Natarajan P, et al. Genetic and phenotypic profiling of supranormal ejection fraction reveals decreased survival and

underdiagnosed heart failure. Eur J Heart Fail 2022;**24**:2118–27. https://doi.org/10. 1002/ejhf.2482

- Ho CY, Lakdawala NK, Cirino AL, Lipshultz SE, Sparks E, Abbasi SA, et al. Diltiazem treatment for pre-clinical hypertrophic cardiomyopathy sarcomere mutation carriers: a pilot randomized trial to modify disease expression. JACC Heart Fail 2015;3: 180–188. https://doi.org/10.1016/j.jchf.2014.08.003
- Semsarian C, Ingles J, Maron MS, Maron BJ. New perspectives on the prevalence of hypertrophic cardiomyopathy. J Am Coll Cardiol 2015;65:1249–1254. https://doi.org/ 10.1016/j.jacc.2015.01.019
- 77. Franssen C, González Miqueo A. The role of titin and extracellular matrix remodelling in heart failure with preserved ejection fraction. Neth Heart J 2016; 24:259–267. https://doi. org/10.1007/s12471-016-0812-z
- Sheydina A, Riordon DR, Boheler KR. Molecular mechanisms of cardiomyocyte aging. *Clin Sci* 2011;**121**:315–329. https://doi.org/10.1042/CS20110115
- Choudhury S, Huang AY, Kim J, Zhou Z, Morillo K, Maury EA, et al. Somatic mutations in single human cardiomyocytes reveal age-associated DNA damage and widespread oxidative genotoxicity. Nat Aging 2022;2:714–725. https://doi.org/10.1038/s43587-022-00261-5
- Rosch S, Kresoja KP, Besler C, Fengler K, Schöber AR, von Roeder M, et al. Characteristics of heart failure with preserved ejection fraction across the range of left ventricular ejection fraction. *Circulation* 2022;**146**:506–518. https://doi.org/10. 1161/CIRCULATIONAHA.122.059280
- Butler J, Packer M, Filippatos G, Ferreira JP, Zeller C, Schnee J, et al. Effect of empagliflozin in patients with heart failure across the spectrum of left ventricular ejection fraction. Eur Heart J 2022;43:416–426. https://doi.org/10.1093/eurheartj/ehab798
- A study of mavacamten in participants with HFpEF and elevation of NT-proBNP with or without elevation of cTnT (EMBARK-HFpEF). https://clinicaltrials.gov/ct2/show/ NCT04766892.
- Kaludercic N, Maiuri MC, Kaushik S, Fernández ÁF, de Bruijn J, Castoldi F, et al. Comprehensive autophagy evaluation in cardiac disease models. *Cardiovasc Res* 2020; 116:483–504. https://doi.org/10.1093/cvr/cvz233
- Kanamori H, Yoshida A, Naruse G, Endo S, Minatoguchi S, Watanabe T, et al. Impact of autophagy on prognosis of patients with dilated cardiomyopathy. J Am Coll Cardiol 2022; 79:789–801. https://doi.org/10.1016/j.jacc.2021.11.059
- Platt FM. Emptying the stores: lysosomal diseases and therapeutic strategies. Nat Rev Drug Discov 2018;17:133–150. https://doi.org/10.1038/nrd.2017.214
- Xu J, Kitada M, Ogura Y, Liu H, Koya D. Dapagliflozin restores impaired autophagy and suppresses inflammation in high glucose-treated HK-2 cells. *Cells* 2021;**10**:1457. https:// doi.org/10.3390/cells10061457
- van Bavel JJA, Vos MA, van der Heyden MAG. Cardiac arrhythmias and antiarrhythmic drugs: an autophagic perspective. Front Physiol 2018;9:127. https://doi.org/10.3389/ fphys.2018.00127
- Gupta SK, Thum T. Non-coding RNAs as orchestrators of autophagic processes. J Mol Cell Cardiol 2016;95:26–30. https://doi.org/10.1016/j.yjmcc.2015.11.012
- Bost JP, Ojansivu M, Munson MJ, Wesén E, Gallud A, Gupta D, et al. Novel endosomolytic compounds enable highly potent delivery of antisense oligonucleotides. *Commun Biol* 2022;**5**:185. https://doi.org/10.1038/s42003-022-03132-2
- Jensen BC, Bultman SJ, Holley D, Tang W, de Ridder G, Pizzo S, et al. Upregulation of autophagy genes and the unfolded protein response in human heart failure. Int J Clin Exp Med 2017;10:1051–1058.
- Ren J, Bi Y, Sowers JR, Hetz C, Zhang Y. Endoplasmic reticulum stress and unfolded protein response in cardiovascular diseases. *Nat Rev Cardiol* 2021L;**18**:499–521. https://doi. org/10.1038/s41569-021-00511-w
- Schiattarella GG, Altamirano F, Tong D, French KM, Villalobos E, Kim SY, et al. Nitrosative stress drives heart failure with preserved ejection fraction. Nature 2019; 568:351–356. https://doi.org/10.1038/s41586-019-1100-z
- Minamino T, Komuro I, Kitakaze M. Endoplasmic reticulum stress as a therapeutic target in cardiovascular disease. *Circ Res* 2010;**107**:1071–1082. https://doi.org/10.1161/ CIRCRESAHA.110.227819