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### Analysis of the Structural Quality of the CASD-NMR 2013 Entries

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#### Abstract

We performed a comprehensive structure validation of both automated and manually generated structures of the 10 targets of the CASD-NMR-2013 effort. We established that automated structure determination protocols are capable of reliably producing structures of comparable accuracy and quality to those generated by a skilled researcher, at least for small, single domain proteins such as the ten targets tested. The most robust results appear to be obtained when NOESY peak lists are used either as the primary input data or to augment Chemical Shift (CS) data without the need to manually filter such lists. A detailed analysis of the long-range NOE restraints generated by the different programs from the same data showed a surprisingly low degree of overlap. Additionally, we found that there was no significant correlation between the extent of the NOE restraint overlap and the accuracy of the structure. This result was surprising given the importance of NOE data in producing good quality structures. We suggest that this could be explained by the information redundancy present in NOEs between atoms contained within a fixed covalent network.

#### Introduction

In the CASD-NMR-2013 effort (see accompanying paper, Rosato et al., REF), 164 entries were submitted across ten targets. Together, these data provide for the opportunity to assess the current state of automated structure calculation methods for small- to medium-sized proteins. Automated methods (summarized in Donald and Martin 2009; Gossert et al. 2011; Guerry and Herrmann 2011; Güntert and Buchner 2015; Herrmann et al. 2002; Huang et al. 2006; Linge et al. 2003a; Williamson and Craven 2009) greatly speed up the process of NMR structure determination by providing an alternative to the manual, labor intensive step of NOESY peak assignment. In addition, it is to be expected that these procedures also provide a more unbiased interpretation of the available data. Some automated methods are even purely chemical-shift (CS) based (Shen et al. 2008), thus requiring no additional data at all and further reducing the required experimental time and associated costs in terms of equipment and labor.

For any structure, whether derived by automated methods or manually, it is imperative that the result is accurate and properly reflects the underpinning data. Ultimately, it is the aim to use the structure to explain biology, either by the researchers that generated them or indirectly by others through deposition in the PDB archive. Prompted by instances of serious errors and allegations of fabricated data underpinning published structures the wwPDB, as curator of the PDB archive, initiated a number of policy changes to improve its quality and integrity. Consequently, it is now mandatory to deposit experimental NMR restraints and assigned NMR chemical shifts. In addition, an expert NMR validation task force (NMR-VTF) has published a set of recommendations for validating NMR-derived structures and accompanying experimental data (Montelione et al. 2013).

In the following, we present a comprehensive validation report on the entries and targets of CASD-NMR-2013 in line with the NMR-VTF recommendations. The analysis draws upon the tools for validating geometric quality in relation to known structural data and the assessment of structural quality in relation to the experimental data. We used commonly available program suites CING (Doreleijers et al. 2012a), Molprobity (Lovell et al. 2003) and PSVS (Bhattacharya et al. 2006). By validating all of the entries in the same way, we are able to show the strengths and weaknesses of the various automated structure generation programs and assess the complementarity of these structure validation tools. In addition, we are able to draw broader conclusions across the range of automated structure generation routines participating in the CASD-NMR-2013 effort.

#### Methods

The CASD-NMR-2013 entries and their metadata were downloaded from the WeNMR (Wassenaar et al. 2012) web site whereas the targets were obtained from the BMRB and RCSB wwPDB repositories. We adopt the definitions of target and entry given in the accompanying paper describing the CASD-NMR-2013 data (see table 1 and the accompanying paper, Rosato et al., REF), where the target comprises all originating data, the manually derived restraints and resulting structural ensemble. An entry denotes an ensemble of conformers and the accompanying restraints generated by a specific program for a specific target.

The data sets were reorganized into a uniform directory structure to allow processing by the software analysis pipeline. Ambiguous header data and missing and damaged files were queried with the depositors, and errors discovered during processing, such as incorrect file formats, unsupported naming conventions or atom name errors in the structure files, etc, were corrected. Structure ensembles were read into CcpNmr Analysis 2.1 from PDB-type files using the Analysis structure reader (built on CcpNmr FormatConverter parsers) (Vranken et al. 2005), which disambiguated the varied naming conventions employed, and reported errors for correction. The deposited sequences were aligned automatically with those read from the target data to identify truncations. Restraint files were read into the same CCPN project using CcpNmr FormatConverter in automatic mode to identify, classify, read, and integrate the restraint files for each submission. The resulting CCPN projects each contained all data for a single target or entry, grouped so that they could be automatically extracted by CING for analysis. Due to technical limitations we were not able to incorporate data from the so-called ARIA 'swap files' that describe conformer-specific stereospecific resonance assignments. Accordingly, we were forced to reduce the precision of the deposited restraints to nonstereospecific for the one program that uses different assignments of prochiral groups in each individual structure of the ensemble.

Entries were assigned three-part names with each part separated by an underscore. The first part of the name is the target dataset. The second part is the program used in generating the entry, merging CS-ROSETTA and CS-DP-ROSETTA together as 'Web Rosetta Server'; CS-HM-ROSETTA and CS-HM-DP-ROSETTA as 'CS-HM-ROSETTA and Cheshire and Cheshire-YAPP as 'Cheshire'. The last part of the name describes what input data were used. The first character indicates curated NOE peaks (c), un-curated NOE peaks (u), raw spectra (r), and CS only (s); if RDCs were used, 'r' has been appended to the end of the data identifier. Finally, if the input sequence was truncated manually, the truncated range used is

indicated in parenthesis. Using this merging strategy, no information is lost – for example Cheshire uses only CS data and entries are listed as 'Cheshire-s', while Cheshire-YAPP uses both CS and NOE data and entries are listed as either 'Cheshire-f' or 'Cheshire-u'.

All analyses were conducted using CING (Doreleijers et al. 2012a), except where noted. CING integrates the results of a number of external programs, such as WHAT-IF (version 6) (Vriend 1990), PROCHECK NMR (Laskowski et al. 1996), Wattos (Doreleijers et al. 2009) and VASCO (Rieping and Vranken 2010), combined with its own internal routines. All analyses were conducted on residues within the well-defined areas of the reference ensembles as determined by CyRange (Table 1) (Kirchner and Güntert 2011). The analysis of Discriminating Power (DP) scores (Huang et al., 2005) and number of atomic clashes was performed using the PSVS (Bhattacharya et al. 2006) server (http://psvs-1 5-dev.nesg.org). PSVS integrates analyses from several widely-used structure quality evaluation tools, including RPF (Huang et al. 2005), PROCHECK (Laskowski et al., 1993), PROCHECK NMR (Laskowski et al., 1996), Ramachandran (Lovell et al. 2003), Verify3D (Lüthy et al. 1992), Prosa II (Sippl 1993) and Probe (Word et al. 1999). For the DP score determination the (curated) NMR peak lists and chemical shifts from the targets were used. The number of clashes was obtained as the number of disallowed atom pair overlaps  $\geq 0.4$  Å given by the Probe (Word et al. 1999) standalone program.

All-by-all RMSD values are calculated as follows. For each of the M conformers in the query ensemble, the RMSD between the backbone N, C $\alpha$  and C' atoms in the well-defined region of the reference ensemble as defined by CyRange (Kirchner and Guentert 2011) (see Table 1) of each of the N conformers in the target ensemble is calculated, yielding a list of M×N RMSD values (or  $\frac{M(M-1)}{2}$  values for convergence calculations where the same ensemble is both the query and target.) If an entry is lacking any atoms within the well-defined range, the corresponding atoms in the compared ensemble are ignored. The average value of this list of values is then reported as the mean RMSD. The accuracy of an ensemble is defined as the all-by-all RMSD of an entry to the appropriate target ensemble. Ensemble convergence values are reported as the average all-by-all RMSD of the conformers in an ensemble. The ensemble convergence calculation is rapid and independent of both the nature of the experimental input data and the structure determination algorithm method used and should not be confused with ensemble precision. Ensemble convergence often underreports the actual precision of an ensemble, as prior research showed that ensembles with a much

larger RMSD could be generated that equally well satisfied the experimental restraints (Buchner and Güntert 2015; Spronk et al. 2003). Accordingly, ensemble convergence is used here as a diagnostic criterion only.

NOE overlap values were calculated using a custom Python script, available from the authors on request. Each value was calculated as follows: each NOE in the query list (row in Figs 5C,D) was compared to each NOE in the subject list (column in Figs 5C,D) until either a match was found or there were no more NOEs in the subject list. To ensure ambiguous restraints were counted only once, the search was terminated once a match was found to any of the options. Note that handling ambiguous restraints in this way has the side effect that multiple ambiguous restraints in the query list can match a single restraint in the subject list. Heatmaps of all restraint overlaps for all ten targets are shown in the Supplementary materials.

Supplementary Table 1 lists entry and validation statistics of all 169 entries, including for reference also six entries marked 'incorrect' by the depositing authors. The CING validation reports and csv files of all the accumulated data, including restraint violation statistics and all values underpinning the Figures in this manuscript, are available from our website http://nmr.le.ac.uk/CASD-NMR-2013. A PostgreSQL database containing the complete CING analysis for all targets and entries in CASD-NMR-2013 is available from the authors upon request.

#### Results

#### Accuracy and ensemble convergence

The ensemble convergence of each of the CASD-NMR-2013 entries and target ensembles and the similarity of the entry to the corresponding target ensemble were assessed using the deviation of the backbone coordinates, expressed as the average of the pairwise root mean square deviation (RMSD) between the conformers in the reference and entry ensembles using the well-defined regions defined for the reference ensemble by CyRange (Kirchner and Güntert 2011). For the targets, the convergence ranges from 0.4 Å to 1.0 Å (Fig. 1). Therefore, we consider 1.0 Å to be an appropriate threshold to identify satisfactorily converged calculations. The median convergence for the entries is 0.6 Å with 77% of the entries having an ensemble convergence of 1.0 Å or less. Only five entries have values larger than 2 Å: three ensembles calculated from augmented CS data and two ensembles calculated from CS data only. For programs that submitted entries based on un-curated and curated NOESY peak lists, we observed a weak tendency to obtain better ensemble convergence with the curated list when the ensemble convergence for the un-curated list was above the 1 Å threshold.

In the CASD-NMR effort, the manually determined target ensemble is assumed to be the correct representation of the three-dimensional structure of the target protein. Hence, the RMSD between the target ensemble and the entry ensemble constitutes a measure of accuracy (Fig. 1A). An entry is considered to be indistinguishable from the target when the RMSD between the two ensembles is less than the sum of their ensemble convergence. Given the average ensemble convergence of 0.63 Å for the targets and 0.74 Å for the entries, a threshold of 1.5 Å appears to be reasonable. Above this threshold, any ensemble describes a structure with differences from the corresponding target beyond experimental uncertainty. Each entry was evaluated relative to the targets with the exception of ensembles marked as not converged by the programs used to generate them. The median accuracy over the entire dataset is 1.14 Å, with 71% of the entries below the 1.5 Å threshold. Approaches using curated NOESY peak lists achieved the highest accuracy with a median accuracy of 1.05 Å and 80% of the entries below the threshold. The performance was essentially the same when un-curated NOESY peak lists were used (median accuracy 1.08 Å; 79% of entries below the threshold). In contrast, calculations based on either raw spectral data or CS only data performed less well, with median accuracies of 1.45 Å and 1.52 Å, respectively, both yielding only 50% of ensembles below the threshold.

The data collected within CASD-NMR-2013 allowed us to evaluate the dependence of the performance of automated structure generation methods on the input data, specifically comparing the use of curated NOESY peak lists relative to un-curated NOESY peak lists and/or raw spectral data. Only programs with multiple submissions using different types of input data for the same target were included in this analysis (cf. Table 2).

Firstly, we compared the use of curated and un-curated NOESY peak lists for methods that rely predominantly on NOESY data. ARIA submitted entries for five targets that allow for such a comparison. The median accuracy is 0.91 Å for the un-curated peak lists and 0.78 Å for the curated peak lists, suggesting that the use of curated peak lists does improve the accuracy. However, it should be noted that the accuracy of each entry is well within the 1.5 Å threshold for good quality ensembles regardless whether un-curated or curated peak lists were used. Similarly, for the ten qualifying entries (for five targets) submitted by CYANA, the median accuracy for entries generated from un-curated peak lists is slightly lower at 0.97

Å when compared to the values obtained for entries generated from the curated peak lists (0.84 Å), again with the accuracy for all entries comfortably within the threshold. Overall, ASDP performed slightly less well than ARIA or CYANA (but see below.) Based on entries for six targets, four ensembles generated by ASDP-Rosetta achieved the required accuracy using either un-curated or curated peak lists and the median accuracies were similar at 1.43 Å and 1.16 Å, respectively. Interestingly, for ASDP-CNS the proportion of entries within the accuracy cutoff rose from five out of eight generated from curated peak lists to six out of eight entries generated from un-curated peak lists, with median accuracies of 1.27 Å and 1.20 Å, respectively.

One deficiency in our analysis is the incomplete nature of the dataset. For example, ARIA and CYANA both submitted five paired entries, four of which were for the same targets, but of the six target pairs submitted by ASDP-Rosetta, only three pairs were also submitted by ARIA and two pairs were also submitted by CYANA (and for one pair RDC restraints are used by ASDP-Rosetta and not by CYANA.) As a result, any comparisons made across programs could lead to inappropriate conclusions. Indeed, ASDP-Rosetta is the only program to submit paired entries for both of the two most challenging targets (StT322 and HR8254A (Rosato et al., REF)), where ARIA and CYANA both failed to generate realistic converged structures from un-curated peak lists. Overall, our results would suggest that for algorithms relying primarily on NOESY data, ensembles of equivalent accuracy can be obtained regardless of whether curated or conservatively chosen un-curated peak lists are used as the input. It is worth noting that the two targets using the most liberal peak picking algorithm (i.e., StT322 and HR8254A,) and therefore including the largest fraction of probable noise peaks, proved the most difficult to solve using these fully automated analysis methods.

Cheshire-YAPP generates ensembles based on CS data then filters these ensembles based on NOESY distance restraints. Using un-curated peak lists, Cheshire-YAPP submitted entries for seven targets achieving a median accuracy of 1.24 Å with four entries within the 1.5 Å threshold. In this case, the use of curated peak lists significantly improved the accuracy of the entries as, out of the eight targets submitted, the median accuracy for the seven matched pairs improved to 1.05 Å and the accuracy of all eight entries was within the threshold. Only one program, UNIO, submitted entries based on both peak lists (Un-curated) and raw

spectral data. Entries were submitted for six targets with all of the ensembles generated based

on un-curated peak lists displaying an accuracy within the 1.5 Å RMSD threshold. Similarly, five of the six entries generated from raw spectral data achieved the desired accuracy, with the sixth entry (OR36) yielding a still acceptable accuracy of 1.6 Å. A small decline in the median accuracy (from 1.01 Å to 1.11 Å) was observed for the UNIO entries derived from raw spectral data compared to those calculated from un-curated peak lists.

Finally, we note that all but one of the entries misidentified the only *cis*-Proline in the target set, Pro142 in HR5460A as a *trans*-proline. *Cis*-prolines are normally identified using the chemical shift difference between  ${}^{13}C^{\beta}$  and  ${}^{13}C^{\gamma}$ , with 0 ppm  $\leq {}^{13}C^{\beta} - {}^{13}C^{\gamma} \leq 4.8$  ppm strongly indicative of a *trans* conformation and 9.15  $\leq {}^{13}C^{\beta} - {}^{13}C^{\gamma} \leq 14.4$  strongly indicative of a *cis* conformation (Schubert 2002) and/or by characteristic sequential H<sup>a</sup>/H<sup>a</sup> NOEs. The  ${}^{13}C^{\beta} - {}^{13}C^{\gamma}$  value for Pro142 in HR5460A is 7.88 ppm, hence in the transition region between these two chemical shift ranges. Only CS-HM-Rosetta successfully identified Pro142 as a *cis*-proline, but only in six out of ten conformers in the ensemble. This suggests that more rigorous determination of proline isomer state may be appropriate for all methods (including CS-HM-Rosetta.)

#### Geometric and packing quality

Structures can be validated by comparison of a set of metrics relative to those obtained from reference structures. We used the scores of the programs Molprobity (Lovell et al. 2003) and WHAT-IF (Vriend 1990), as implemented in the CING framework. Fig. 2 displays four such metrics, i.e. the fraction of backbone dihedrals in the Ramachandran disallowed region (Fig. 2A), the number of high energy interatomic contacts per 1000 atoms in the ensemble (Fig. 2B), the Ramachandran backbone angle distribution (Fig. 2C) and the side chain dihedral angle distribution (Fig. 2D) and, for both the targets and the entries. WHAT-IF values (Fig 2C,D) are given as the mean of the values calculated for each conformer in the ensemble. The scores reveal that the targets constitute well-refined structures, with near-zero percent of Ramachandran outliers (Fig. 2A) and Ramachandran Z-scores generally larger than -2 (Fig. 2C). Relatively few clashes are observed (Fig. 2B) and WHAT-IF side-chain Z-scores (Fig. 2D) of around zero are comparable to those observed in well-refined X-ray and NMR structures.

The values observed for the different entries vary considerably and correlate to some extent with the structure generation method, i.e. the engine used to generate the entry, with some targets displaying relatively better or worse scores across all programs. The effect of the generating method is most clearly observed from the WHAT-IF side chain dihedral scores (Fig. 2D). Structures from the Rosetta web server, CS-HM-Rosetta, Autonoe (also Rosetta-based) and ASDP-Rosetta, all display excellent median Z-scores. Conversely, CYANA, UNIO, Ponderosa, and Cheshire-YAPP have scores of around -5, whereas the other entries are intermediate between these two extremes. The excellent scores for the Rosetta-based protocols is no surprise, as its conformational sampling engine draws upon structural reference data similar to those used to generate the WHAT-IF side chain dihedral score. Low scores are indicative of non-optimal local geometry and do not imply errors in the overall fold.

The quality of the backbone geometry is expressed by the MolProbity fraction of Ramachandran outliers (Fig. 2A) and the WHAT-IF Ramachandran Z-Score (Fig. 2C). These two scores are complementary, as the MolProbity score reports the fraction of residues having nearly impossible dihedral angles while the WHAT-IF score reports the overall dihedral distribution. The entries generated by curated or un-curated Cheshire and UNIO display the poorest scores, with Cheshire also showing a large variability of the outliers scores within its submitted entries. ARIA also shows a substantial number of entries with larger outlier percentages, yet the WHAT-IF Ramachandran Z-scores are often better than those of the targets. The three Rosetta-based protocols (Autonoe, CS-HM-Rosetta and Web server Rosetta) and Ponderosa are generally good according to these two criteria, with nearly all entries displaying only small fractions of outliers and generally better Z-scores than the targets. Finally, the scores of ASDP-CNS and CYANA entries appear en-par with those of the targets.

Figure 2B shows the number of high-energy interatomic interactions, per 1000 residues, as determined by MolProbity. Here, the three Rosetta-based protocols, CYANA, and ASDP-Rosetta display the best values, below ten clashes per thousand atoms. ARIA, ASDP-CNS and Cheshire have values around twenty clashes per 1000 atoms, and UNIO, Ponderosa, and Cheshire-YAPP have median clash scores somewhat higher than the targets, with values of thirty or higher and extending up above forty. Close examination of the scores obtained for individual targets across the different entries reveals that entries for HR6470A and OR135 tend to be among the best scoring, whereas entries for StT322, HR2876C and OR36 are among the worst.

In addition to overall validation scores as provided by programs such as MolProbity and

WHAT-IF, it is also advantageous to examine residue-specific validation criteria. The CING program suite (Doreleijers et al. 2012a) implements such a residue-specific score as the socalled ROG-score. The ROG (red-orange-green) score represents a compounded measure of confidence for an individual entity, such as a residue, expressed as red for potentially problematic, orange for suspect and green for likely correct. The residue ROG score includes an assessment of the Ramachandran quality, the Omega dihedral and side chain dihedrals. The criteria are detailed in Table 2 of Doreleijers et al (Doreleijers et al. 2012a). ROG-scores reported in the current paper represent the fractions of the total number of residues in the well-defined range (as identified by CyRange,) with a specific red or green classification. As a rule of thumb, the number of green residues should exceed 50% while the number of red residues should be below 30%. Figs 3A and 3B display the green and red ROG-scores, respectively, for all entries and targets. In line with the overall scores, the data show that the targets display generally very good ROG scores, with all of the targets scoring better than the 50/30% criteria. For the entries the results are more diverse. As expected, and as observed before (Rosato et al. 2012), the CS-only based methods display very good ROG scores. Good scores were also obtained for many of the NOE/RDC based protocols, e.g. ARIA, ASDP-CNS, ASDP-Rosetta, Autonoe and most of the CYANA entries. In contrast, Cheshire using peak lists, Ponderosa and UNIO score substantially worse. StT322 appears to be a problematic target, which has the worst scores of all the reference structures and consistently scores poorly for the entries as well. Most automated methods had their poorest performance with the filtered peak list for StT322, and did not even provide a submission for this target with un-curated peak lists. For methods which did provide results using the un-curated or raw StT322 data, including ASDP-CNS, ASDP-Rosetta and Ponderosa, the resulting structures were clear outliers, with low accuracies. Potentially, this target has some special features, e.g. related to either the distribution of the chemical shifts, the quality of the data or the occurrence of conformational equilibria in solution, that distinguish it from the other targets.

#### Agreement with experimental data

The completeness of the experimental data and its agreement with the ensemble of conformers constitutes another class of useful metrics to assess the structural results. The quality of the structure produced by any given method is expected to depend on the amount of experimental data, i.e. to a large extent the number of correct NOEs that can be assigned and their information content. During the evaluation of the CASD-NMR-2010 round the

discriminating power (DP) score was used as a measure of the goodness-of-fit of the unassigned NOESY peak lists to the obtained structures. The DP score compares the unassigned NOESY peak lists to the generated structure, using the improvement in fit relative to a random coil reference structure to evaluate the structure quality. With possible values of 0 to 1, a DP lower score cutoff of ~0.7 has been considered to represent a reliable structure (Huang et al. 2012). For this round, the DP scores of the targets and the different entries are shown in Fig. 4A. As in the CASD-NMR-2010 results (Rosato et al. 2012,) a correlation was observed between DP score and the structural accuracy (Fig. S1). The DP scores of most of the entries are above 0.7, with the large majority in the 0.85-0.95 range, indicative of excellent agreement between experimental data and structural results. These CASD-NMR-2013 data further suggest a more practical cutoff for DP scores of reliable models should be above ~0.75 (RMSDs to reference < 3 Å.)

The expected number of NOEs that should be observed for a given structure ensemble presents a related measure correlating structural accuracy to experimental data. The so-called NOE completeness score can be calculated using the program Wattos, which is part of the CING suite. In practice, it is impossible to obtain all possible NOEs due to relaxation, peak overlap and alternating local conformations that can lead to conflicting assignments. In the NRG-CING database (Doreleijers et al. 2012b), the median NOE completeness is 44%, and this represents a realistic goal for modern structure determination by NMR. Fig. 4B shows the NOE completeness score for each of the targets and entries calculated from NOESY lists. For all the targets, the NOE completeness was well above the median database completeness consistent with the high quality of the target input data and the resulting structures. On average, the entries generated 64% (range: 31% - 87%) of the expected number of restraints. With the exception of the algorithms using methods that rely exclusively on raw spectral data, i.e. UNIO and Ponderosa, all other entries produced assigned NOE peak lists that yielded a completeness well above the database median. In comparison to the targets, three tools (ARIA, CYANA and I-TASSER) performed better than the expert in assigning NOE peaks.

This high level of NOE completeness, combined with the high level of accuracy and the generally good quality of the structures generated, led us to hypothesize that there would be significant overlap between the restraints identified and used by the expert researcher and any one of the automated protocols as implemented in the different programs. To investigate this, we started from the list of restraints generated manually by the expert researcher or by each of the programs for each target. We removed the differences between the expert's and the

algorithms' treatment of stereochemical assignment by treating all restraints as pseudo-atom restraints. This list was then curated to include only long-range restraints, i.e. those between atoms at least five residues apart, since these are the restraints that are known to carry the majority of the structural information (Nabuurs et al. 2003). The results of this analysis are shown in Fig. 5A. To our surprise they revealed that none of the automated methods identified more than 50% of the restraints produced manually and some even obtained less than 10%. As an example, Fig. 5C shows the restraint overlap between target OR36 and all entries (the majority of which were within 1.5 Å from the target structure). The aforementioned low degree of overlap between the manually derived, i.e. target, restraints is evident from the top row of this graph. The overlap between entries derived by the different programs is also highly variable, albeit that within a single program group the values are, as expected, consistently much higher.

Following this somewhat unexpected result, we decided to look at the overlap on a residueto-residue basis rather than atom-to-atom, as it could be expected that the proximity of two residues rather than the exact NOE is the determining factor. Starting from the same curated pseudo-atom restraint lists generated above, we then generated a list containing only residueto-residue restraint information. The results shown in Fig. 5B were much more consistent with our hypothesis, as for most of the targets the restraints obtained by the automated algorithms were now overlapping by more than 80% with the manually derived restraints. A similar increase was obtained for the restraint overlap between the entries generated by the different programs, as illustrated for the OR36 target (Fig. 5D).

Given the importance of NOE data in producing an accurate structure for many of the programs used in the CASD-NMR-2013 effort, we wanted to explore a possible correlation between the accuracy of the results, as expressed by RMSD (cf. Fig. 1) to the target, and the NOE-derived restraint overlap (Fig. 6). Although an extremely weak correlation can be inferred for some targets and methods (data not shown) no overall correlation patterns are supported by the data, neither on a pseudo-atom basis nor on a per-residue basis. Some clustering based on program group is also observed for some targets, however the clustering observed is not consistent across targets in either the pseudo-atom or per-residue based plots. We speculate that this unexpected lack of correlation can be explained by the information redundancy present in NOEs between atoms contained within a fixed covalent network.

It is clear from the Figures in this paper that both RMSD values and validation scores are correlated for individual targets across different calculations; some targets tend to be either among the best or among the worst for all calculation protocols. The observation is semiquantitative at best, given the variability of results across programs and program types, and the fact that not all targets were attempted (or resulted in converged structures) for all programs. It was not possible to determine any systematic variation with structure type: CASD-2013 included six  $\alpha/\beta$  proteins and three all- $\alpha$  proteins, and both groups contained both 'good' and 'bad' targets. Two of targets stood out for unrelated reasons, as discussed in the accompanying paper (Rosato et al. REF). Producing converged entries from the uncurated peak lists for HR8254A and StT322 proved difficult, and entries for these targets were missing for a number of programs. These targets were both small (ca. 40 defined residues), were the only targets to use non-uniformly-sampled NMR data, had no RDC data, and had a high proportion of probable noise peaks in their un-curated peak lists. HR8254A gave consistently good validation scores, but had among the highest RMSD values for both accuracy and convergence. HR8254A is a three-helix protein with one very long helix extending outside a small core; clearly RMSD calculations will be quite sensitive to small variations in the inter-helical angle that, in the absence of RDC data, is difficult to determine precisely. StT322 is the only all-β protein in the set, and has a particularly large ill-defined tail. It gave consistently poor validation scores, and also gave high RMSD values for both convergence and accuracy.

#### Discussion

The above results provide for a comprehensive evaluation of the performance of the currently available programs for automated protein structure generation from NMR data. The reference structures for all of the ten CASD-NMR-2013 targets are well converged (ensemble convergence 0.4 Å to 1.0 Å). The quality of these structures is higher than in the CASD-NMR-2010 round (ensemble convergence 0.4 Å to 1.7 Å) suggesting that either the quality of the input data has improved, most likely as a result of improved NMR hardware and acquisition schemes, or that there have been significant improvements in manual data analysis and structure calculation routines. Most likely, it resulted from a combination of both these factors.

Overall, the performance of the automated structure determination methods, in terms of accuracy and ensemble convergence, was excellent (median ensemble convergence 0.6 Å; median accuracy 1.14 Å). An accuracy threshold of 1.5 Å was imposed in this work to

identify acceptable structures, which was achieved by 71% of the entries. The less stringent 2.5 Å threshold imposed previously (Rosato et al. 2012) was achieved by 72% of the entries in CASD-NMR-2010. Applying the same more relaxed criterion to the present CASD-NMR-2013 effort raises the success rate to 85% of the entries. This improvement relative to CASD-NMR-2010 may partly be attributed to the quality of the input data as discussed above, but more likely also to advances in the structure generation engines, as also the overall quality indicators have improved. The success rate is even higher (78% using the 1.5 Å cutoff) if the most challenging targets, HR8254A and StT322, are excluded. As for CASD-NMR-2010, the performance of the programs depended on the nature of the input data and once again the best results were obtained by methods utilizing NOESY data, either as the primary input or to augment CS data.

The comparison between the entries obtained from either the use of curated NOESY peak lists *versus* un-curated peak lists revealed that for programs relying on NOESY data as the primary input the use of curated peak lists does not lead to significantly better structures. It appears that the iterative procedures implemented in these protocols are efficient at filtering the peaks for consistent information. For example, CYANA performs the so-called network-anchoring and restraint-combination methods to perform such peak filtering (Herrmann et al. 2002). However, programs such as Cheshire-YAPP that use NOESY data to augment the input CS data performed significantly better when using curated peak lists. The data also suggest that UNIO performs better with an un-curated peak list than with raw spectral data. The use of curated peak lists may however improve convergence. It is interesting to note that five out of ten ARIA calculations and three out of eight CYANA calculations with un-curated peaks failed to converge, where the equivalent calculations using curated peaks led to good quality structures. The non-converging seemed to correlate with the proportion of extraneous peaks in the un-curated peak lists (data not shown).

As for CASD-NMR-2010, three validation parameters were used to assess the geometric and packing quality of the CASD-NMR-2013 submissions: Ramachandran backbone angle distribution, side-chain angle distribution and the number of high-energy interatomic contacts (Fig. 2). As in the previous CASD-NMR-2010 round, these parameters varied over a wide range of Z-score (up to 15 standard deviations,) and were overwhelmingly determined by the choice of structure calculation protocol. The three Rosetta-based protocols (Autonoe, CS-HM-Rosetta, and Web server Rosetta) and also the Rosetta-refined ASDP-Rosetta all did extremely well based on geometric criteria. This result is not surprising, given that these

programs derive their backbone conformations and refinement parameters from databases of known good geometries. ARIA performed moderately well across all criteria, whereas ASDP-CNS, CYANA, Cheshire, and Ponderosa achieved acceptable but more mixed validation scores. Nevertheless, the relatively good validation scores for ARIA and ASDP-CNS confirm that water refinement in a realistic force field has a very positive effect on the geometry of the final conformer, also when performed in automation. Finally, UNIO and Cheshire-YAPP consistently received the lowest scores across all criteria, which is likely due to the lack of the aforementioned refinement procedures. Our investigations did not reveal any promising correlations between any of the geometric parameters and either accuracy or convergence.

Residue-specific ROG scores are indicators of local conformation and sensitive to errors in restraints. Within the set of entries, Rosetta/chemical-shift based methods show very good ROG scores (Fig. 3). Again, this is an expected result as some of the parameters that underpin the ROG score are also based upon comparison with fragments from structures contained within the PDB database. The results for Cheshire however, display an interesting phenomenon; whereas the chemical-shift-only entries display the expected good ROG scores, inclusion of the NOE peak lists does improve accuracy, albeit at the expense of much poorer ROG scores. It has been shown that for NOE/RDC-based structures consistently poor local conformation, as expressed by poor ROG scores, correlates with propensity for errors in the overall fold. It is notable that the entries for the StT322 target generally display among the worst ROG scores combined with the lowest accuracy scores (Fig. 1A). Generally however, the accuracy of the entries is high, suggesting that other factors may also depress the ROG scores. Proper refinement in a force-field that implements an explicit water-shell has been shown to substantially improve local conformation as well as the agreement with experimental restrains (Linge et al. 2003b; Spronk et al. 2002). Whereas some protocols, e.g. ARIA, ASDP and Autonoe, do implement such a refinement step as a standard procedure, for others like CYANA and UNIO this is generally not the case. As the accuracy of the latter protocols is similar to the accuracy of the former, it is likely that the observed differences in their ROG score patterns could be the result of the (lack of) final refinement, rather than of significant differences in the interpretation of the underlying data.

For the CASD-NMR-2013 entries we investigated the quality of the NOE input data and the accuracy of the structure generated by the different methods. For both the reference structures

and the CASD-NMR-2013 entries the NOE completeness scores (Fig. 4B) were well-above the median in the CING database and the DP-scores (Fig. 4A) largely exceeded the lower cutoff of 0.7. Together, this indicates that all of the automated methods perform well with regard to assigning NOE restraints. We also determined the extent of NOE restraint overlap between different entries using either pseudo-atoms or residues as the basis for comparison (Fig. 5). When using pseudo-atoms as the basis for comparison, the restraint overlap was surprisingly low between the reference structure and the CASD-NMR-2013 entries but the degree of overlap increased significantly when determined at the residue level. This discrepancy between the extent of restraint overlap observed when using pseudo-atom to pseudo-atom restraints compared to residue to residue restraints could be explained by experimenter bias when manually generating restraint lists. For example, a human researcher, having identified NOE patterns consistent with an alpha helix, may invest more time and energy identifying all of the NOEs for the helix than the unbiased automated methods. Similarly, the human researcher may devote more attention to assigning as many NOE's as possible in an under-restrained portion of an intermediate structure while the automated methods would be expected to spend equivalent amounts of effort on each region in the molecule. In support of this notion, we observed that at the pseudo-atom level the overlap between any two of the entries was higher when compared to the overlap between the target restraint list and any one of the entries. This is exemplified by the OR36 target (Fig. 5C,D). The results obtained for the pseudo-atom basis (Fig. 5C) show that, in general, overlap is greater between automated methods than between an automated method and the manual assignment. However, the overlap does not reach 100% in either case, suggesting that differences cannot be entirely due to experimenter bias alone. It has been previously observed that multiple calculations starting from the same data may result in different restraint sets, with only a subset of restraints common to all calculations (Buchner and Güntert 2015). In contrast, there is little difference at the residue level in the extent of overlap between two automated algorithms or between an automated algorithm and the expert researcher (Fig. 5D). Notably, the methods based on raw spectral data have a lower overlap with all other methods, also on the per-residue basis. Given that they generated accurate structures for the majority of targets, this suggests that not all long-range contacts are equally important to define the correct protein fold. Finally, we found that there was no significant correlation between the extent of NOE restraint overlap and the accuracy of the structure. This result was surprising given the importance of NOE data in producing good quality structures. We suggest that this could be explained by the information redundancy present in NOEs between atoms contained within a fixed covalent network.

#### Conclusions

The ten targets of CASD-NMR-2013 constitute a high-quality set of NMR structures, exemplifying the quality that can be attained by a skilled researcher using state-of-the-art techniques. Overall, the results from CASD-NMR-2013 demonstrate that automated structure determination protocols are capable of reliably producing structures of comparable accuracy and quality, at least for small, single domain proteins such as the ten targets tested. The most robust results appear to be obtained when NOESY peak lists are used either as the primary input data or to augment CS data, with limited need to manually refine such lists. Since no single method performed consistently better than the others for all ten targets it is advisable to use more than one program routinely and combine the results.

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Table 1. CASD-2013 targets. The PDB ID, valid ranges and reference ensemble sources for comparison of each target is given.

TARGET ID	PDB ID	Valid Range(s)	Reference Ensemble Authors
HR2876B	2LTM	13-105	Liu, G., Xiao, R., Janjua, H., Hamilton, K., Shastry, R., Kohan, E., Acton, T.B., Everett, J.K., Lee, H., Huang, Y.J., Montelione, G.T.
HR2876C	2M5O	17-91	Liu, G., Xiao, R., Janjua, H., Hamilton, K., Shastry, R., Kohan, E., Acton, T.B., Everett, J.K., Pederson, K., Huang, Y.J., Montelione, G.T.
HR5460A	2LAH	14-25, 33-158	Liu, G., Shastry, R., Ciccosanti, C., Hamilton, K., Acton, T.B., Xiao, R., Everett, J.K., Montelione, G.T.
HR6430A	2LA6	14-99	Liu, G., Xiao, R., Janjua, H., Lee, H., Ciccosanti, C.T., Acton, T.B., Everett, J.K., Huang, Y.J., Montelione, G.T.
HR6470A	2L9R	554-608	Liu, G., Xiao, R., Lee, HW., Hamilton, K., Ciccosanti, C., Wang, H.B., Acton, T.B., Everett, J.K., Huang, Y.J., Montelione, G.T.
HR8254A	2M2E	15-56	Lemak, A., Yee, A., Houliston, S., Garcia, M., Ong, M., Arrowsmith, C.
OR135	2LN3	4-74	Liu, G., Koga, R., Koga, N., Xiao, R., Lee, H., Janjua, H., Kohan, E., Acton, T.B., Everett, J.K., Baker, D., Montelione, G.T.
OR36	2LCI	2-46, 53-125	Liu, G., Koga, N., Koga, R., Xiao, R., Lee, H.T., Janjua, H., Ciccosanti, C., Acton, T.B., Everett, J., Baker, D., Montelione, G.T.
StT322	2LOJ	23-63	Wu, B., Yee, A., Houliston, S., Garcia, M., Savchenko, A., Arrowsmith, C.H.
YR313A	2LTL	17-41, 45-115	Liu, G., Xiao, R., Hamilton, K., Janjua, H., Shastry, R., Kohan, E., Acton, T.B., Everett, J.K., Lee, H., Huang, Y.J., Montelione, G.T.

Table 2. Median accuracy of paired entries. Only targets calculated using both curated and un-curated data (or un-curated and raw data, in the case of UNIO) are included. Note that no program submitted paired entries for all targets and therefore comparison of accuracies made across programs is potentially inappropriate (see text.)

Program	Curated (Å)	Un-curated (Å)	Raw (Å)	Number of targets
ARIA	0.78	0.91	-	5
ASDP-CNS	1.27	1.20	-	8
ASDP-ROSETTA	1.16	1.43	-	6
CHESHIRE-YAPP	1.05	1.24	-	7
CYANA	0.84	0.97	-	5
UNIO	-	1.01	1.11	6

#### **Figure captions**

#### Figure 1.

Comparison of targets and entries. (A) Structural similarity (accuracy): the mean all *vs* all pairwise backbone RMSD for well-defined residues for each of the entries with respect to the target. The dashed line at 1.5 Å indicates a reasonable upper threshold for identity within experimental uncertainty (see text for details). (B) The pairwise backbone RMSD for well-defined residues within each ensemble for each of the targets and entries. The dashed line at 1.0 Å indicates an estimated upper threshold for a converged structure. Symbols for each target are indicated on the left. Open symbols indicate entries generated from truncated input sequences. Horizontal axis labels: targets are labeled in green, entries generated from curated lists in black, curated lists plus RDCs in bold-black, un-curated lists in blue, un-curated lists plus RDCs in bold-blue, CS only in magenta, CS plus RDCs in bold magenta and raw data in purple.

#### Figure 2.

Overall quality scores of the targets and the entries. (A) Molprobity Ramachandran outliers (Lovell et al. 2003). (B) Molprobity number of clashes per thousand atoms in the ensemble. (C) WHAT-IF Ramachandran Z-scores (Vriend 1990). (D) WHAT-IF side chain Z-scores. Symbols and labels are explained in the legend for Figure 1.

#### Figure 3.

ROG scores (Doreleijers et al. 2012a) of the targets and the entries. (A) The fraction of residues with a green ROG score. The lower threshold of 0.5 is indicated by a dashed line. (B) The fraction of residues with a red ROG score. The upper threshold of 0.3 is indicated by a dashed line. Symbols and labels are explained in the legend for Figure 1.

#### Figure 4.

Agreement with experimental data of the targets and the entries. (A) The DP score (Huang et al. 2005). The dashed line indicates the lower threshold of 0.75 for agreement between the structure and the input data. (B) The NOE completeness determined by Wattos. The dashed line indicates the median NOE completeness (44.2%) for all structures in the NRG-CING database (Doreleijers et al. 2012b). Symbols and labels are explained in the legend for Figure 1. Only entries calculated from NOESY lists have been included.

#### Figure 5.

Overlap of long-range NOE restraints between the targets and the entries. Fraction of overlapping NOE restraints between the target and each entry determined on the basis of (A) pseudo-atom or (B) residue. Symbols and labels are explained in the legend for Figure 1. (C,D) Heatmaps of the fractions of overlapping long-range NOE restraints between the OR36 target and entries, determined on the basis of pseudo-atom (C) or residue (D). The total number of long-range restraints present for each target/entry is shown on the diagonal. The off-diagonal values denote the percentage of restraints used in the entry indicated along the row that are also found in the entry indicated along the column. The top row shows the percentage of NOE's used in the reference structure that were found in each entry, while the left-most column shows the percentage of NOE's used by each entry that were found in the reference structure. For example, the entry in the square marked by the black box in (C) shows that 238 restraints (22%) used in the OR36 target are also present in the OR36\_ASDP-CNS\_c entry.

#### Figure 6.

Correlation between entry pairwise RMSD and NOE restraint overlap. For every pair of entries for a given target, the all-by-all RMSD and NOE restraint overlap between those entries is shown. NOE restraint overlap are calculated on a (A) pseudo-atom or (B) residue basis. Symbols are explained in the legend for Figure 1.









### Analysis of the Structural Quality of the CASD-NMR 2013 Entries

### **Supplementary materials**

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Supplementary Figure 1. DP score *vs* Accuracy. The DP score *vs* mean all by all pairwise backbone RMSD to the reference structure is plotted for all valid entries. The dashed line indicates the more stringent lower threshold of 0.75 for identifying reliable (RMSDs to reference <  $\sim$ 3 Å) models. Symbols for each target are indicated on the right. Open symbols indicate entries generated from truncated input sequences.

						At least	5 residue	es apart.					
HR2876B	1123	44%	44%	43%	44%	42%	37%	42%	37%	49%	30%	27%	40% -
HR2876B_ARIA_c	63%	1038	96%	91%	91%	88%	79%	88%	79%	95%	69%	60%	83% -
HR2876B_ARIA_cr	63%	97%	1046	91%	91%	87%	80%	88%	80%	95%	70%	60%	83%
HR2876B_ARIA_u	- 30%	44%	44%	2458	90%	42%	47%	42%	47%	46%	36%	30%	58% -
HR2876B_ARIA_ur	- 31%	46%	46%	93%	2389	44%	49%	44%	49%	48%	38%	32%	60% -
HR2876B_ASDP-CNS_c	- 66%	86%	85%	86%	87%	725	80%	94%	80%	91%	64%	55%	80% -
HR2876B_ASDP-CNS_ur	- 53%	70%	69%	89%	90%	73%	795	73%	100%	74%	58%	49%	80% -
HR2876B_ASDP-Rosetta_c	65%	86%	86%	86%	88%	94%	81%	721	81%	92%	64%	55%	81% -
HR2876B_ASDP-Rosetta_ur	- 53%	70%	69%	89%	90%	73%	100%	73%	795	74%	58%	49%	80% -
HR2876B_CYANA_c	63%	89%	88%	89%	88%	83%	76%	84%	76%	1063	65%	55%	80% -
HR2876B_Ponderosa_r	- 56%	83%	82%	91%	<b>92%</b>	82%	81%	82%	81%	87%	715	61%	85%
HR2876B_UNIO_r	66%	90%	89%	95%	96%	90%	89%	90%	89%	93%	76%	579	95%
HR2876B_UNIO_u	- 48%	66%	66%	95%	95%	65%	70%	65%	70%	70%	55%	49%	1168 -
	HR2876b	HR2876b	HR2876b-	HR28760-	HR28T6D	HR2876b	HR2876b	HR2876b-	HR28765	HR2876b-	HR2876b-	HR2876b	HR2876B-
		ζ.	AAIAS	ARIA_CI	ARIA	ARIA	ASDP-CNS	ASDP-CNS	ASDP-Roset	ASDP-Rose	CYANAC	ponderosa	UNIO
							<u>`</u> ە	È.	hac	itta ur		5	

Supplementary Figure 2A. Heatmap of the fractions of overlapping long-range NOE restraints between the HR2876B target and entries, determined on the basis of pseudo-atom.



Supplementary Figure 2B. Heatmap of the fractions of overlapping long-range NOE restraints between the HR2876B target and entries, determined on the basis of residue.

						At least	5 residue	es apart.					
HR2876C	907	26%	26%	31%	30%	20%	40%	31%	38%	33%	14%	17%	26% -
HR2876C_ARIA_cr	- 53%	555	100%	81%	75%	55%	92%	78%	94%	83%	34%	56%	73% -
HR2876C_ARIA_cr(10-97)	- 53%	100%	555	81%	75%	55%	92%	78%	94%	83%	34%	56%	73%
HR2876C_ARIA_ur	- 41%	56%	56%	879	55%	47%	73%	78%	80%	76%	32%	40%	73% -
HR2876C_ASDP-Rosetta_cr	- 57%	64%	64%	73%	474	59%	85%	76%	90%	81%	31%	43%	65% -
HR2876C_ASDP-Rosetta_ur	- 43%	51%	51%	69%	64%	435	64%	70%	77%	71%	26%	37%	61% -
HR2876C_CYANA_c	- 50%	63%	63%	74%	65%	45%	866	73%	88%	78%	29%	43%	64% -
HR2876C_CYANA_u	- 40%	53%	53%	80%	59%	48%	74%	835	81%	77%	31%	39%	71% -
HR2876C_I-TASSER_c	- 28%	37%	37%	47%	39%	30%	51%	46%	1521	71%	19%	25%	40% -
HR2876C_I-TASSER_u	- 22%	28%	28%	38%	31%	24%	39%	38%	62%	1699	16%	20%	32% -
HR2876C_Ponderosa_r	- 47%	52%	52%	73%	57%	44%	67%	73%	81%	81%	317	43%	67% -
HR2876C_UNIO_r	52%	78%	78%	87%	75%	59%	90%	83%	94%	87%	41%	345	85% -
HR2876C_UNIO_u	- 42%	60%	60%	88%	62%	53%	77%	84%	85%	80%	35%	48%	648 -
	HR28To	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876	HR2876
	C	.* C \.	, ARIA CI	, ARIA crt	, ARIA UT	, ASDP-AG	, ASDP-R	CYANA	, CHANA	, LTASSE	, LTASSE	, ponderos	UNIO-
				(10-9T)		osetta_cr	osetta ut	<u>`</u> م	'L	AC	AL	58-5	

Supplementary Figure 3A. Heatmap of the fractions of overlapping long-range NOE restraints between the HR2876C target and entries, determined on the basis of pseudo-atom.

						At least	5 residue	es apart.					
HR2876C	- 778	96%	96%	98%	93%	83%	99%	97%	98%	97%	65%	85%	93%
HR2876C_ARIA_cr	98%	555	100%	99%	95%	86%	97%	98%	99%	97%	69%	88%	95% -
HR2876C_ARIA_cr(10-97)	98%	100%	555	99%	95%	86%	97%	98%	99%	97%	69%	88%	95%
HR2876C_ARIA_ur	92%	90%	90%	878	86%	81%	92%	94%	95%	92%	64%	82%	90%
HR2876C_ASDP-Rosetta_cr	95%	93%	93%	97%	172	85%	95%	97%	96%	97%	71%	78%	90%
HR2876C_ASDP-Rosetta_ur	84%	81%	81%	89%	84%	172	84%	88%	90%	92%	63%	70%	81% -
HR2876C_CYANA_c	98%	95%	95%	98%	92%	84%	816	97%	97%	96%	64%	84%	93%
HR2876C_CYANA_u	94%	92%	92%	98%	90%	85%	95%	803	96%	96%	64%	83%	93%
HR2876C_I-TASSER_c	80%	77%	77%	86%	74%	68%	80%	83%	693	88%	49%	63%	73% -
HR2876C_I-TASSER_u	68%	66%	66%	73%	64%	60%	69%	71%	77%	751	45%	54%	63% -
HR2876C_Ponderosa_r	97%	93%	93%	97%	92%	91%	96%	96%	97%	99%	303	88%	96% -
HR2876C_UNIO_r	99%	98%	98%	100%	96%	89%	100%	99%	100%	98%	75%	335	98% -
HR2876C_UNIO_u	95%	94%	94%	99%	92%	85%	96%	97%	98%	97%	68%	88%	509 -
	HR2876	HR2876	HR2876	HR2876	HR2876	HR28T	HR2876	HR28To	HR2876	HR2876	HR2876	HR2876	HR28764
	c	, c	, ARIA CI	, ARIA CIL	ARIA II	, ASOP. AO	ASDP-Ro	, CHANA-0	, CYANA-V	LTASSEP	, LTASSEP	, ponderosi	UNIO-1
				(Te-0)		setta_cr	setta	0	5	10	14	10,00	

Supplementary Figure 3B. Heatmap of the fractions of overlapping long-range NOE restraints between the HR2876C target and entries, determined on the basis of residue.



Supplementary Figure 4A. Heatmap of the fractions of overlapping long-range NOE restraints between the HR5460A target and entries, determined on the basis of pseudo-atom.



Supplementary Figure 4B. Heatmap of the fractions of overlapping long-range NOE restraints between the HR5460A target and entries, determined on the basis of residue.



Supplementary Figure 5A. Heatmap of the fractions of overlapping long-range NOE restraints between the HR6430A target and entries, determined on the basis of pseudo-atom.



Supplementary Figure 5B. Heatmap of the fractions of overlapping long-range NOE restraints between the HR6430A target and entries, determined on the basis of residue.

At least 5 residues apart. HR6470A 313 20% 20% 20% 20% 18% 16% 18% 14% 17% 16% 16% 14% 20% 18% 10% 15% -																	
HR6470A -	313	20%	20%	20%	20%	18%	16%	18%	14%	17%	16%	16%	14%	20%	18%	10%	15% -
HR6470A_ARIA_c = :	25%	375	99%	98%	98%	90%	84%	89%	79%	77%	75%	71%	69%	91%	81%	56%	74% -
HR6470A_ARIA_c(10-59) - ;	24%	96%	385	95%	97%	87%	82%	86%	78%	74%	73%	68%	66%	87%	78%	54%	71%
HR6470A_ARIA_cr -	25%	98%	98%	375	98%	89%	84%	89%	78%	76%	75%	70%	68%	89%	81%	55%	73% -
HR6470A_ARIA_cr(10-59) - ;	23%	96%	98%	96%	385	87%	83%	86%	77%	75%	74%	68%	67%	87%	78%	54%	71% -
HR6470A_ARIA_u - ;	23%	87%	87%	87%	87%	355	87%	97%	82%	69%	67%	71%	68%	81%	81%	52%	75%
HR6470A_ARIA_u() - ;	20%	86%	87%	86%	87%	91%	341	91%	86%	69%	68%	69%	68%	80%	79%	52%	72% -
HR6470A_ARIA_ur - ;	23%	86%	85%	87%	85%	97%	86%	360	81%	68%	67%	71%	68%	81%	81%	51%	74%
HR6470A_ARIA_ur()	17%	81%	83%	81%	82%	87%	88%	87%	334	65%	64%	67%	67%	76%	76%	49%	70% -
HR6470A_ASDP-CNS_c -	34%	93%	92%	93%	93%	85%	81%	85%	73%	156	95%	81%	80%	95%	82%	55%	74%
HR6470A_ASDP-CNS_cr -	33%	92%	93%	92%	93%	84%	80%	84%	72%	95%	155	79%	81%	95%	81%	55%	75% -
HR6470A_ASDP-CNS_u -	34%	90%	90%	90%	90%	92%	86%	92%	82%	88%	85%	144	87%	91%	90%	51%	81%
HR6470A_ASDP-CNS_ur - 3	31%	89%	89%	89%	90%	92%	86%	92%	84%	89%	90%	89%	140	92%	89%	54%	81%
HR6470A_CYANA_c - ;	27%	95%	95%	95%	95%	86%	82%	86%	75%	82%	82%	73%	74%	280	83%	55%	75% -
HR6470A_CYANA_u -	29%	97%	97%	97%	97%	99%	94%	99%	87%	82%	81%	83%	80%	95%	235	60%	87%
HR6470A_UNIO_r - :	28%	99%	99%	99%	99%	96%	93%	96%	83%	87%	87%	76%	79%	99%	92%	135	91%
HR6470A_UNIO_u - :	27%	96%	97%	96%	97%	100%	94%	100%	87%	84%	86%	86%	85%	96%	97%	67%	203 -
	HREATOA	HR64TOA.	HRBATOA.	HPIGATOR.	HR64TOA.	HRIGATOR.	HR64TOA-	HREATOA	HR6ATOA.	HR6ATOA.	HABATOA	HREATOR	HREATOA	HREATOR	HR64TOA.	HABATOA	HREATOA
		ź,	AAIA-C	ARIA-C(10-5	AllAct	ARIA-CI(10-	ARIA	ARIA-UI	ARIA	ARIA_UT()	ASOP-CNS	SDP-CNS	NSDP-CNS	ASDP-CNS	CHANA-C	CHANALU	UNIO-5

Supplementary Figure 6A. Heatmap of the fractions of overlapping long-range NOE restraints between the HR6470A target and entries, determined on the basis of pseudo-atom.

At least 5 residues apart.																	
HR6470A	282	100%	100%	100%	100%	96%	93%	96%	93%	96%	96%	90%	93%	100%	93%	90%	91%
HR6470A_ARIA_c	97%	375	100%	100%	99%	96%	91%	94%	92%	91%	90%	86%	89%	97%	89%	85%	87%
HR6470A_ARIA_c(10-59)	95%	99%	384	99%	99%	95%	90%	93%	90%	90%	89%	85%	86%	95%	87%	83%	84%
HR6470A_ARIA_cr	95%	99%	98%	375	98%	96%	91%	94%	91%	90%	90%	86%	88%	95%	87%	84%	86%
HR6470A_ARIA_cr(10-59)	96%	99%	99%	99%	385	95%	91%	93%	90%	91%	90%	86%	88%	96%	88%	84%	85% -
HR6470A_ARIA_u	89%	92%	92%	93%	92%	355	91%	99%	91%	86%	85%	86%	87%	89%	87%	81%	86%
HR6470A_ARIA_u()	88%	91%	91%	92%	91%	95%	341	96%	97%	87%	86%	86%	87%	88%	87%	82%	84%
HR6470A_ARIA_ur	87%	90%	90%	93%	90%	99%	89%	360	89%	84%	83%	86%	86%	88%	86%	79%	84% -
HR6470A_ARIA_ur()	88%	90%	91%	90%	90%	93%	94%	93%	334	87%	84%	84%	84%	88%	86%	80%	83% -
HR6470A_ASDP-CNS_c	100%	100%	100%	100%	100%	93%	91%	93%	91%	43	98%	88%	93%	100%	88%	84%	86%
HR6470A_ASDP-CNS_cr	100%	100%	100%	100%	100%	93%	90%	93%	90%	100%	41	88%	95%	100%	88%	85%	88%
HR6470A_ASDP-CNS_u	93%	98%	98%	98%	98%	98%	90%	98%	90%	90%	88%	42	93%	95%	93%	79%	86%
HR6470A_ASDP-CNS_ur	95%	98%	98%	98%	98%	98%	93%	98%	93%	95%	95%	93%	43	95%	88%	84%	91%
HR6470A_CYANA_c	100%	100%	100%	100%	100%	95%	93%	95%	93%	96%	95%	90%	92%	265	92%	89%	91%
HR6470A_CYANA_u	100%	100%	100%	100%	100%	100%	98%	100%	97%	98%	97%	97%	97%	100%	231	93%	96%
HR6470A_UNIO_r	99%	100%	100%	100%	100%	97%	96%	97%	94%	99%	99%	90%	96%	100%	93%	134	96%
HR6470A_UNIO_u	98%	98%	98%	98%	98%	100%	97%	100%	96%	98%	98%	97%	99%	98%	97%	93%	199 -
	HRBATOA	HABATORS	HABATOA	HREATOR	HAGATOR	HREATOR	HREATON	HRBATOA	HREATON	HRBATOA	HAGATOA	HREATON	HREATON	HAGATOR	HREATON	HABATOR	HRIEATOAL
		4	AIAC	AIA-0(10-59	AIACI	AIA_01(10-55	AIAU	AIA_UO	ALA UT	AIA_UI()	SOP-CNS-C	SDP.CN5.C	SOP-CNS-U	SDP-CNS-V	AMAC	NAMAL	10 Juno Ju

Supplementary Figure 6B. Heatmap of the fractions of overlapping long-range NOE restraints between the HR6470A target and entries, determined on the basis of residue.



Supplementary Figure 7A. Heatmap of the fractions of overlapping long-range NOE restraints between the YR313A target and entries, determined on the basis of pseudo-atom.



Supplementary Figure 7B. Heatmap of the fractions of overlapping long-range NOE restraints between the YR313A target and entries, determined on the basis of residue.



Supplementary Figure 8A. Heatmap of the fractions of overlapping long-range NOE restraints between the HR8254A target and entries, determined on the basis of pseudo-atom.



Supplementary Figure 8B. Heatmap of the fractions of overlapping long-range NOE restraints between the HR8254A target and entries, determined on the basis of residue.



Supplementary Figure 9A. Heatmap of the fractions of overlapping long-range NOE restraints between the StT322 target and entries, determined on the basis of pseudoatom.



Supplementary Figure 9B. Heatmap of the fractions of overlapping long-range NOE restraints between the StT322 target and entries, determined on the basis of residue.

At least 5 residues apart.													
OR36	- 1080	31%	32%	22%	22%	16%	17%	30%	24%	7%	9%	22% -	
OR36_ARIA_c	- 34%	1309	97%	71%	72%	54%	55%	89%	71%	23%	29%	62% -	
OR36_ARIA_cr	- 34%	94%	1366	69%	71%	54%	55%	89%	71%	23%	27%	62% -	
OR36_ASDP-CNS_c	- 37%	87%	87%	639	80%	55%	56%	84%	67%	22%	27%	59% -	
OR36_ASDP-CNS_cr	- 39%	88%	89%	82%	626	57%	60%	87%	71%	24%	28%	61% -	
OR36_ASDP-CNS_u	- 35%	79%	81%	71%	72%	496	83%	78%	79%	25%	31%	70% -	
OR36_ASDP-CNS_ur	- 34%	77%	80%	68%	72%	79%	522	77%	77%	23%	32%	67% -	
OR36_CYANA_c	- 33%	91%	93%	65%	67%	49%	50%	1052	66%	21%	25%	55% -	
OR36_CYANA_u	- 25%	62%	64%	47%	50%	44%	45%	60%	1177	21%	22%	66% -	
OR36_Ponderosa_r	- 27%	66%	69%	57%	62%	50%	48%	67%	70%	336	34%	65% -	
OR36_UNIO_r	- 42%	95%	94%	83%	83%	75%	80%	91%	91%	41%	253	86%	
OR36_UNIO_u	- 30%	68%	71%	55%	56%	52%	52%	66%	85%	25%	28%	844 -	
	OR36	OR36-M	OR36-M	A-96HO	OR36_A	OH36-M	OH36-A	0436-0	0436-0	OR36_P	0R36_U	OR36-U	
			ANA-C	SA SA	CUB-CNS	ONP.CNS.	-OP-CNS-	-OP-CNS	ANAC	MNA-1	Merosa	MOL	
					`O	,õ	14	` <b>E</b>					

Supplementary Figure 10A. Heatmap of the fractions of overlapping long-range NOE restraints between the OR36 target and entries, determined on the basis of pseudo-atom.



Supplementary Figure 10B. Heatmap of the fractions of overlapping long-range NOE restraints between the OR36 target and entries, determined on the basis of residue.

						At I	east 5 re	sidues ap	art.					
OR135	945	41%	41%	31%	31%	36%	20%	36%	20%	45%	30%	9%	20%	23% -
OR135_ARIA_c	- 58%	888	96%	75%	75%	83%	52%	83%	52%	91%	74%	25%	52%	60% -
OR135_ARIA_cr	- 58%	95%	894	76%	75%	82%	51%	82%	51%	91%	74%	25%	52%	59% -
OR135_ARIA_u	- 53%	82%	84%	733	90%	78%	62%	78%	62%	83%	88%	31%	56%	75% -
OR135_ARIA_ur	- 52%	79%	80%	88%	742	73%	60%	73%	60%	81%	88%	28%	53%	72% -
OR135_ASDP-CNS_c	- 60%	84%	85%	69%	66%	560	51%	100%	51%	90%	68%	23%	47%	54% -
OR135_ASDP-CNS_u	- 52%	77%	77%	81%	78%	80%	362	80%	100%	80%	80%	33%	57%	68% -
OR135_ASDP-Rosetta_c	- 60%	84%	85%	69%	66%	100%	51%	560	51%	90%	68%	23%	47%	54% -
OR135_ASDP-Rosetta_u	- 52%	77%	77%	81%	78%	80%	100%	80%	362	80%	80%	33%	57%	68% -
OR135_CYANA_c	- 60%	83%	84%	67%	68%	78%	46%	78%	46%	880	68%	21%	44%	53% -
OR135_CYANA_u	- 51%	77%	79%	85%	86%	72%	57%	72%	57%	81%	684	26%	51%	67% -
OR135_Ponderosa_r	- 51%	83%	82%	89%	84%	82%	77%	82%	77%	84%	83%	192	73%	78%
OR135_UNIO_r	- 59%	88%	90%	84%	82%	86%	68%	86%	68%	90%	84%	36%	399	74% -
OR135_UNIO_u	53%	84%	84%	91%	89%	81%	68%	81%	68%	87%	91%	33%	61%	504 -
	0R135	0A135-	0A135	0A135	OR135	0R135-	0A135	0R135-	0A135-	0A135	0A135-	0A135	0R135	0A135
		, N	-alk-c	A GIA CI		Vally III	ADP.CN	NO.40P.CN	Pop-Po	COP-Ro	CT.	-VANA-	, UIT	UNIO-
							5,0	1511	setta_c	setta_u		-	2	

Supplementary Figure 11A. Heatmap of the fractions of overlapping long-range NOE restraints between the OR135 target and entries, determined on the basis of pseudo-atom.

	At least 5 residues apart. OR135 811 98% 99% 96% 95% 95% 88% 95% 88% 98% 95% 46% 82% 8													
OR135	811	98%	99%	96%	95%	95%	88%	95%	88%	98%	95%	46%	82%	88%
OR135_ARIA_c	98%	888	99%	97%	97%	96%	90%	96%	90%	98%	96%	51%	85%	90%
OR135_ARIA_cr	98%	99%	894	97%	97%	96%	89%	96%	89%	98%	96%	49%	84%	88%
OR135_ARIA_u	96%	97%	97%	731	98%	94%	90%	94%	90%	96%	96%	51%	86%	90%
OR135_ARIA_ur	92%	93%	93%	95%	742	91%	87%	91%	87%	94%	95%	48%	80%	88%
OR135_ASDP-CNS_c	96%	96%	97%	90%	92%	200	82%	100%	82%	99%	92%	42%	78%	80% -
OR135_ASDP-CNS_u	92%	92%	94%	95%	95%	93%	154	93%	100%	94%	93%	45%	82%	84%
OR135_ASDP-Rosetta_c	96%	96%	97%	90%	92%	100%	82%	200	82%	99%	92%	42%	78%	80%
OR135_ASDP-Rosetta_u	92%	92%	94%	95%	95%	93%	100%	93%	154	94%	93%	45%	82%	84%
OR135_CYANA_c	98%	97%	97%	95%	96%	96%	88%	96%	88%	824	96%	46%	81%	87%
OR135_CYANA_u	95%	95%	95%	96%	98%	94%	87%	94%	87%	97%	669	46%	81%	88%
OR135_Ponderosa_r	99%	99%	99%	99%	99%	99%	98%	99%	98%	99%	99%	187	96%	99%
OR135_UNIO_r	98%	98%	99%	99%	98%	97%	95%	97%	95%	97%	98%	55%	392	92%
OR135_UNIO_u	96%	98%	98%	98%	98%	96%	93%	96%	93%	97%	98%	53%	87%	488 -
	0R135	0R135_M	ORISS	ORISS	ORISS	OR135-M	ORI35-M	ORIJS	ORISS	OR135_C	ORISS	0R135_P	OR135_U	0R135_U
		3	MA/C	MA CI	MALL .	ALA UI	DP-CNS-C	DP.ONS.U	DP-Rosetta	DP-Rosetta	ANAC	AMAL	inderosal	MIOL

Supplementary Figure 11B. Heatmap of the fractions of overlapping long-range NOE restraints between the OR135 target and entries, determined on the basis of residue.

### Supplementary Table 1.

						Conversions		NOF		WI	MP	MP Clash coore	WI	MP Ø Romosbondron	PC_NMR	WI	PC_NMR					Ambia	Queen	NOE	NOE	NOE	dihedral	dihedral	dihedral
EntryID Target	Program Type	Entry	Invalid	Conformers	Accuracy	e	ROG	completeness	Score	quality	Z-score	(ndb style)	n annearance	disallowed	φ/ψ Z-score	7.	score	Intra-NOFs	Sea-NOEs	Med-NOEs	Long-NOEs	NOFs	n				dibedrals >1°	> 3°	> 5° RDCs
HR2876B HR2876B	Troplan type	HR28768	invana	20	riccurucy	0.62	green	73.2	0.93	2 10	0.02	9.06	=1 93	disaliowed	-0.04	-0.44	-0.30	699	628	519	1147	0	66 278	119	11	1 0.5 1	112 8	38	15 0
255 HR2876B	ARIA	HR2876B ARIA c		10	0.72	0.36	green	75.2	0.93	2.09	-0.74	12.36	-1.55	2.1	-0.35	-0.45	-0.53	619	1333	451	942	580	45.138	208	112	53	172 173	91	59 0
256 HR2876B	ARIA	HR2876B_ARIA_cr		10	0.78	0.53	green	75.5	0.93	2.00	-1.32	16.63	-2.24	3.4	-0.98	-0.67	-1.06	627	1340	447	946	609	75.970	315	202	143	172 272	170	138 121
240 HR2876B	ARIA	HR2876B_ARIA_u		10	0.91	0.58	green	74.4	0.90	2.42	-3.56	27.49	-2.23	1.6	-0.98	-2.37	-1.54	1041	1224	1024	2180	1447	66.538	7755	6749	5793	172 267	175	107 0
241 HR2876B	ARIA	HR2876B_ARIA_ur		10	1.05	0.70	green	74.5	0.91	1.72	-4.18	31.03	-3.06	2.4	-1.77	-2.14	-2.37	1057	1201	1067	2132	1461	80.438	7473	6483	5580	172 497	345	257 121
313 HR2876B	ASDP-CNS	HR2876B_ASDP-CNS_c		20	0.97	0.54	green	66.4	0.92	2.05	-1.70	21.32	-1.82	0.1	-0.59	-1.51	-1.01	493	441	381	759	0	87.979	85	0	0	164 362	116	64 0
2/8 HR28/0B	ASDP-CNS ASDP-Rosetta	HR2876B_ASDP-CINS_UF		20	0.76	0.82	green	66.8	0.91	2.35	-0.92	15.57	-2.41	0.1	-0.55	-2.95	-1.42	509	429	297	751	0	75.089	189	9	0	164 422	167	118 120
279 HR2876B	ASDP-Rosetta	HR2876B ASDP-Rosetta ur		20	0.88	0.53	green	66.2	0.92	2.04	-0.22	10.32	-0.03	0	0.24	2.86	0.83	569	440	440	809	0	75.689	466	309	221	164 183	118	88 120
244 HR2876B	autonoe-Rosetta-alpha	HR2876B_autonoe-Rosetta-alpha_ur		10	1.50	1.17	green			1.77	0.99	2.78	-1.36	0.8	-0.59	5.07	0.41	0	0	0	0	0		0	0	0	0 215	183	143 0
325 HR2876B	Cheshire	HR2876B_Cheshire_s		10	3.06	3.19	green			0.37		8.21	-3.47			-1.09		0	0	0	0	0		0	0	0	0 360	323	284 0
326 HR2876B	Cheshire-YAPP	HR2876B_Cheshire-YAPP_c		50	0.80	0.46	orange		0.93	0.60		19.36	-5.16			-6.86		0	0	0	0	0		0	0	0	0 2576	2291	1979 0
262 HR2876B	Cheshire-YAPP	HR2876B_Cheshire-YAPP_u		100	1.00	0.48	orange		0.91	1.52	-10.04	30.71	-3.21	1.6	-1.57	-7.20	-4.91	0	0	0	0	0		0	0	0	0 1585	991	675 0
283 HK28/0B	CS-HM-ROSELLA	HR2876B_CS-HM-ROSetta_Sr		20	2.30	0.28	orange	76.1	0.02	-1.72	0.67	5.12	-2.19	0.1	-1.02	-4.09	.2.21	675	502	525	1194	0	90 712	22	0	0	0 26	25	20 0
227 HR2876B	Ponderosa	HB2876B_Ponderosa_r		20	1.29	0.30	orange	70.1	0.89	0.52	-2.24	23.86	-0.55	0.1	-0.35	-6.03	-2.31	577	482	380	795	0	67.914	64	0	0	316 513	417	366 0
239 HR2876B	UNIO	HR2876B_UNIO_r		20	1.05	0.58	orange	54.8	0.89	0.27	-5.09	36.05	-4.78	0.4	-2.52	-6.63	-5.14	462	435	265	579	0	66.952	238	51	27	274 1092	956	851 0
238 HR2876B	UNIO	HR2876B_UNIO_u		20	1.03	0.45	red	65.5	0.87	0.84	-12.88	75.19	-5.56	3.1	-3.62	-6.74	-6.39	795	405	617	1228	0	88.163	521	134	82	274 1279	1091	958 0
HR2876C HR2876C		HR2876C		20		0.53	green	68.0	0.89	2.41	-0.63	14.75	-1.70	0	0.12	-0.39	0.24	744	648	739	907	0	54.959	105	2	0	120 104	32	7 0
303 HR2876C	ARIA	HR2876C_ARIA_cr		15	1.05	0.12	green	62.5	0.90	2.39	-1.23	20.13	-1.32	2.8	-0.39	-0.70	-0.47	431	1008	451	500	325	54.387	94	19	12	140 251	175	152 95
304 HR2876C	ARIA	HR2876C_ARIA_cr(10-97)		15	1.05	0.12	green	62.5	0.90	2.39	-1.50	20.13	-1.32	2.8	-0.39	-0.70	-0.47	431	1008	451	500	325	54.387	94	19	12	140 240	164	142 95
307 HR2876C	ASDP-Rosetta	HR2876C_ASDP-Rosetta_cr		20	0.81	0.42	green	62.2	0.80	2 91	-2.23	20.35	-0.18	0.1	-0.04	3.91	-0.18	529	425	515	506	440	82 340	1434	476	109	140 280	139	85 0
301 HR2876C	ASDP-Rosetta	HR2876C ASDP-Rosetta ur		20	1.41	0.65	green	58.2	0.83	1.93	-1.20	15.18	-0.07	0	0.67	2.08	1.12	569	393	515	443	0	80.670	1299	557	229	158 334	251	184 0
288 HR2876C	autonoe-Rosetta-alpha	HR2876C_autonoe-Rosetta-alpha_ur		10	1.16	0.79	green		0.85	3.18	0.92	3.63	0.45	0	0.79	6.20	2.19	0	0	0	0	0		0	0	0	0 138	98	76 0
310 HR2876C	BE-metadynamics	HR2876C_BE-metadynamics_s		28	1.68	1.66	green		0.77	0.70	1.13	2.35	-3.97	0.4	-1.06	-4.91	-2.07	0	0	0	0	0		0	0	0	0 1036	859	725 0
309 HR2876C	Cheshire	HR2876C_Cheshire_s		10	1.84	1.32	green			2.05	-1.70	19.03	-2.14	0.6	-0.04	-0.68	0.06	0	0	0	0	0		0	0	0	0 324	291	254 0
308 HR2876C	Cheshire-YAPP	HR2876C_Cheshire-YAPP_c		50	1.05	0.56	red		0.90	1.68	-5.08	32.06	-5.05	1.3	-2.36	-7.62	-5.03	0	0	0	0	0		0	0	0	0 2389	2040	1773 0
300 HR28/6C	Cheshire-YAPP	HR2876C_CS_HM_DR_Posetta_ur		50	2.07	1.14	red		0.85	0.01	-10.58	2 95	-7.18	4.9	-4.48	-7.74	-7.69	0	0	0	0	0		0	0	0	0 3896	3627	3396 0
285 HB2876C	CS-HM-Bosetta	HB2876C_CS-HM-Br-Hosetta_ur		11	1.40	0.00	green		0.01	1.99	0.99	3.30	-1.00	0.7	0.12	6.31	1.24	0	0	0	0	0		0	0	0	0 245	197	171 0
286 HR2876C	CS-Rosetta	HR2876C CS-Rosetta s		10	0.96	0.82	green		_	3.07	0.19	6.57	0.01	0	0.75	5.75	1.89	0	0	0	0	0		0	0	0	0 125	105	78 0
287 HR2876C	CS-Rosetta3	HR2876C_CS-Rosetta3_s		6	1.00	0.85	green			2.70	0.55	5.91	0.01	C	0.59	6.62	1.83	0	0	0	0	0		0	0	0	0 95	74	64 0
302 HR2876C	CYANA	HR2876C_CYANA_c		20	0.84	0.34	green	71.0	0.89	1.83	-0.20	12.37	-2.52	0	-0.16	-5.42	-1.60	739	607	735	940	0	73.930	64	0	0	0 466	338	274 0
297 HR2876C	CYANA	HR2876C_CYANA_u		20	0.97	0.30	orange	66.8	0.87	1.49	-2.03	21.89	-4.40	2.6	-1.65	-6.15	-3.61	724	615	690	879	0	74.135	111	0	0	0 750	612	502 0
305 HR28/6C	I-TASSER	HR28/bC_I-TASSER_C		1	1.32		green	86.2	0.82	-1.14	-1.19	18.17	-0.50	22	0.04	-3.21	-1.30	1/81	2253	1/92	2011	2/5	89.898	216	/5	28	138 3	24	16 0
238 HR2876C	Ponderosa	HB2876C_Ponderosa_r		20	0.95	0.17	orange	00.5	0.83	1.64	-2.66	27.68	-2.00	2.5	-0.04	-7.22	-2.07	526	428	419	326	0	31,966	3829	2463	1688	307 863	733	621 0
293 HR2876C	UNIO	HR2876C_UNIO_r		20	1.32	0.63	orange	48.0	0.86	1.03	-3.52	34.99	-5.13	0.1	-1.81	-6.94	-4.67	404	406	314	348	0	55.568	332	167	108	259 1218	1017	907 0
294 HR2876C	UNIO	HR2876C_UNIO_u		20	1.12	0.50	red	57.7	0.86	0.84	-7.89	64.62	-6.04	0.3	-2.91	-7.24	-5.62	648	395	624	678	0	69.220	424	166	122	258 1286	1054	937 0
HR5460A HR5460A		HR5460A		20		0.60	green	67.8	0.87	2.43	-1.36	18.85	-1.63	0	1.49	-1.71	0.83	1063	1139	1422	1349	0	47.608	408	11	0	220 6:	. 7	1 0
176 HR5460A	ARIA	HR5460A_ARIA_c		10	1.56	0.67	green	77.1	0.87	2.64	-2.76	25.98	-1.83	0.6	1.26	-1.58	0.65	1306	2657	1600	1547	3120	30.707	3511	2057	1241	266 468	288	189 0
331 HR5460A	ARIA	HR5460A_ARIA_CF	TRUE	10	1.24	0.68	green	51.4	0.86	2.80	-2.72	25.93	-1.28	0.5	1.20	-1.58	0.59	1307	2053	1588	1368	1615	65 178	5903	2139	1312	200 397	241	181 83
173 HR5460A	ASDP-CNS	HR5460A ASDP-CNS c	mor	20	1.70	0.90	green	64.3	0.87	2.29	-0.94	15.14	-2.12	0.1	1.30	-2.45	0.24	876	774	1006	770	0	76.358	965	63	5	272 955	612	433 0
174 HR5460A	ASDP-CNS	HR5460A_ASDP-CNS_cr		20	1.52	0.80	green	62.1	0.86	2.47	-1.61	17.20	-2.08	0	1.38	-2.31	0.35	858	772	1041	773	0	78.956	971	60	3	272 980	658	465 83
164 HR5460A	ASDP-CNS	HR5460A_ASDP-CNS_u		20	1.53	1.15	green	57.0	0.84	2.01	-1.00	15.99	-2.27	0	1.34	-3.22	-0.18	831	733	920	612	0	39.015	1030	85	14	274 1078	687	433 0
165 HR5460A	autonoe-Rosetta-alpha	HR5460A_autonoe-Rosetta-alpha_ur		10	3.46	2.56	green			2.85	0.86	4.17	-0.02	0	2.01	6.90	2.78	0	0	0	0	0		0	0	0	0 259	181	129 0
321 HR5460A	Cheshire Cheshire VADD	HR5460A_Cheshire_s		1	3.52	0.00	green		0.61	-0.01		5.66	-0.96			2.50		0	0	0	0	0		0	0	0	0 32	20	17 0
323 HR5460A	Chechire-YAPP	HR5460A_Cheshire-YAPP_C	TDUE	25	5.79	4.95	orange		0.87	-2.17	_	37.47	-0.28			-7.32		0	0	0	0	0		0	0	0	0 10/2	6224	5919 0
138 HR5460A	CS-HM-Rosetta	HR5460A_CS-HM-Rosetta sr	mor	10	1.70	1.33	green		3.13	3.58	0.62	5.48	1.00	C	2.32	6.95	2.96	0	0	0	0	0		0	0	0	0 201	145	103 0
168 HR5460A	CS-Rosetta	HR5460A_CS-Rosetta_s		5	3.12	2.87	green		0.75	3.28	0.02	9.32	0.82	0	2.20	6.88	2.84	0	0	0	0	0		0	0	0	0 111	63	38 0
171 HR5460A	CYANA	HR5460A_CYANA_c		20	1.03	0.44	green	67.5	0.87	1.96	-0.03	10.13	-2.78	1.4	0.83	-5.52	-1.66	1100	1149	1402	1437	0	70.903	124	0	0	0 842	582	407 0
175 HR5460A	Ponderosa	HR5460A_Ponderosa_r		20	9.33	0.93	orange		0.69	1.81	-5.54	41.76	-1.06	0	1.57	-7.57	-2.78	865	747	786	288	0	27.522	5890	4473	3569	531 1134	835	639 0
1/2 HK5460A				20	1.22	0.67	orange	60.2	0.85	0.72	-0.48	12.98	-4.45	1.3	-0.20	-6.54	-3.25	864	975	1109	926	0	70.561	874	381	316	419 1597	1237	994 0
162 HR5460A	UNIO	HR5460A_UNIO_C		20	2.26	1.40	orange	36.6	0.87	0.09	-2.70	27.52	-5.90	1	-1.61	-7.28	-3.23	475	674	375	376	0	44.366	405	410	150	415 1400	1946	1688 0
HR6430A HR6430A		HR6430A		20		0.52	green	79.7	0.94	2.66	-0.69	14.33	-1.55	0	-0.31	-0.98	-0.30	658	570	479	1211	0	41.613	174	3	0	80 44	24	18 0
166 HR6430A	ARIA	HR6430A_ARIA_c		10	0.80	0.46	green	82.2	0.94	2.97	-1.19	15.89	-1.04	1.2	-0.79	-0.90	-0.77	751	1310	552	1331	1290	38.006	817	451	247	158 311	212	132 0
144 HR6430A	ARIA	HR6430A_ARIA_u		10	0.82	0.43	green	81.1	0.94	3.02	-1.41	16.87	-0.96	1.2	-0.63	-0.96	-0.65	749	1309	556	1336	1290	36.959	844	498	255	158 316	220	139 0
167 HR6430A	ASDP-CNS	HR6430A_ASDP-CNS_c		20	1.08	0.81	green	71.9	0.92	2.59	-0.72	14.49	-1.55	0.1	-0.75	-2.56	-1.42	491	421	338	763	0	78.591	291	7	0	152 610	415	314 0
148 HK6430A	ASUP-CNS	HK043UA_ASUP-CNS_U		20	0.95	0.77	green	69.6	0.91	2.33	-0.72	14.67	-1.60	0.1	-0.87	-2.64	-1.66	503	426	352	807	0	82.795	353	28		155 640	424	301 0
147 HR6430A	Cheshire	HR6430A Cheshire s		20	1./9	0.00	red		J.04	-1.12	-1.75	32.10	-4.47	57	-3.42	-5.20	-4.55	0	0	0	0	0		0	0	0	0 210	104	56 0
169 HR6430A	Cheshire-YAPP	HR6430A_Cheshire-YAPP_c		100	0.77	0.40	orange		0.94	1.71	-3.53	13.43	-4.77	1.2	-2.60	-6.51	-4.44	0	0	0	0	0		0	0	0	0 4308	3825	3372 0
146 HR6430A	Cheshire-YAPP	HR6430A_Cheshire-YAPP_u		100	1.03	0.75	red		0.94	1.33	-4.60	13.38	-5.73	1.6	-3.19	-6.43	-5.38	0	0	0	0	0		0	0	0	0 5046	4529	4090 0
150 HR6430A	CS-DP-Rosetta	HR6430A_CS-DP-Rosetta_u		5	2.02	2.30	green		0.80	1.78	-0.66	10.45	0.25	0	0.00	4.14	0.71	0	0	0	0	0		0	0	0	0 94	74	59 0
149 HR6430A	CS-HM-Rosetta	HR6430A_CS-HM-Rosetta_sr		10	1.21	0.59	green		0.89	3.01	0.66	4.92	0.76	0	0.08	5.17	1.18	0	0	0	0	0		0	0	0	0 185	153	130 0
152 HR6430A	CYANA	HK643UA_CYANA_c		20	0.84	0.36	green	79.2	0.93	1.66	0.89	4.16	-1.71	1.2	-1.22	-4.72	-2.31	656	582	470	1204	0	79.969	10	0	0	0 417	337	284 0
163 HR6430A	Ponderosa	HR6430A_Ponderosa_r		20	1 0.90	0.34	orange	41 1	0.85	1.60	-3.72	34.94	-2.23	0.1	-1.30	-5.5/	-2.34	514	441	294	497	0	49,395	5484	3685	2485	100 1141	676	445 0
141 HR6430A	UNIO	HR6430A UNIO r		20	1.03	0.74	green	59.8	0.92	1.04	0.84	4.39	-3.58	0.7	-1.65	-5.11	-3.25	420	365	221	578	0	60.817	157	65	56	253 665	547	479 0
142 HR6430A	UNIO	HR6430A_UNIO_u		20	0.91	0.73	orange	72.3	0.94	1.34	0.48	6.70	-3.60	0.3	-1.49	-5.62	-3.13	610	523	432	1006	0	78.679	194	33	13	255 587	501	436 0

						Com		NOE DR	WI	Clack	MP Clarb coord	WI	MP % Ramachandrar	PC_NMR	WI	PC_NMR dibodrol 7				Ambig	Queen	NOE	NOE	NOE		dihedral	dihedral dih	edral
Entryl	Target	Program Type	Entry	Invalid	Conformers	s Accuracy	e ROG	completeness Score	quality	Z-score	(pdb style)	n appearance	disallowed	Z-score	Z-	score	Intra-NOEs	Seg-NOEs Med-NOEs	Long-NOEs	NOEs	n	> 0.1 Å	> 0.3 Å	> 0.5 Å	dihedrals	> 1°	> 3° >	5° RDCs
HR6470	AHR6470A		HR6470A		20		0.40 green	68.4 0.91	2.32	0.76	12.87	-0.09	0 0	0 1.57	1.62	2.66	368	429 462	2 313	0	31.906	66	2		140	28	10	0 7
15	4 HR6470A	ARIA	HR6470A ARIA c		10	0.57	0.38 green	81.7 0.91	2.76	0.16	7.95	0.26		0 1.97	1.17	1.77	414	1011 448	3 314	633	21.942	281	192	126	92	87	55	36
15	6 HR6470A	ARIA	HR6470A_ARIA_c(10-59)		10	0.58	0.30 green	80.9 0.88	3.04	0.01	10.11	0.53	1 (	0 2.08	0.42	1.66	397	878 443	310	518	34.692	312	217	142	92	83	56	26
15	5 HR6470A	ARIA	HR6470A_ARIA_cr		S	9 0.57	0.32 green	80.1 0.91	2.97	-0.10	0 10.78	1.09	) (	0 2.12	1.59	2.19	430	1009 442	2 304	633	30.763	255	185	113	92	58	42	26 7
15	7 HR6470A	ARIA	HR6470A_ARIA_cr(10-59)		10	0.60	0.37 green	79.9 0.88	2.87	-0.07	10.78	0.36	6 (	0 2.01	0.76	1.77	386	874 446	5 316	509	24.788	320	221	162	92	66	47	30 7
12	9 HR6470A	ARIA	HR6470A_ARIA_u		10	0.49	0.44 green	77.2 0.91	2.88	-0.48	12.94	0.81	0.2	2 1.81	1.12	1.71	444	1002 461	284	667	38.921	395	288	182	2 92	60	38	26
12	8 HR6470A	ARIA	HR6470A_ARIA_U(10-59)		10	0.58	0.36 green	76.5 0.88	2.88	-0.47	13.61	0.72		0 1.85	1.07	1.42	404	8/8 458	3 260	508	34.670	4//	335	232	92	91	54	35
12	/ HR64/UA	ARIA	HR6470A_ARIA_UF		10	0.47	0.36 green	76.5 0.90	2.98	-0.30	11.19	0.60		0 2.12	1.48	1.89	443	1004 456	281	650	30.880	393	261	184	92	120	74	42 7
15	8 HR6470A	ASDP-CNS	HR6470A_AKIA_UI(10-33)		20	0.70	0.72 green	67.2 0.80	2.78	-0.23	11.40	-1.31		0 1.73	-1.39	0.83	288	320 315	5 156	0	62 887	145	407	200	106	358	219	128
15	9 HR6470A	ASDP-CNS	HR6470A_ASDP-CNS_cr		20	0.75	0.80 green	58.8 0.90	2.15	-0.13	13.48	-1.45		0 1.69	-1.50	0.83	231	162 309	150	0	36,233	72	5		100	335	200	120 7
13	6 HR6470A	ASDP-CNS	HR6470A_ASDP-CNS_u		20	0 1.00	0.51 green	58.7 0.85	2.77	-1.11	23.65	-0.14	1 (	0 0.83	-2.40	0.06	297	322 329	144	0	63.040	658	515	425	106	240	97	42
13	5 HR6470A	ASDP-CNS	HR6470A_ASDP-CNS_ur		20	0 1.00	0.51 green	60.1 0.85	2.77	-1.11	23.65	-0.14	1 (	0 0.83	-2.40	0.06	302	323 328	3 140	0	62.120	73	3	3	3 106	240	97	42
18	5 HR6470A	autonoe-Rosetta-alpha	HR6470A_autonoe-Rosetta-alpha_u(11-58)		10	0.78	0.83 green		3.47	-0.47	4.99	1.28	8 (	0 1.85	7.96	1.42	0	0 (	0 0	0		0	0	0	0 0	61	39	25
18	6 HR6470A	autonoe-Rosetta-alpha	HR6470A_autonoe-Rosetta-alpha_ur		10	0.83	0.77 green	0.88	3.01	0.76	6.75	0.63	s (	0 1.57	7.29	2.66	0	0 (	0 0	0		0	0	0	0 0	98	72	48
13	1 HR6470A	Cheshire	HR6470A_Cheshire_s		1	1 0.81	green		-0.18	0.00		1.02	1		0.29	0.00	0	0 0	0	0		0	0		0	5	3	0
15	3 HR6470A	Cheshire-YAPP Cheshire YAPP	HR6470A_Cheshire-YAPP_C		20	0.57	0.38 orange	0.89	1.81	-0.32	15.61	-4.35		0 -0.08	-6.16	-3.08	0	0 (		0		0	0			527	399	302
13	2 HR6470A	CRESNIRE-YAPP	HK6470A_CREShire-YAPP_U		50	0.64	0.34 red	0.87	1.50	0.05	12.11	-4.//	0.4	4 -0.39	-6.28	-3.49	0	0 0		0		0	0			1538	1203	908
14	1 HR6470A	CS-Rosetta	HR6470A_CS-HWI-ROSetta_s		10	5 0.54	0.32 green	0.89	2.03	0.06	4.50	0.37		0 2.30	7.00	3.13	0	0 0				0	0			38	22	17
13	7 HR6470A	CYANA	HR6470A CYANA c		20	0.53	0.42 green	73.0 0.91	2.30	1.27	2.16	-0.85	5 0	0 1.65	-2.53	0.59	357	438 428	3 327	0	42.061	2	0		0 0	220	146	90
13	3 HR6470A	CYANA	HR6470A_CYANA_u		20	0.59	0.46 green	72.2 0.90	1.92	1.07	3.78	-1.54		0 1.61	-2.91	0.53	385	461 397	262	0	36.170	7	0	0	0 0	242	152	92
12	5 HR6470A	Ponderosa	HR6470A_Ponderosa_r		20	0.65	0.07 green	0.83	2.28	-0.81	20.95	1.63	8 (	0 1.22	-6.08	-1.83	399	401 361	1 258	0	53.100	2	0	0	256	421	333	266
12	2 HR6470A	UNIO	HR6470A_UNIO_r		20	0 1.09	0.61 green	52.8 0.86	0.75	1.25	2.36	-2.55	5 2.1	1 0.90	-4.77	-1.42	206	291 179	135	0	28.179	23	7	4	1 188	519	416	360
12	3 HR6470A	UNIO	HR6470A_UNIO_u		20	0.66	0.51 orange	65.2 0.90	2.30	1.03	4.05	-1.32	2 (	0 1.42	-4.65	-0.83	334	417 344	1 204	0	37.227	72	17	14	1 185	323	238	157
HR8254	IA HR8254A		HR8254A		20	0	0.72 green	59.2 0.83	2.79	-0.88	15.61	-2.26	6 0.1	1 1.53	-1.30	1.01	387	408 444	1 300	0	29.160	999	811	746	5 125	64	2	0
27	3 HR8254A	ARIA	HR8254A_ARIA_c	20110	10	0 1.50	0.52 green	66.0 0.83	4.23	-1.50	22.61	. 0.90	0 (	0 2.48	1.10	2.66	362	865 396	242	325	48.112	377	135	66	5 130	80	23	16
33	5 HR8254A	AKIA	HK8254A_ARIA_U	TRUE	10	0 10.71	0.70 orange	57.2 0.42	-1.04	0.02	68.05	-5.81	10	0 2.44	-3.20	3.55	648	852 584	669	497	61.400	1988	1555	85/	130	1100	/29	493
27	A HR8254A	ASDP-Rosetta	HR8254A_ASDP-Rosetta_u		20	1.74	1.08 green	53.6 0.84	4.17	-0.03	0.75	0.81	1.0	6 2.44	4.58	3.55	305	316 414	149	0	70.915	959	350	42	108	521	506	425
26	5 HR8254A	autonoe-Rosetta-aloba	HR8254A_HSDF-Rosetta_alpha_c		10	2 68	0.99 green	34.0 0.81	3.05	-0.08	1 17	0.73	0.0	0 2.05	7.50	3.25	303	0 (	120	0	70.915	1008	330	112	110	330	71	50
26	4 HR8254A	autonoe-Rosetta-alpha	HR8254A autonoe-Rosetta-alpha u		10	3 91	0.97 green	0.73	3.84	1.01	3.20	1.30		0 2.05	6.89	3.19	0	0 0		0		0	0			145	116	102
32	9 HR8254A	Cheshire	HR8254A Cheshire s		1	1 2.77	green		0.22			-1.75	5		1.90	0.25	0	0 0	0 0	0		0	0		0 0	35	30	23
27	0 HR8254A	CS-HM-Rosetta	HR8254A_CS-HM-Rosetta_s		11	1 1.38	0.48 green	0.81	4.21	1.40	0.97	1.49	) (	0 2.60	8.49	3.78	0	0 (	0 0	0		0	0	0	0 0	78	59	47
26	9 HR8254A	CS-Rosetta	HR8254A_CS-Rosetta_s		5	5 1.52	1.31 green	0.79	4.18	0.93	3.82	1.80	0 (	0 2.71	9.25	4.02	0	0 (	0 0	0		0	0	0	0 0	36	25	14
29	9 HR8254A	CYANA	HR8254A_CYANA_c		20	0 1.04	0.80 green	61.3 0.82	2.44	0.72	5.89	-2.18	3 (	0 1.26	-3.56	0.06	368	403 444	1 295	0	56.637	20	0	0	0 0	375	279	227
27	1 HR8254A	I-TASSER	HR8254A_I-TASSER_c		10	0 1.31	0.28 green	86.6 0.77	3.84	-1.04	19.32	3.81		0 3.11	0.02	2.37	1241	1562 1630	854	65	56.519	863	392	229	130	64	27	7
27	2 HR8254A	I-TASSER	HR8254A_I-TASSER_u		10	0 1.31	0.25 green	80.0 0.74	3.22	-2.75	32.27	2.62	2 (	0 2.56	-0.80	1.71	1616	1434 1925	5 534	92	65.600	1486	806	552	130	128	90	43
26	8 HR8254A	Ponderosa	HR8254A_Ponderosa_r		20	3.01	0.02 green	38.9 0.75	2.59	-1.76	24.89	-0.62	2 (	0 2.36	-6.10	-0.18	398	451 366	5 211	0	41.794	3493	2768	2239	95	845	421	141
26	3 HR8254A	UNIO	HR8254A_UNIO_u		20	0 1.45	0.95 green	49.2 0.81	2.81	-4.24	38.69	-2.52	2 0.1	1 1.49	-2.69	0.53	267	341 278	167	0	38.784	116	0		140	579	436	359
21	E OP125	ADIA	OR135		20	0.74	0.84 green	71.9 0.90	4.00	-0.81	14.76	-1./0		0 0.31	-0.29	0.41	643 560	1112 52	967	767	50.203 65.240	201	112	71	108	122	52	41
21	4 08125	ARIA	OR135_ARIA_C		10	0.74	0.34 green	77.4 0.90	5.20	-1.03	14.57	0.80		0 0.79	0.90	0.95	500	1122 532	784	767	65.349 53.936	201	113	/1	120	98	194	122 10
19	6 OR135	ARIA	OR135_ARIA_LI		10	0.84	0.45 green	68.4 0.89	4.01	-1.19	14.57	0.09		0 0.43	-0.13	0.12	457	930 433	641	666	66 264	482	283	173	128	147	92	72 10
19	9 OR135	ARIA	OR135 ARIA ur		10	1.00	0.60 green	69.6 0.89	3.89	-1.76	18.65	-2.25	0.2	2 -0.28	-0.81	-0.41	462	924 408	652	636	53.221	509	341	226	128	360	240	157 10
31	2 OR135	ASDP-CNS	OR135_ASDP-CNS_c		20	0.88	0.46 green	64.9 0.89	4.94	-2.34	26.02	-0.86	5 (	0 -0.04	-1.10	-0.59	445	435 401	590	0	83.467	133	4		138	276	113	59
20	7 OR135	ASDP-CNS	OR135_ASDP-CNS_u		20	0 1.06	0.56 green	54.6 0.86	4.79	-2.14	24.57	-0.75	5 (	0 0.00	-2.07	-0.95	334	379 273	388	0	39.594	46	0	0	136	308	145	103
31	1 OR135	ASDP-Rosetta	OR135_ASDP-Rosetta_c		20	0.83	0.14 green	65.8 0.89	5.64	0.88	4.34	2.51		0 0.47	5.54	1.36	445	435 401	1 590	0	83.467	366	113	44	1 138	56	47	45
20	8 OR135	ASDP-Rosetta	OR135_ASDP-Rosetta_u		20	0.85	0.19 green	54.7 0.88	5.39	1.10	2.85	2.41		0 0.67	5.23	1.89	334	379 273	388	0	39.594	153	25	1	136	76	72	68
20	9 OR135	autonoe-Rosetta-alpha	OR135_autonoe-Rosetta-alpha_ur		9	9 1.01	0.52 green	0.85	5.50	1.46	0.38	0.28	8 (	0 0.67	7.96	2.07	0	0 0	0	0		0	0		0	136	99	82
21	6 OP125	Chechire VARR	OR125_Chechico VARR c		1 100	1.29	0.85 rod	0.05	1.16	0.70	8.29	-3.46		0 -1.22	0.49	-0.65	0			0		0				E147	19	4016
21	6 OR135	Cheshire-YAPP Cheshire YAPP	UR135_Cheshire-YAPP_C		100	1.21	0.85 red	0.85	1.45	-4.73	16.59	-5.35	5.9	9 -3.11	-7.38	-6.27	0	0 0		0		0	0			514/	4596	4016
21	1 OR135	CS-HM-Rosetta	OR135_CIESHIRE-TAPP_0 OR135_CS-HM-Rosetta_sr		100	1.5/	0.94 red	0.82	5.67	-1.80	0.90	-6.54	0.5	0 0.75	7.52	-0.15	0	0 0		0		0				230	181	141
20	5 OR135	CS-Rosetta	OR135 CS-Rosetta s		20	5 0.93	0.59 green	0.85	5.35	1.12	2.56	0.08		0 0.87	8.36	2.25	0	0 0	0 0	0		0	1 0			233	18	13
21	2 OR135	CYANA	OR135 CYANA c		20	0.75	0.38 green	75.7 0.91	4.03	0.78	4.17	-2.58	3 0	0 -0.47	-4.96	-2.13	626	659 584	966	0	74.336	20	0		0 0	359	277	198
19	8 OR135	CYANA	OR135_CYANA_u		20	0.98	0.38 green	66.3 0.89	3.55	0.88	1.70	-2.10		0 -0.35	-4.24	-1.71	509	524 464	733	0	74.119	46	0		0 0	377	292	228
22	6 OR135	Ponderosa	OR135_Ponderosa_r		20	0 1.91	0.28 orange	0.79	2.47	-2.79	27.68	0.93	0.3	3 0.87	-7.39	-2.48	300	392 184	1 209	0	25.484	896	637	507	298	774	719	647
20	4 OR135	UNIO	OR135_UNIO_r		20	0 1.01	0.56 orange	54.1 0.88	2.34	-2.06	23.08	-3.93	8 (	0 -0.71	-6.86	-4.08	353	438 249	403	0	51.397	75	19	9	226	620	472	377
20	3 OR135	UNIO	OR135_UNIO_u		20	0.98	0.60 orange	60.2 0.87	2.97	-3.84	28.58	-3.62	2 0.2	2 -0.55	-7.17	-3.67	395	396 354	508	0	63.971	63	21	17	7 225	743	623	510
OR36	OR36		OR36		20	2	0.77 green	60.2 0.91	4.65	-1.73	18.92	-1.71	4	0 0.90	-1.71	0.41	749	1080 824	1090	0	46.892	134	2	-	196	155	54	10
18	9 UR36	AKIA	UK36_AKIA_C		10	1.02	0.80 green	70.4 0.91	4.71	-1.23	17.38	-0.48		0 1.06	-1.11	0.71	909	2298 890	1105	1717	40.379	921	567	344	236	334	216	147
19	2 0826	ARIA	OR30_ARIA_CC	TRUE	9	1.04	0.72 green	/1.5 0.91	4.55	-1.35	20.40	-2.19	0.2	2 0.35	-1.11	U.18	912	2292 856	1161	1/40	51.904	10010	392	229	236	496	329	243 16
10	2 OR36	ASDP-CNS	OB36_ASDP-CNS_C	TRUE	20	11.74	0.98 green	57.1 0.90	-0.51	-1.83	21 44	-5.78	03	3 0.30	-3.07	-0.65	586	734 53	673	1//1	35.997	701	50	7215	236	1058	689	411
19	3 OR36	ASDP-CNS	OR36 ASDP-CNS cr		20	1.38	0.96 green	56.5 0.90	4,38	-1.98	22 76	-1.62	2 (	0 0.75	-3.46	-0,53	591	737 543	662	0	80.217	569	30		240	855	530	309 16
17	8 OR36	ASDP-CNS	OR36_ASDP-CNS_u		20	0 1.35	1.00 green	50.7 0.88	3.96	-1.72	20.99	-2.03		0 0.63	-3.43	-0.59	590	683 520	502	0	63.010	577	34	2	2 246	1074	626	402
17	9 OR36	ASDP-CNS	OR36_ASDP-CNS_ur		20	0 1.33	0.91 green	48.2 0.87	4.38	-2.57	26.57	-1.90		0 0.43	-3.80	-0.89	591	684 502	2 540	0	74.609	629	83	3	3 246	1107	678	449 16
18	4 OR36	autonoe-Rosetta-alpha	OR36_autonoe-Rosetta-alpha_ur		10	0 1.46	1.24 green	0.89	5.26	0.82	3.98	-0.27	/ (	0 1.26	6.62	2.25	0	0 0	0 0	0		0	0	0	0 0	224	154	115
32	4 OR36	Cheshire	OR36_Cheshire_s		1	1 2.64	green		1.59			-1.95	5		-0.65		0	0 (	0 0	0		0	0	0	0 0	39	34	28
19	4 OR36	Cheshire-YAPP	OR36_Cheshire-YAPP_c		50	0 1.20	0.96 red	0.88	2.26	-3.39	32.86	-5.46	0.1	1 -1.02	-7.92	-4.55	0	0 0	0 0	0		0	0	0	0 0	4246	3669	3068
18	8 OR36	Cheshire-YAPP	OR36_Cheshire-YAPP_u		50	0 1.24	0.91 red	0.86	2.26	-4.20	23.87	-5.67	0.1	1 -1.26	-7.74	-5.03	0	0 0	0	0		0	0		0	4491	3840	3232
18	3 UR36	CS-UP-Rosetta	UK36_CS-DP-Rosetta_u	L	5	1.42	1.07 green	0.88	5.32	0.80	4.58	1.14	9 9	U 1.69	6.10	2.31	0	0 0	0	0	06.42	0	0		1 0	91	66	45
18	/ UK3b	CYANA	OR30_CTANA_C		20	1.19	0.51 green	58.0 0.91	3.95	0.34	15.04	-2.08		0 0.15	-4.54	-1.42	/74	1037 816	1200	0	85.234	73	1 0			786	5/6	432
1/	2 0836	Ponderosa	OR36 Ponderosa r		20	2 78	0.33 green	30.7 0.79	3.14	-0.90	22 79	-3.26		0 1.12	-0.30	-2.60	907	900 8bl	2/ 1262		35.044	4660	3304	2200	102	/90	433	270
18	0 0836	UNIO	OB36_UNIO_r		20	1.57	1.06 green	36.1 0.88	1.80	-2.19	4 61	-3 20	0.0	3 -0.35	-6.22	-2.00	321	608 221	252	0	31,632	230	5304	2390	3 372	1161	942	757
18	1 OR36	UNIO	OR36 UNIO u		20	1.43	0.70 orange	47.6 0.86	2.19	-0.80	14.01	-3.25	0.3	0 -0.39	-7.31	-3.08	773	600 705	872	0	73.111	239	307	249	3/2	1159	922	740
		12 2										5.51				0.50	. / 3	/00	1 5/1									

										WI	MP	MP	WI	MP	PC_NMR	wi	PC_NMR						Queen M	4OE	NOE	NOE		dihedral	dihedral	dihedral	
	-		_			Converg	nc	NOE	DP	packing	Clash	Clash score	Ramachandra	% Ramachandran	φ/ψ	X	dihedral Z-				Am	nbig- Ir	nformatio viol	ations	violations	violations		violations v	riolations	violations	
EntryID	Target	Program Type	Entry	Invalid	Conformers	Accuracy e	ROG	completeness	Score	quality	Z-score	(pdb style)	n appearance	disallowed	Z-score	Z-	score	Intra-NOEs	Seq-NOEs Me	I-NOEs Long-N	OEs NO	DEs	n >1	0.1 A	> 0.3 A	> 0.5 A	dihedrals	> 1°	> 3*	>5°	RDCs
StT322	StT322		StT322		20	) (	.57 green	63.4	0.81	2.39	-0.06	12.63	-4.23	0	-2.91	1 -3.68	-4.02	280	320	137	371	0	32.415	252	1	0	70	28	0	0	1
225	StT322	ARIA	StT322_ARIA_c		10	2.19 (	.51 green	52.0	0.73	3.13	-1.08	17.35	-1.60	0.2	-2.12	2 -0.58	-2.66	218	694	111	232	393	31.127	249	148	87	78	244	128	97	
333	StT322	ARIA	StT322_ARIA_u	TRUE	10	9.27	.02 orange	62.4	0.51	-4.23		63.77	-5.92			-5.54		742	741	716	1789	2074	105.501	13546	11966	10478	78	557	364	254	1
229	StT322	ASDP-CNS	StT322_ASDP-CNS_c		20	1.91 (	.67 red	62.7	0.80	1.05	-0.75	19.92	-2.66	0.1	-2.71	1 -7.79	-6.62	213	284	105	258	0	43.591	42	11	1	92	368	171	91	
222	StT322	ASDP-CNS	StT322_ASDP-CNS_u		20	2.85	.43 red	48.8	0.77	0.07	-1.08	21.45	-2.71	0	-1.49	-7.76	-5.14	237	247	79	143	0	39.702	52	2	0	92	399	249	155	
230	StT322	ASDP-Rosetta	StT322_ASDP-Rosetta_c		20	1.49 (	.81 green	61.5	0.81	2.48	-0.71	17.73	-0.50	0	-1.30	3.58	-1.06	213	284	105	258	0	43.591	175	52	10	92	266	195	130	
223	StT322	ASDP-Rosetta	StT322_ASDP-Rosetta_u		20	2.28	.50 green	48.0	0.79	1.96	0.03	9.89	0.66	0	-0.47	7 5.15	0.06	237	247	79	143	0	39.702	189	75	14	92	275	225	177	
220	StT322	autonoe-Rosetta-alpha	StT322_autonoe-Rosetta-alpha_u		10	4.34 (	.44 green			1.69	1.41	0.00	0.62	1.7	-1.30	5.83	-0.06	0	0	0	0	0		0	0	0	0	116	104	84	
231	StT322	Cheshire	StT322_Cheshire_s		10	1.30 (	.82 green		0.70	4.47	0.65	4.39	-3.14	0.5	-2.40	0.01	-2.13	0	0	0	0	0		0	0	0	0	190	174	158	
224	StT322	CYANA	StT322_CYANA_c		20	2.83 (	.50 orang	e 60.6	0.77	1.73	0.79	6.41	-3.34	1.1	-3.38	3 -6.44	-5.32	278	296	132	329	0	38.716	7	0	0	0	337	294	238	
221	StT322	Ponderosa	StT322_Ponderosa_r		20	3.69 (	.25 orang	2	0.59	-0.14	-2.12	32.37	-0.38	0.1	-1.46	5 -5.92	-4.02	231	178	74	81	0	14.679	74	0	0	211	329	286	244	
YR313A	YR313A		YR313A		20	) (	.97 green	61.2	0.87	2.70	-0.58	12.18	-2.87	0	-0.55	5 -1.31	-0.59	631	743	413	754	0	49.414	74	1	0	122	130	37	3	4
253	YR313A	ARIA	YR313A_ARIA_c		10	1.12 (	.50 green	62.0	0.84	2.93	-0.93	16.27	-1.25	1	-0.63	3 -0.77	-0.83	608	1495	354	655	575	28.782	372	230	105	174	163	115	78	
254	YR313A	ARIA	YR313A_ARIA_cr		10	1.39 (	.64 green	61.3	0.83	2.42	-1.03	16.66	-3.21	0	-1.26	-0.49	-1.24	602	1500	346	616	545	47.096	386	218	123	174	417	235	154	11
334	YR313A	ARIA	YR313A_ARIA_ur	TRUE	10	14.15 12	.74 orang	68.4	0.59	-2.20		34.61	-5.83			-4.20		822	1308	454	176	519	26.691	3931	3217	2629	174	1113	840	643	11
315	YR313A	ASDP-CNS	YR313A_ASDP-CNS_c		20	1.81 (	.74 green	54.6	0.80	2.33	-2.64	27.03	-2.08	0.1	-1.06	6 -2.78	-2.01	478	555	297	484	0	79.991	105	4	0	198	412	146	51	
276	YR313A	ASDP-CNS	YR313A_ASDP-CNS_u		19	1.44	.11 green	51.4	0.84	2.48	-1.42	20.02	-2.24	0.1	-0.55	5 -3.12	-1.36	506	529	297	455	0	60.701	124	7	0	198	519	202	99	1
316	YR313A	ASDP-Rosetta	YR313A_ASDP-Rosetta_c		20	1.75	.06 green	60.0	0.83	2.82	-0.18	10.02	-0.19	0	0.20	4.65	1.06	487	558	301	475	0	77.327	259	82	25	198	211	149	103	
277	YR313A	ASDP-Rosetta	YR313A_ASDP-Rosetta_u		20	1.45 (	.75 green	48.3	0.84	2.97	-0.11	9.98	-0.03	0.1	0.12	2 4.07	0.83	506	529	297	455	0	60.701	395	306	241	198	139	90	66	1
245	YR313A	autonoe-Rosetta-alpha	YR313A_autonoe-Rosetta-alpha_ur		10	1.20 (	.75 green			3.83	0.83	3.82	-0.90	0	-0.04	4 6.40	1.30	0	0	0	0	0		0	0	0	0	169	138	112	
327	YR313A	Cheshire	YR313A_Cheshire_s		1	2.04	green			-1.21			-3.28			-1.84		0	0	0	0	0		0	0	0	0	40	34	31	
328	YR313A	Cheshire-YAPP	YR313A_Cheshire-YAPP_c		50	1.12 (	.80 red		0.86	1.10		31.19	-5.86			-7.11		0	0	0	0	0		0	0	0	0	2989	2628	2301	
261	YR313A	Cheshire-YAPP	YR313A_Cheshire-YAPP_u		100	1.73	.53 orang	2	0.82	1.14	-6.95	21.30	-3.19	1.5	-0.63	3 -7.88	-4.44	0	0	0	0	0		0	0	0	0	1605	1123	821	
282	YR313A	CS-HM-Rosetta	YR313A_CS-HM-Rosetta_sr		1	2.17	orang	2		-0.37	0.64	5.19	-2.72	0	-1.02	2 6.03	0.06	0	0	0	0	0		0	0	0	0	31	22	16	
260	YR313A	CYANA	YR313A_CYANA_c		20	1.21 (	.67 green	64.3	0.86	1.84	0.96	3.70	-2.88	0.3	-1.46	-4.65	-2.78	626	724	389	778	0	82.907	51	0	0	0	545	424	338	
228	YR313A	Ponderosa	YR313A_Ponderosa_r		20	1.67 (	.12 green		0.78	2.23	-3.21	27.18	-0.80	0	-0.43	3 -6.48	-2.90	638	723	426	737	0	73.187	14	2	0	412	586	523	440	
237	YR313A	UNIO	YR313A_UNIO_r		20	2.94	.81 red	40.7	0.82	-0.24	-2.71	24.38	-4.85	0.1	-1.46	6 -7.07	-4.49	291	529	102	192	0	27.243	264	77	47	314	1315	1177	1031	