



Morphological and Functional Characterization of IL-12R β 2 Chain on Intestinal Epithelial Cells: Implications for Local and Systemic Immunoregulation

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Interaction between intestinal epithelial cells (IECs) and the underlying immune systems is critical for maintaining intestinal immune homeostasis and mounting appropriate immune responses. We have previously showed that the T helper type 1 (T_H1) cytokine IL-12 plays a key role in the delicate immunological balance in the gut and the lack of appropriate levels of IL-12 had important consequences for health and disease, particularly with regard to food allergy. Here, we sought to understand the role of IL-12 in the regulation of lymphoepithelial cross talk and how this interaction affects immune responses locally and systemically. Using a combination of microscopy and flow cytometry techniques we observed that freshly isolated IECs expressed an incomplete, yet functional IL-12 receptor (IL-12R) formed solely by the IL-12R β 2 chain that albeit the lack of the complementary IL-12 β 1 chain responded to *ex vivo* challenge with IL-12. Furthermore, the expression of IL-12R β 2 on IECs is strategically located at the interface between epithelial and immune cells of the lamina propria and using *in vitro* coculture models and primary intestinal organoids we showed that immune-derived signals were required for the expression of IL-12R β 2 on IECs. The biological relevance of the IEC-associated IL-12R β 2 was assessed *in vivo* in a mouse model of food allergy characterized by allergy-associated diminished intestinal levels of IL-12 and in chimeric mice that lack the IL-12R β 2 chain on IECs. These experimental models enabled us to show that the antiallergic properties of orally delivered recombinant *Lactococcus lactis* secreting bioactive IL-12 (rLc-IL12) were reduced in mice lacking the IL-12 β 2 chain on IECs. Finally, we observed that the oral delivery of IL-12 was accompanied by the downregulation of the production of the IEC-derived proallergic cytokine thymic stromal lymphopietin (TSLP). However, further analysis of intestinal levels of TSLP in IL-12R β 2^{-/-} mice suggested that this event was not directly linked to the IEC-associated IL-12R β 2 chain. We interpreted these data as showing that IEC-associated IL12R β 2 is a component of the cytokine network operating at the interface between the intestinal epithelium and immune system that plays a role in immune regulation.

Keywords: IL12R β 2 chain, intestinal epithelial cell, immunoregulation, mucosal immunity, interleukin 12

INTRODUCTION

The constant dialog between the intestinal epithelium and underlying immune system plays a key role in intestinal immune homeostasis by shaping the appropriate immunological microenvironment (1). Intestinal epithelial cells (IECs) signal to the immune system *via* the secretion of regulatory cytokines; in the presence of commensal bacteria, IECs secrete anti-inflammatory molecules that include thymic stromal lymphopoietin (TSLP) (2) and transforming growth factor- β (3). By contrast, pathogens cause epithelial cells to release proinflammatory factors, such as IL-8 (CXCL-8), MCP-1 (CCl2) and MIP-3 α (4, 5) and might trigger rapid response to clear the infection (6). In turn, cytokines produced by the intestinal immune system may signal back to the epithelium and regulate epithelial functions such as, for example, antigen sampling by specialized M cell (7). The T helper type 1 (T_H1) cytokine IL-12 plays a variety of roles in intestinal immunity ranging from response to pathogens (8), interaction with group 1 innate lymphoid cells in maintaining barrier integrity (9) to protecting against food allergy (10). With regard to the latter we have previously shown that the intestinal levels of IL-12p70 in the gut are critical for the resistance to food allergy (10). Indeed, sensitization to the food allergen peanuts was characterized by significantly reduced intestinal levels of IL-12. One of the most convincing evidence for the regulatory role of IL-12 in the intestinal mucosa was brought about by the observation that blocking IL-12 in the gut at the time of antigen presentation by targeted oral delivery of microencapsulated neutralizing antibody significantly increased susceptibility to food allergy in mice (10). The notion that IL-12 is critical in food allergy is further strengthened by the observation that oral delivery of liposome encapsulated IL-12 suppressed allergic reaction (11). However, aspects of its mechanism(s) of action remained to be determined. We observed that IECs expressed only the low affinity IL-12 binding site IL-12R β 2 chain, but not the complementary high affinity binding site IL-12R β 1 chain. IEC-associated IL-12R β 2 operates as an incomplete, yet still operational IL12R the expression of which is under the control of immune cell-derived signals. This observation prompted us to test the hypothesis that the presence of IEC-associated IL-12R β 2 could directly affect the regulatory properties of IECs. This was done by adopting a strategy based on oral administration of a *Lactococcus lactis* strain genetically engineered to produce biologically active IL-12 (rLc-IL12) (12) in allergic wild-type (wt) and chimeric mice that lacked IL-12R β 2 on the intestinal epithelium. We observed that the lack of IL-12R β 2 on IECs significantly reduced the anti-allergy activity of rLc-IL12. Also, IL-12 mediated suppression of food allergy was accompanied by the downregulation of the IEC-derived cytokine TSLP.

These data suggested that IEC-associated IL-12R β 2 contributed to control the finely tuned regulatory cytokine network operating at the lymphoepithelial interface in the gut and impacted on the host's ability to mount appropriate immune responses.

MATERIALS AND METHODS

Mice and Bacterial Strain

Mice were maintained in an access-restricted room in conventional conditions. B6.129S1-Il12rb2tm1Jm/J (IL-12R β 2^{-/-})

and wt C57BL/6J mice that were previously used to generate chimeric mice (13) were purchased from (Jackson Laboratory, Bar Harbor, ME, USA). IL-12R β 2^{-/-} mice were maintained on BL/6 background. Chimeric mice were produced by adopting a slightly modified busulfan-based method that allowed a stable bone marrow (BM) chimerism (14). Briefly, recipient mice were treated with daily injections of 20 mg/kg busulfan (6 consecutive days for a total dose of 120 mg/kg busulfan). Twenty-four hours after the last BU injection, 5×10^6 BM cells collected from the femur and tibia of wt or IL-12R β 2^{-/-} donors and injected i.v. (tail vein) into recipient mice. Age- and sex-matched busulfan-treated IL12R β 2-deficient mice were reconstituted either with BM from wt mice (WT \rightarrow IL12R β 2^{-/-}) to generate mice lacking IL-12R β 2 in IECs (IL-12R β 2^{ΔIEC}) or BM from syngeneic IL-12R β 2-deficient mice (IL-12R β 2^{-/-} \rightarrow IL-12R β 2^{-/-}) to control for the effects of the busulfan treatment and BM reconstitution. Mice were then allowed to rest for at least 8 weeks before the beginning of the experiments. Chimerism was functionally assessed by determining the ability to produce IFN γ *in vivo* following i.p. administration of mouse IL-12 (1 μ g/mouse for 5 consecutive days) as previously described (15). Furthermore, wt C57BL/6J and chimeric mice were sensitized according to a well-established model of food (peanuts) allergy (10, 16). Ground whole peanuts were used as antigen and crude peanut extract (CPE) was prepared as described in detail previously (10, 17). Briefly, mice received intragastric (i.g.) administration (oral gavage) of 1 mg/mouse of ground whole peanut together with 10 μ g of cholera toxin (CT) (Quadrant Ltd., Surrey, UK) in PBS on days 0, 7, 14, 21 and 28. On day 35, all mice were challenged by oral gavage with CPE (1 mg/mouse divided into two doses at 30 min intervals). The administration regimen of the sensitizing mixture was based on previously established protocol (10, 16). To assess the role of rLc-IL12 in suppressing food allergy, wt and chimeric mice were gavaged with PBS (untreated) or *L. lactis* FI10611 secreting bioactive IL-12p70 (rLc-IL12) (8–10 mice/group). Description of the wt *L. lactis* strain, generation of the recombinant IL-12 producing strain, levels and regulation of IL-12 secretion and its biological activity were described by us in detail previously (12). Strains were grown to an OD₆₀₀ of approx 1.0. Cell pellets were collected, washed with PBS and resuspended in 2% NaHCO₃. A suspension of 100 μ l containing 10⁹ colony-forming units/ml was administered by oral gavage in a single dose to the mice 24 and 2 h before administration of the sensitizing mixture. The percentage of mice that developed Type I hypersensitivity reaction was assessed as previously described (10). Serum levels of peanut-specific IgE antibody were determined by ELISA in all groups of sensitized and control mice as described previously in detail (18). Plasma levels of histamine were detected by using an enzyme immunoassay kit (Immunotech S.A.S., Marseille, France) whereas the vascular leakage was monitored by measuring the hematocrit. Body temperature was monitored using a rectal probe (Kent Scient. Corp. Torrington, USA). Levels of the cytokines IL-4 and IL-5 produced by splenocytes *in vitro* following antigen (CPE 20 μ g/ml) recall challenge (72 h); plasma levels of IFN γ were determined by ELISA kits (ELISA MAXTM Deluxe Sets, Biologend-Cambridge Bioscience, Cambridge, UK).

Gene Expression

Total RNA was extracted from gut tissues using RNeasy Mini Kit (Qiagen Ltd.). For cDNA synthesis, total RNA was reverse transcribed with Quantitect Reverse Transcription Kit (Qiagen Ltd). Q-PCR was performed on 5 μ l of diluted (1:5) template cDNA, using the ABI 7500 Real-Time PCR System (Applied Biosystems, Warrington, UK) and SYBR Green Detection. Data were normalized against an invariant endogenous control, 18S ribosomal RNA, and threshold cycle number (Ct) values obtained were converted into fold of relative induction using the $\Delta\Delta$ Ct. The following primers were used: 18s: F CGG ACA GGA TTG ACA GAT TG-3', R 5'-CAA ATC GCT CCA CCA ACT AA; OX40L: F: CCC TCC AAT CCA AAG ACT; R: ATC CTT CGA CCA TCG TTC; TSLP: F:CCG ATG GGG CTA ACT TAC; R: TCC TGA TTT GCT CGA ACT; IL-33: F: GAT GGG AAG AAG CTG ATG GTG; R: TTT TGA AGG ACG AAG AAG.

Immunohistochemistry and Transmission Electron Microscopy (TEM)

Frozen sections (8–10 μ m) were fixed with 10% buffered formalin for 30 min and endogenous peroxidases were quenched with 0.3% hydrogen peroxide for 1 h and stained with biotinylated goat anti-mouse TSLP (R&D Systems) and then treated with Vectastain Elite ABC kit (Vector Laboratories). Fluorescein deposit was achieved on the site of the immunoreaction with the TSA Plus fluorescence system (Perkin-Elmer). Sections were counterstained with mouse anti- β -actin Mab (Sigma-Aldrich, Milan) followed by TRITC-conjugated donkey anti-mouse IgG antibody (Jackson ImmunoResearch Lab-Starfish, Milan, Italy). For TEM analysis, tissues were fixed in 4% paraformaldehyde and, following dehydration, embedded in Epon 812. Sections were incubated overnight at 4°C with rabbit anti-IL12R β 2 (Bioss Inc., Boston, MA, USA) followed by 15 nm gold-conjugated anti-rabbit IgG (BB International, Cardiff, UK) and examined with a 201 Philips TE microscope.

Isolation of IECs From Mouse Small Intestine and Evaluation of IL-12R Expression and Functional Integrity

Mice were sacrificed; small intestines were removed, dissected longitudinally and washed with sterile PBS. Tissue was cut into 0.5 cm pieces and washed with sterile PBS. To remove IECs, the tissue was subjected to a first wash in PBS containing 10% FCS, 1 mM DTT (20 ml), and a second wash in PBS containing 10% FCS, 15 mM EDTA (20 ml) at 37°C, 250 rpm for 15 and 20 min, respectively. Preceding each wash, tissue-buffer mix was vortexed for 1 min. After washings the resultant pellets were resuspended in PBS containing 10% FCS and stored on ice. CD45⁺ cells were removed from crude IEC preparations using magnetic labeling with Anti-PE Microbeads (Miltenyi Biotec) and magnetic separation with LD MidMACS depletion columns (Miltenyi Biotec). The expression and function of IL-12R β 1 and IL-12 β 2 chains on viable IECs was assessed by flow cytometry. Briefly, isolated IECs were double-stained with anti-IL-12R β 1-FITC and anti-IL-12R β 2-PE or mouse IgG1-PE (R&D Systems) and mouse IgG1-FITC (eBiosciences, UK) isotype controls. After incubation for 1 h on ice, cells

were washed once and pellets were resuspended and fixed with an equal volume of 4% formalin for 20 min at room temperature. Fixed cells were washed once with PBSA. The functional integrity of IL-12R β 2 through MAPKp38 phosphorylation was tested as follows. Freshly isolated IECs were placed in a 37°C water bath in the presence of 500 μ M H₂O₂ or 2 μ g/ml IL-12 (Biolegend, UK) for various times. Control cells were left untreated. At the end of the incubation period, an equal volume of 8% buffered formalin was added. After 10 min, fixed cells were washed with PBSA and allowed to cool on ice; cells were then permeabilized on ice for at least 30 min with Perm Buffer III (BD Biosciences, UK). Cells were washed and stained for 30 min at room temperature with rabbit antiphospho-MAPKp38 (Cell Signaling Technology, Danvers, MA, US) or normal rabbit IgG (Senta Cruz Biotechnology, Dallas, TX, US) as control. Cells were then incubated for a further 30 min with donkey anti-rabbit IgG-FITC (Santa Cruz). For viability assessment cells were washed and resuspended in 1 ml PBS. Dead cells were stained using Live/Dead Fixable Violet Dead Cell Stain Kit (Life Technologies, UK) according to the manufacturer's instructions and data were acquired by flow cytometry.

Intestinal Organoids and Cell Culture

Small intestine organoids were produced according to Sato et al. (19). Crypts were counted and diluted to obtain a final concentration of 75,000 crypts/ml. Pelleted crypts were resuspended in Matrigel matrix (Corning) (1 ml/75,000 crypts) and 20 μ l were seeded per well of a preheated (37°C) 48-well tissue culture plate (Greiner Bio One). Organoids were grown at 37°C and 5% CO₂ and media was replaced daily. The media used were as follows. Small intestinal culture condition (ENR) (19) medium: 18.4 ml advanced DMEM/F12⁺⁺⁺, 0.02% (vol/vol) B27 supplement (Invitrogen), 0.01% (vol/vol) N2 supplement (Invitrogen), n-acetylcysteine (final conc. 1.25 mM, Sigma-Aldrich), recombinant mouse epidermal growth factor (final conc. 50 ng/ml, Invitrogen), recombinant mouse Noggin (final conc. 4 ng/ml, Peprotech), recombinant mouse R-spondin (final conc. 1 ng/ml, R&D systems) and ROCK kinase inhibitor, γ -27632 (final conc. 10 μ M, Sigma-Aldrich). Advanced DMEM/F12⁺⁺⁺ media: 500 ml advanced DMEM/F12 (Invitrogen), 1% (vol/vol) Glutamax 100 \times (Invitrogen), HEPES (final conc. 5 mM, Invitrogen) and penicillin (final conc. 100 U/ml)/streptomycin (final conc. 100 μ g/ml, Invitrogen). In some experiments, conditioned medium (CM) from 3-day culture of Peyer's Patch (PP)-derived immune cells was used as part of the culture medium for the development of the organoids. Passaging was done after 7 days by breaking the Matrigel matrix dome with a 1 ml pipette. The organoids were dissociated from the matrix dome by pipetting with a 200 μ l pipette. Organoids were centrifuged (600 rcf for 5 min), resuspended into fresh Matrigel (ratio 1:3) and seeded as described above. MODE-K cells (20) were used to study the influence of immune derived-signals on the expression of IL-12R β 2 on IECs. MODE-K cells were seeded in the top compartment of Transwells culture system (Costar) at 1×10^5 cells/well. After 4 days of growth PP-derived lymphocytes were added to the culture. Lymphocytes were isolated from PPs by mashing between frost edged sterile glass slides and filtered through strainer. Following the scraping of the MODE-K monolayer, PP-lymphocytes and MODE-K cells

were seeded together to the top compartments of the Transwells to enable cell-to-cell contact. For non-contact coculture, 1×10^6 PP-lymphocytes were added to the bottom compartments of the Transwell. Expression of IL-12R β 2 was examined by flow cytometry after 72 h.

Statistical Analysis

The animal study data were analyzed by (ANOVA) Wilcoxon rank test. The results for the gene expression data show the average relative fold of induction of IL-12R β 2, TSLP, OX40L and IL-33. Data were analyzed using a Student's *t*-test (two-tailed, equal variance) followed by application of the Bonferroni correction the level of which was set at $p \leq 0.0125$. In either case, *p*-value of ≤ 0.05 was considered statistically significant. The results are expressed as mean \pm SD.

RESULTS

IECs Express the IL-12R β 2 Chain but Not the Complementary IL-12R β 1 Chain

The T_H1 cytokine IL-12 binds to a heterodimeric IL-12R β 1/ β 2 complexes on a variety of immune cells (21). First, flow cytometry analysis was carried out using epithelial cell suspension to identify intestinal cells expressing IL-12R. We observed that approximately 58% of small intestine CD45⁻ cells and in a small proportion (>1%) of CD45⁺ cells (Figure 1A) expressed IL-12R β 2 chain. IL-12R β 2 positive cells that did not express CD45 (IL-12R β 2⁺CD45⁻) were identified and selected. The epithelial origin of the IL-12R β 2⁺CD45⁻ cells was confirmed through expression of cytokeratin as seen in Figure 1B. Figure 1C demonstrated that these IL-12R β 2⁺CD45⁻cytokeratin⁺ cells did not coexpress IL-12R β 1. Side scatter (SSC) analysis is also shown (Figures 1D,E). This showed that the IL-12 β 2 chain is expressed *in vivo* by IECs in absence of the complementary IL-12R β 1 chain.

It has been shown that the presence of the IL-12R β 2 chain is indispensable for IL-12 mediated signaling (15) although IL-12R β 2 alone was insufficient to induce the canonical signal transducer and activator of transcription (Stat) 4 activation in response to IL-12 (22). However, an IL-12R β 1/STAT4-independent pathway and based on MAP kinase (MAPK) activation in the presence of only the IL-12R β 2 chain was observed (22). Thus, to assess the function of IL-12R β 2 in the absence of the complementary IL-12R β 1 chain, freshly isolated IECs were cultured in the presence of IL-12p70 and levels of phospho-MAPK38 were assessed. Rapidly, after the application of the stimulus, expression of phospho-MAPKp38 reached similar levels observed following challenge of IECs with hydrogen peroxide (H₂O₂) a non-specific activation stimulus (Figure 1F). By contrast, IECs isolated from IL-12R β 2^{-/-} mice did not respond to challenge with IL-12p70 demonstrating that activation of MAPK38 was an IL-12R β 2 specific event. The expression of the IL-12R β 2 chain on IECs was further confirmed by TEM by immunogold staining. Positive staining was observed within the Golgi area (Figure 2A), in the proximity of the basolateral membrane (Figure 2B) and, most relevant to lympho-epithelial cross talk, at the interface between IECs and immune cells harboring the lamina propria (lp) (Figure 2C).

The Expression of IL-12R β 2 Chain on IECs Is Controlled by Immune System-Derived Signals

The *in vivo* observation that IL-12R β 2 was expressed by IECs at the interface with local lp-immune cells prompted us to assess whether its expression was under the control of immune-derived signals. Quantitative RT-PCR analysis showed that the IL-12R β 2 was expressed on freshly isolated primary IECs; however, its expression was negligible on the murine IEC line MODE-K (Figure 3A). By contrast, when using coculture systems (Figure 3B) we observed that PP-lymphocytes seeded in the bottom compartment of Transwells culture system induced the expression of the IL-12R β 2 in MODE-K (1.5-fold increase). Yet, a more pronounced increase (3.2-fold) was seen when PP-lymphocytes were allowed cell-to-cell interaction with the MODE-K cells. The importance of immune-derived signals was further investigated in primary small intestine organoids that were developed *in vitro*. In the complete absence of immune cell-derived signals organoids IECs lacked the expression of the IL-12R β 2 chain, however, when the organoids were grown in the presence of PP-derived conditioned medium (PP-CM) significantly higher number of IL-12R β 2⁺ cells were observed (Figure 3C; Figure S1 in Supplementary Material). This experimental approach also showed that in the presence of PP-CM intestinal organoids also tend to increase the expression of cytokeratin (Figure S1 in Supplementary Material). Taken together these data demonstrated that cross-talk between IECs and immune cells is instrumental to trigger the expression of the IL-12R β 2 on IECs.

IEC-Associated IL-12R β 2 Chain Plays a Role in Lympho-Epithelial Cross Talk

To test the biological role of IEC-associated IL-12R β 2 chain, we analyzed the ability of a recombinant bacterial vector secreting bioactive IL-12p70 (rLc-IL12) (12) to suppressing food allergic response in wt and chimeric IL-12R β 2^{-/-} and IL-12R β 2^{ΔIEC} mice that lack IL-12R β 2 on IECs. First, we assessed the production of IFN γ in response to systemic administration of IL-12 in IL-12R β 2^{-/-} mice following BM transplant from wt mice (WT \rightarrow IL-12R β 2^{-/-}) at week 8 posttransplant. (Figure 4) In agreement with a previous report (15), the presence of an intact IL-12R enabled wt mice to produce *in vivo* significant amount of IFN γ following i.p. administration of IL-12. By contrast, IL-12R β 2^{-/-} and chimeric IL-12R β 2^{-/-} (IL-12R β 2^{-/-} \rightarrow IL-12R β 2^{-/-}) mice did not produce detectable levels of IFN γ . However, IFN γ production was completely restored in IL-12R β 2^{-/-} mice by BM transplant from wt mice (WT \rightarrow IL-12R β 2^{-/-}). The result of the functional assay was paralleled by the observation that post-BM transplant mice displayed similar levels of MHCII⁺ cells in the lp compared to wt mice (Figure S2 in Supplementary Material). Similar results were obtained at week 28 posttransplant (not shown). Taken together these data show a successful BM engraftment. Subsequently we investigated the role of IEC-associated IL-12R β 2 in the regulation of immune response using a well-established mouse model of food allergy. The IL-12 secreting bacterial vector rLc-IL12 was delivered prior to each administration of the sensitizing mixture and its effects on food

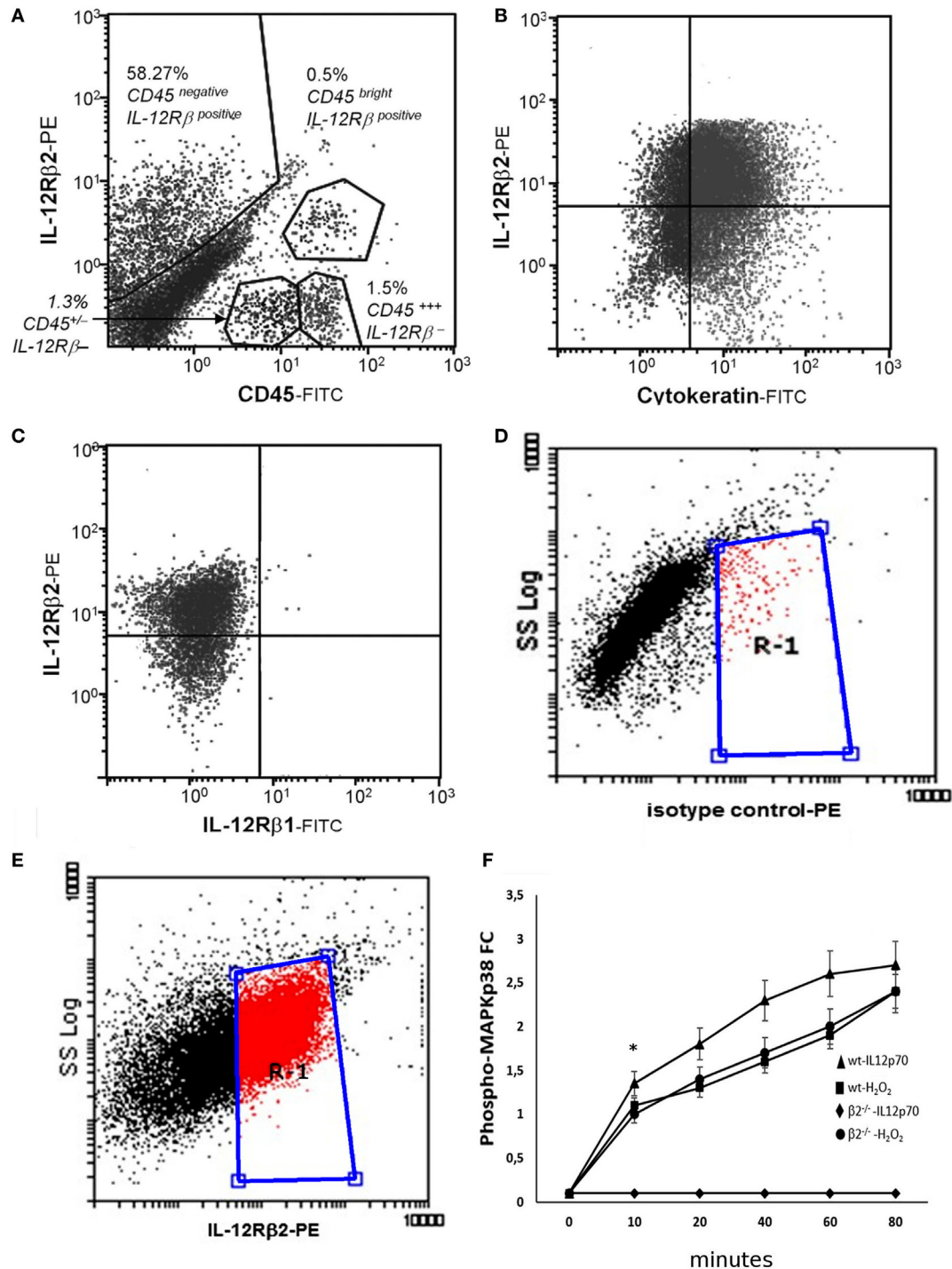


FIGURE 1 | Expression and functional integrity of IL-12R β 2 on intestinal epithelial cells (IECs). Flow cytometry analysis was carried out using epithelial cell suspension to identify intestinal cells expressing IL-12R. IL-12R β 2 chain was detected on approximately 50% of small intestine CD45⁺ (IECs) cells and in a small proportion (>1%) of CD45⁺ cells (A). IL-12R β 2 positive cells that did not express CD45 (IL-12R β 2⁺CD45⁻) were identified and selected. Their epithelial origin was confirmed through expression of cytokeratin (B). Furthermore, we showed that the expression of IL-12R β 2 on these cells was not paralleled by the presence of the complementary IL-12R β 1 chain (C). Also, plots of SSC vs. IL-12R β 2 are shown (D,E); Cells were stained with PE-coupled isotype control antibody to facilitate gating and enable detection of non-specific fluorescence (D). Following staining with PE-coupled anti-IL-12R β 2 antibody, positive cells were identified (E) within the gate R-1 constructed in D above. Finally, *ex vivo* stimulation of freshly isolated IECs from the mouse small intestine with IL-12p70 led to a rapid increase in intracellular phosphorylation of MAPKp38 only in IEC from wt mice but not IL-12R β 2^{-/-} mice showing that the upregulation of phospho-MAPK38 is an IL-12R β 2-specific event; stimulation with H₂O₂ was used as a positive control (F). Experiments on MAPKp38 were performed in triplicate using freshly isolated IECs (five mice/group). (*) indicates statistical difference. $p < 0.05$.

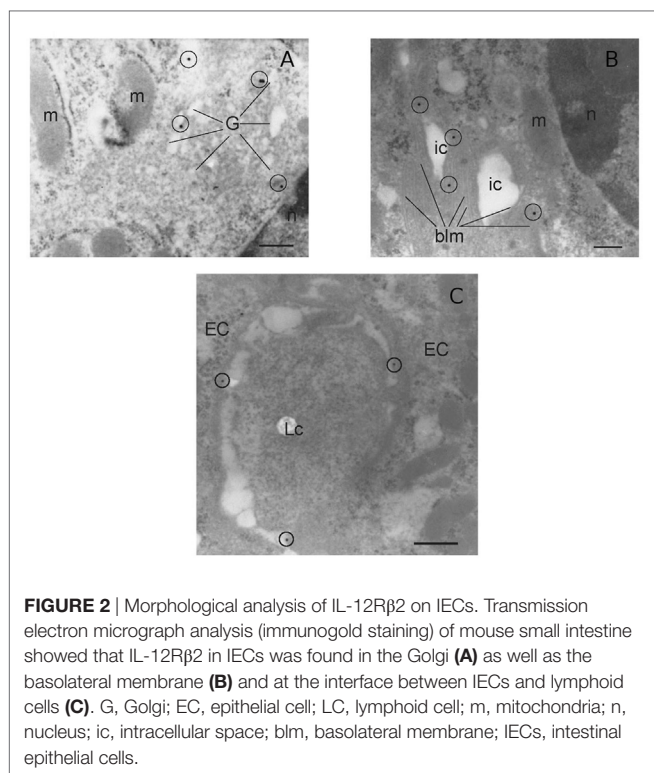


FIGURE 2 | Morphological analysis of IL-12R β 2 on IECs. Transmission electron micrograph analysis (immunogold staining) of mouse small intestine showed that IL-12R β 2 in IECs was found in the Golgi (A) as well as the basolateral membrane (B) and at the interface between IECs and lymphoid cells (C). G, Golgi; EC, epithelial cell; LC, lymphoid cell; m, mitochondria; n, nucleus; ic, intracellular space; blm, basolateral membrane; IECs, intestinal epithelial cells.

allergic reaction were monitored upon final allergen challenge by monitoring a panel of allergic symptoms. Oral delivery of rLc-IL12 significantly reduced plasma levels of IgE in sensitized wt mice compared to sensitized untreated mice (Figure 5A). Also, in these mice we observed that rLc-IL12 prevented the drop in temperature associated with allergic reaction following the final challenge with the causal allergen (Figure 5B). By contrast, IgE response was not affected by the administration of rLc-IL12 in chimeric IL-12R β 2^{-/-} mice (Figure 5C); also, these mice showed a markedly higher production of IgE compared to wt throughout the allergen sensitization period (summary in Table 1). In this group of mice, rLc-IL12 also failed to prevent the allergy-associated drop in body temperature (Figure 5D). The pattern in IL-12R β 2^{ΔIEC} appeared to be different. In this group, the administration of rLc-IL12 significantly reduced plasma levels of IgE but the magnitude of the reduction was smaller compared to wt mice (41 and 93%, respectively) (Figure 5E). A similar profile was observed in regard to the effects of rLc-IL12 on body temperature (Figure 5F). Here too the treatment with rLc-IL12 significantly prevented the drop of body temperature observed in wt mice but not as much as observed in the wt cohort. A similar trend was observed for all the other parameters of Type I allergic reactions monitored (Figure 6). Plasma levels of histamine in sensitized wt mice reached 2,100 ± 185 ng/ml and were reduced to 320 ± 21 ng/ml ($p < 0.01$) in rLc-IL12-treated mice leading to a 75% reduction (Figure 6A). Furthermore, while no effects were seen on histamine levels in IL-12R β 2^{-/-} mice, the administration of rLc-IL12 led to a reduction of plasma levels in IL-12R β 2^{ΔIEC} from 2,600 ± 212 to 1,450 ± 104 ng ml

totaling to a 45% reduction. Treatment with rLc-IL12 also affected IL-4 production following *in vitro* recall antigen challenge of splenocytes in both wt and chimeric IL-12R β 2^{ΔIEC} but not in chimeric IL-12R β 2^{-/-} mice. Levels of IL-4 in culture supernatants following splenocytes challenge with the causal allergen were significantly reduced ($p < 0.01$) by treatment with rLc-IL12 (Figure 6B). By contrast, production of IL-5 was downregulated by rLc-IL12 only in wt mice but not in the two chimeric groups (Figure 6C). Hematocrit was utilized to monitor vascular permeability leakage, an additional feature of type I hypersensitivity reaction (Figure 6D). Hematocrit levels were increased in sensitized untreated wt mice (60.5%) compared to baseline levels (41.5%) and, in these mice was significantly reduced by rLc-IL12 (43.5%). Administration of rLc-IL12 did not suppress vascular leakage in chimeric IL-12R β 2^{-/-} mice, however, rLc-IL12 treatment reduced hematocrit levels in chimeric IL-12R β 2^{ΔIEC} mice compared to untreated syngeneic mice (42.2% and 61.2%, respectively). Finally, we observed that the expression of the IL-12R β 2 on IECs did not differ between sensitized and control naïve wt mice (Figures 6E,F) suggesting that allergy-associated skewed T_H1/T_H2 ratio did not affect the expression of the IL-12R β 2 chain on IECs.

Downregulation of IEC-Derived TSLP Following Oral Delivery of rLc-IL12 Did Not Occur Directly via IEC-Associated IL-12R β 2

The overexpression of the IEC-derived cytokines TSLP and IL-33 is critical in food allergy (23–25). We then tested the hypothesis that the engagement of IL-12 with the functional IEC-associated IL12R β 2 chain could play a role in the regulation of these pro-allergic cytokines. First, quantitative RT-PCR analysis (Figure 7A) showed that TSLP gene expression was significantly upregulated in the gut epithelium of sensitized mice as well as in mice treated with CT alone, compared to control non-sensitized (PBS-treated) mice. Furthermore, administration of rLc-IL12 led to a significant downregulation of TSLP gene expression in the gut epithelium of sensitized mice. The regulatory effect of rLc-IL12 on IEC-derived TSLP was further confirmed by immunohistochemistry. The latter approach did not detect TSLP neither in PBS- nor in rLc-IL12-treated mice (Figures 7B,C) but only in sensitized mice (Figures 7D,E). Taken together, these results concurred with reports that suggested a critical role of the epithelium-derived TSLP in allergic reactions (23, 24) and pointed to an important role of IL-12 in the regulation of the epithelium-derived TSLP. Furthermore, immunofluorescence staining was carried out in both IL-12R β 2^{-/-} and IL-12R β 2^{ΔIEC} chimeric mice undergoing allergic sensitization and treated with rLc-IL12. Both sensitized chimeric mice retained the ability to produce TSLP by IECs (Figure S3 in Supplementary material). The latter observation further suggested that IEC-associated IL-12R β 2 plays a role in the control of TSLP production by IECs. However, when the expression of TSLP was assessed in the epithelium of naïve IL-12R β 2^{-/-} mice we did not observe an increase in the level of TSLP compared to naïve wt counterparts (Figure 7F). Taken together these data point to a scenario suggesting that IEC-associated IL-12R β 2 is an important, but

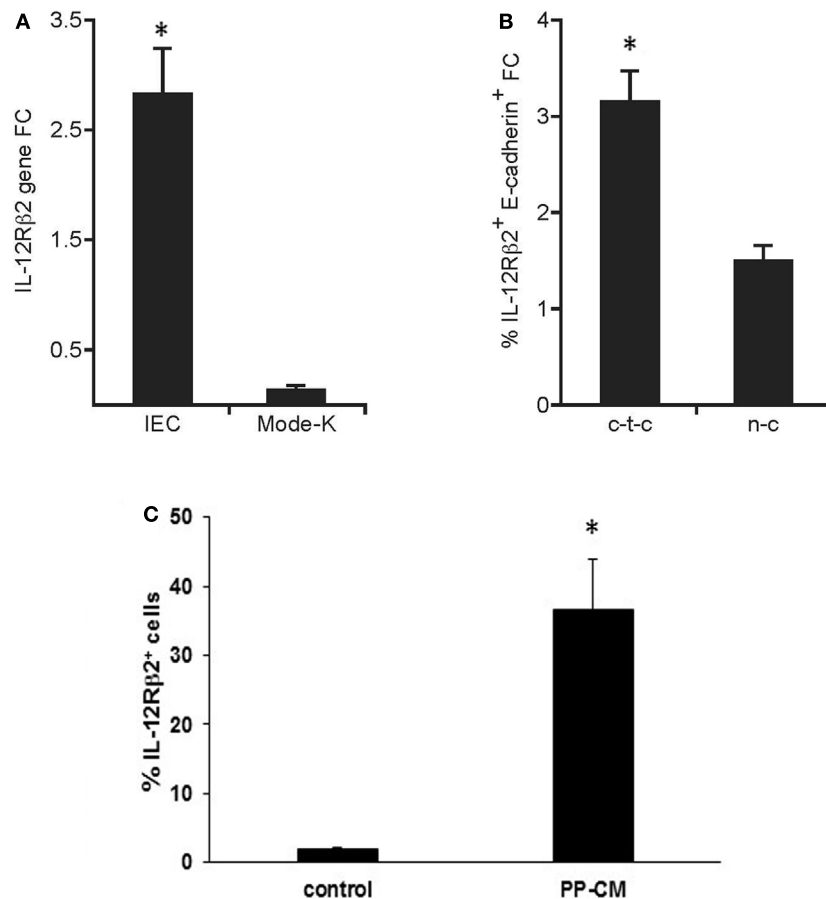
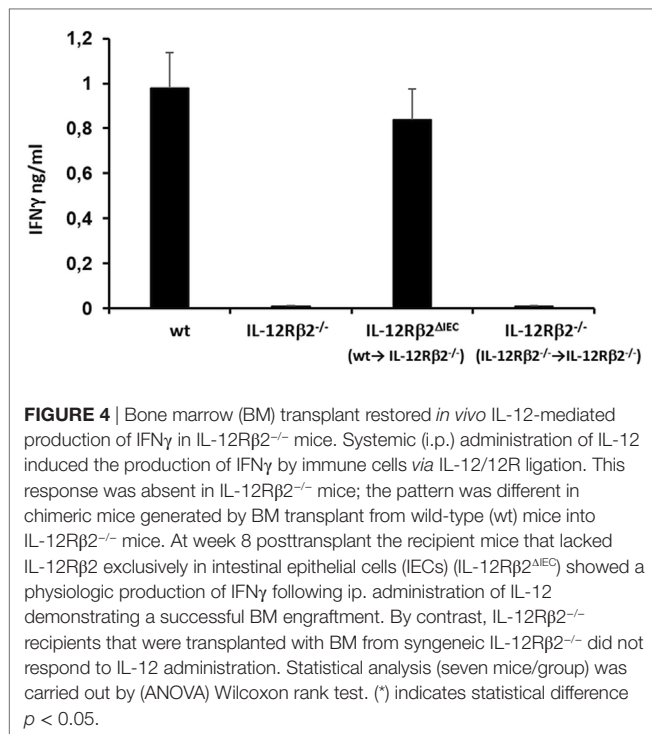


FIGURE 3 | Immune system-derived signal(s) are required for the expression of functional IL-12R β 2 on intestinal epithelial cells (IECs). Quantitative RT-PCR analysis showed that the expression of IL-12R β 2 was significantly higher on freshly isolated primary IECs compared to cultured intestinal epithelial cell line MODE-K (A). By contrast (B), higher expression of IL-12R β 2 (1.5-fold change, FC, increase compared IL12R β 2 expression levels in MODE-K untreated control cells) was seen when Peyer's patch-derived lymphocytes were seeded in the bottom compartment of the Transwells culture system not in contact with MODE-K cells (n-c). However, a more pronounced increase (3.2-fold) was seen when PP-lymphocytes were allowed cell-to-cell interaction with the MODE-K cells (c-t-c). Expression of IL-12R β 2 was also assessed on primary intestinal organoids developed in culture in the absence (control) or in presence of conditioned medium (CM) from 3-day cultures of PP-derived immune cells (PP-CM) chain (C). A significant increase in the number of organoid cells expressed the IL-12R β 2 was seen when the organoids were grown in the presence of CM. Transcript analysis was carried out in replicate mice (five mice/group) in two independent assays. Data were analyzed using a Student's *t*-test (two-tailed, equal variance) followed by application of the Bonferroni correction the level of which was set at $p \leq 0.0125$. Mode-K cells were pooled in two independent experiments and samples run in triplicate. Two batches of intestinal organoids/treatment were prepared and samples were run in triplicate. The results are expressed as mean \pm SD. (*) indicates statistical difference, $p < 0.05$.

not the only factor controlling TSLP production by IECs. Overall, taken together these data demonstrated that the downregulation of IEC-derived TSLP following oral administration of IL-12 is not directly linked to IL-12 engagement with the IL-12R β 2 chain on IECs, thus suggesting that IL-12 might trigger the production of additional factor(s) that in turn controlled the production of TSLP by the intestinal epithelium. Finally, as a corollary parallel analysis showed that administration of rLc-IL12 suppressed allergic reaction without downregulating levels of IEC-derived IL-33 while at the same time partially suppressing allergy-associated expression of OX40L (Figure S4 in Supplementary Material). The latter observation pointed to a synergistic action of TSLP and IL-33 on the expression of the allergy-critical OX40L rather than an exclusive role of IL-33 as previously suggested (25).

DISCUSSION

Here, we report that mouse IECs expressed the IL-12R β 2 but not the complementary IL-12R β 1 chain that functions as an incomplete, yet functional IL-12R that plays a role in lympho-epithelial cross talk. In the mouse system, the two β 1 and β 2 components of the IL-12R have specific role in IL-12-mediated binding and signaling. IL-12R β 1 chain is the primary binding site conferring both high and low affinity, instead the IL-12R β 2 only binds IL-12 with low affinity (26). However, these studies carried out in IL-12R β 1^{-/-} mice initially suggested that the β 1 component was required for both high affinity binding and for immune cells response to IL-12 it was later showed that only the IL-12R β 2 chain was strictly required for IL-12 signaling (15). Indeed, no IL-12



mediated biological functions could be detected in IL-12R β 1⁺/ β 2⁻ mice. Combined, these studies suggested that in the mouse system the IL-12R β 1 is primarily responsible for binding and IL-12R β 2 is further required for IL-12 signaling. Our study extends this concept showing that while this is the case for immune cells the expression of the sole β 2 chain of the IL-12R on IECs might be an important component of the lymphoepithelial cross talk.

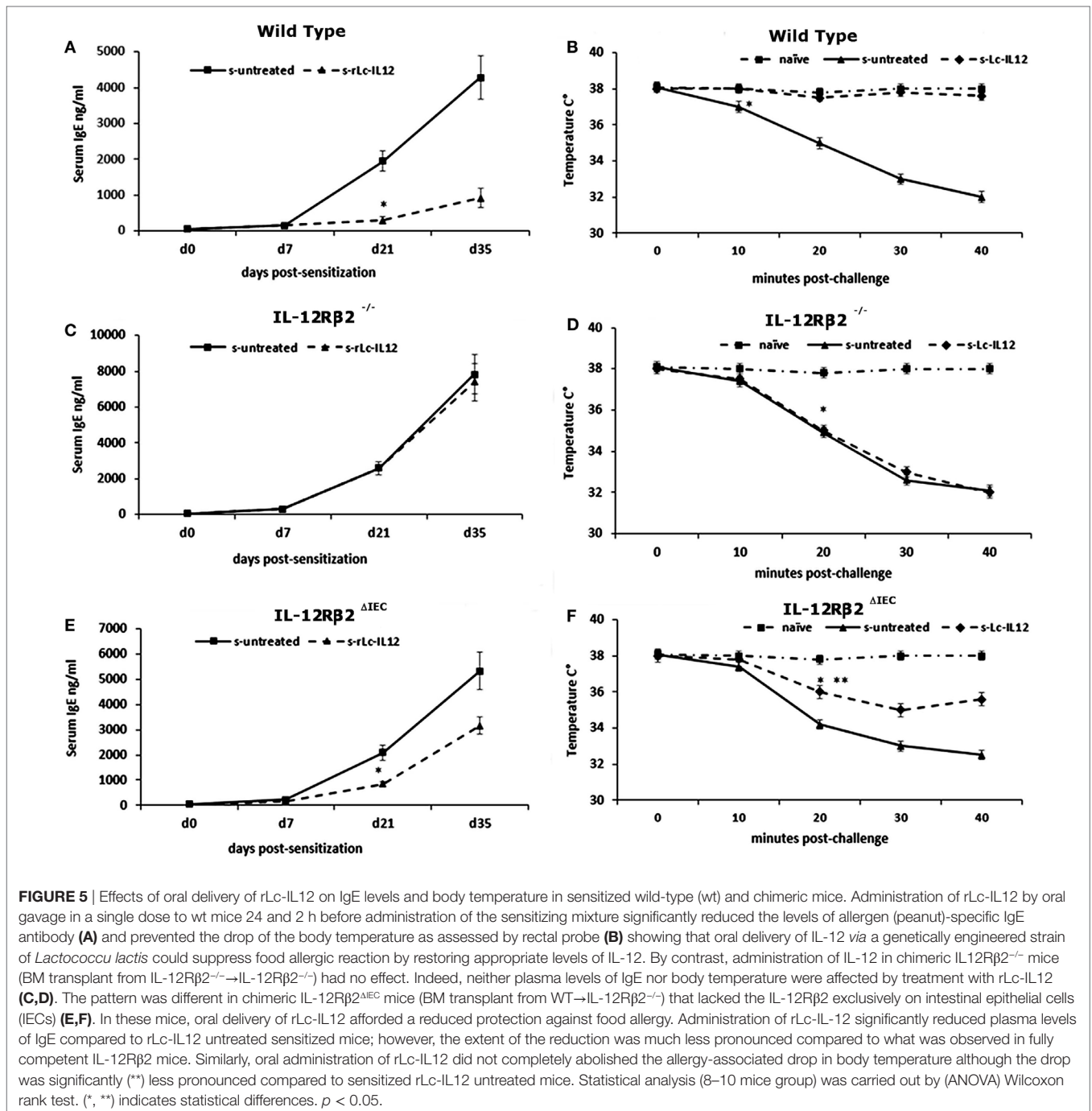
The expression of IL-12R β 2 on IECs depended on immune-derived stimuli. This concept stemmed from several observations. A mouse small-intestine derived epithelial cell line and primary intestinal organoids cultured in the absence of gut-derived immune cells did not express the IL-12R β 2 chain. By contrast, the presence of gut-derived immune cells triggered the expression of the IL-12R β 2 subunit by both an IEC line and primary intestinal organoids. This is in line with previous reports showing that the expression of the IL-12R β 2 but not of the IL-12R β 1 is regulated by signals (i.e., cytokines) originating from the immune system (22). Although the nature of the signals remains to be determined it appeared that both soluble factor(s) and cell-to-cell interaction are important to trigger the expression of IL-12R β 2. This was also suggested by the observation that *in vivo*, the expression of the IL-12R β 2 chain was located at the basolateral domain of IECs at the interface with local Ip-immune cells.

IL-12 ligation with the conventional IL-12R β 1/ β 2 complex signaled through the Janus kinase (JAK)-signal transduce pathway in both human and mouse (27, 28); in particular, the critical role of Stat-4 in IL-12R signaling has been demonstrated *in vivo* *via* the use of Stat 4-deficient mice (29). On the other hand, although in IL-12R β 1-deficient humans the presence of the sole IL-12R β 2 chain was not sufficient to confer full IL-12 responsiveness *via* JAK-mediated pathway, T cells from these individuals could still

develop into T_H1 cells and respond to IL-12 (22). In this case, IL-12 signaling occurred *via* an IL-12R β 1-independent pathway that involved MAPK pathways. In agreement with these observations, we reported that IL-12/IL-12R β 2 ligation on freshly isolated IL-12R β 1⁻/ β 2⁺ IECs induced activation of phospho-MAPKp38 in a manner similar to that obtained by a non-specific MAPK activator. At present the possibility that other surface molecules might be associated with IL-12R β 2 on IECs to form an additional variant of the IL-12R that operates *via* MAPK cannot be ruled out; however, our results, strongly suggested that the presence of IL-12R β 2 alone on IECs is sufficient to confer responsiveness to IL-12.

The presence of a functional, albeit incomplete IL-12R on IECs suggested that the IL-12/IL-12R β 2 ligation could be important in controlling the production by IEC-derived cytokines. We tested this hypothesis in a mouse model of food allergy. Its utilization was prompted by the notions that in this model the allergic sensitization to food components (peanuts) was characterized by the lack of IL-12 in the gut but not in the systemic immune system (10); in addition, using the same model others have previously shown that oral delivery of liposome encapsulated recombinant IL-12 suppressed allergic reaction (11). We report here that in chimeric mice the lack of IEC-associated IL-12R β 2 significantly reduced but not completely abolished the antiallergic capability of a *L. lactis* strain genetically engineered to secrete bioactive IL-12. This result suggested that IL-12 delivered orally *via* the recombinant bacterial vector produced the antiallergic activity by engaging a variety of IL-12 responding cells either locally or systemically that included IECs. This result could not be attributed to an unsuccessful BM engraftment in IL-12R β 2^{-/-} recipients; in these mice the production of IFN γ following systemic administration of IL-12 was completely restored at 8 weeks after transplant of BM from wt mice.

Oral delivery of Lc-IL12 had a significant impact on the levels of TSLP produced by IECs in sensitized mice. The production of TSLP by IECs is an important example of the importance of the tight regulation of the finely tuned cytokine network in the gut. On one hand, TSLP plays a critical role in maintaining the intestinal immune homeostasis by shaping the anti-inflammatory features of dendritic cells (2) and Treg (30); on the other, strong evidence suggested that its over-expression is instrumental for the development of allergic reactions (23, 24). We found, in agreement with another report (25) that mice undergoing the sensitization protocol employed in our study displayed a significant increase of TSLP production by IECs during and following the sensitizing procedure. However, oral administration of rLc-IL12 significantly downregulated TSLP production by IECs while at the same time suppressing allergic symptoms. The action of IL-12 on TSLP production however did not appear to be directly linked to the IL-12/IL-12R ligation on IECs. Naive IL-12R β 2^{-/-} mice showed only a slight increase of IEC-derived TSLP, thus suggesting that the production of TSLP by IECs is under the control of multiple factors. It is known that the production of certain cytokines, such as IL-13 required the simultaneous presence of two cytokines (i.e., IL-33 and IL-7) (31) and this notion lends support to the hypothesis that a dual signal might be required also to control the production of TSLP by IECs. This would suggest that IL-12 mediated regulation of TSLP production required the synergy with factor(s) that

**TABLE 1** | Effects of rLc-IL12 on IgE production (ng/ml): day 35.

Group (n = 8/10) reduction	Control	s-Untreated	s-rLc-IL12	p	%
Group I (wt)	<10	4,280 ± 950	275 ± 72	<0.001	93.6
Group II (IL-12R β 2 ^{-/-})	<10	7,820 ± 1,050	7,250 ± 1,220	>1	8
Group III (IL-12R β 2 ^{ΔIEC})	<10	5,320 ± 980	3,150 ± 650	0.05	41

Levels of plasma IgE were evaluated in wt (Group I) and chimeric mice deficient for IL-12R β 2 chain (Group II, IL-12R β 2^{-/-}) or deficient for IL-12R β 2 chain in intestinal epithelial cells (Group III, IL-12R β 2^{ΔIEC}).

Values refer to levels of IgE prior to the beginning of the sensitization period (control) and in mice sensitized and left untreated (s-untreated) or treated with rLc-IL12 (s-rLc-IL12) on day 35 following the final antigenic challenge.

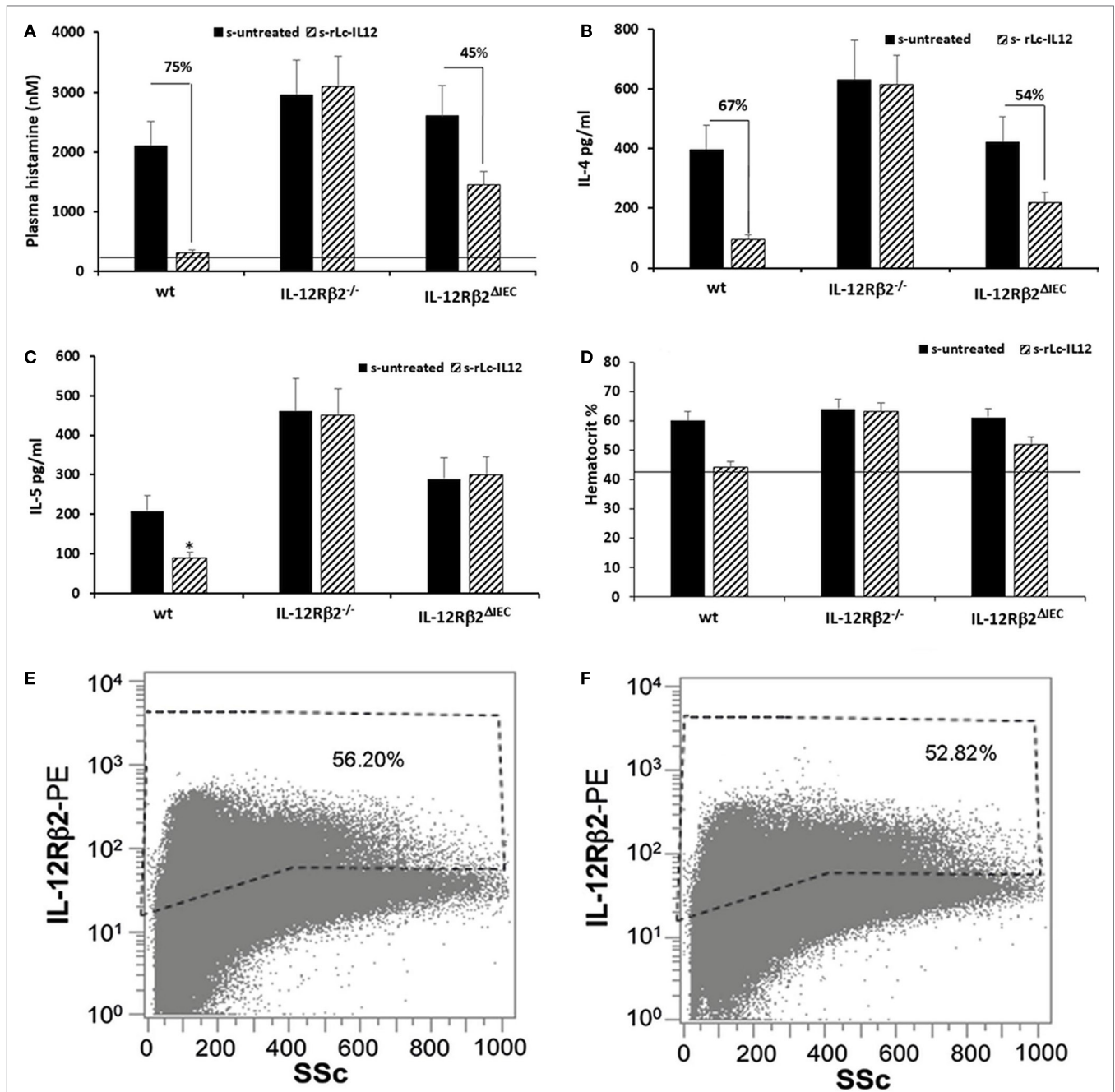


FIGURE 6 | Efficacy of rLc-IL12 on additional parameters of food allergy in sensitized wild-type (wt) and chimeric mice. Oral administration of IL-12 reduced levels of plasma histamine by 75% in sensitized wt mice compared to IL-12 untreated syngeneic mice (A). By contrast, no effects were seen in chimeric IL-12R β 2^{-/-} mice; instead the IL-12 mediated reduction of plasma histamine in chimeric IL-12R β 2 ^{Δ IEC} totaled only to 45%. Also, levels of IL-4 produced by splenocytes following *in vitro* recall allergen challenge showed a similar pattern (B). Production of IL-4 in wt mice treated with rLc-IL12 dropped 67% compared to sensitized untreated mice; the same treatment was not effective in chimeric IL-12R β 2^{-/-} mice and only partially effective in IL-12R β 2 ^{Δ IEC} mice (54%). By contrast, levels of IL-5 following *in vitro* recall allergen challenge were reduced by rLc-IL12 only in wt but not in either chimeric mouse variants (C). Hematocrit was evaluated as a measure of vascular leakage (D). Administration of rLc-IL12 restored near physiological levels of hematocrit in wt mice; no effects were seen in chimeric IL-12R β 2^{-/-} mice and only partial restoration of the hematocrit was seen in chimeric IL-12R β 2 ^{Δ IEC}. Finally, representative histograms showed that levels of IL-12R β 2 on intestinal epithelial cells (IECs) did not vary between sensitized (F) and naïve mice (E). Statistical analysis (8–10 mice group) was carried out by (ANOVA) Wilcoxon rank test. **p* < 0.05.

are suppressed/downregulated in sensitized mice but that are still retained in non-sensitized IL-12R β 2^{-/-} mice. Overall, our study suggested that IL-12 and TSLP are part of an important regulatory

cytokine loop operating at the interface between the intestinal epithelium and the underlying immune system that ultimately plays a role on the host's ability to mount appropriate immune

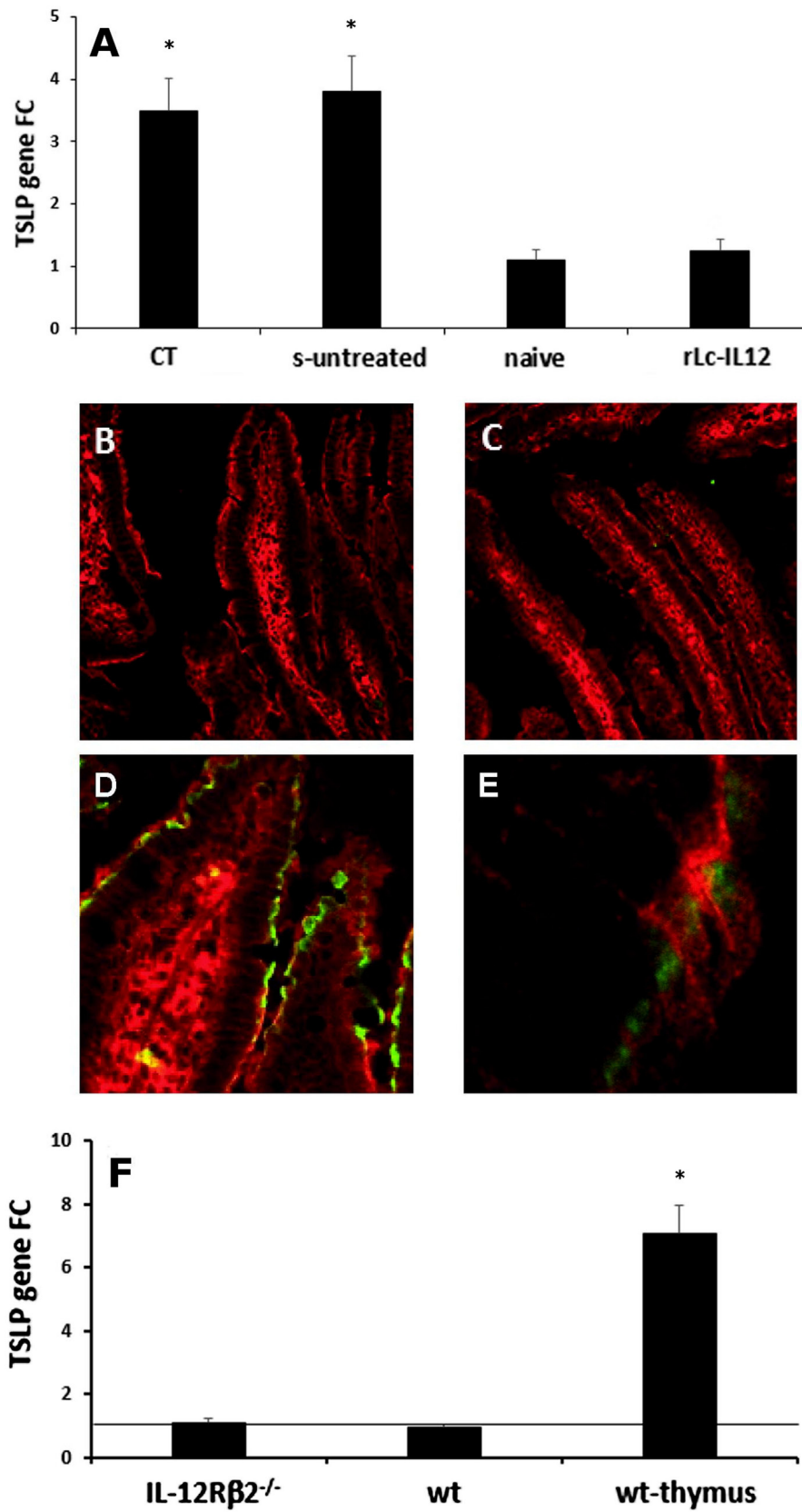


FIGURE 7 | Continued

FIGURE 7 | IL-12-mediated downregulation of IEC-derived TSLP does not occur directly *via* IEC-associated IL-12R β 2. RT-PCR analysis of the gut epithelium (A) showed that oral delivery of the adjuvant cholera toxin (CT) induced a significant increase of TSLP in the gut epithelium similar to that observed following delivery of the complete sensitizing mixture (s-untreated) compared to control mice (naive). Allergy-associated levels of TSLP were abrogated by treatment with rLc-IL12. This notion was further confirmed by immunohistochemistry. Immunofluorescence analysis with anti-TSLP antibody did not detect levels of TSLP in the IECs of both control (B) and rLc-IL12-treated mice (C). In contrast, specific anti-TSLP staining (green) was observed in the epithelium of sensitized rLc-IL-untreated mice (D), detail in (E). Sections were counter-stained with antiactin antibody (red). Finally, expression of TSLP gene in the intestinal epithelium of naïve IL-12R β 2^{-/-} mice (F) showed only a minor increase that did not reach statistical significance compared to wt syngeneic mice. In this case, the thymus from wt mice was used as positive control. Transcript analysis was carried out in replicate mice (four to six mice/group) in two independent assays. Data were analyzed using a Student's *t*-test followed by application of the Bonferroni correction the level of which was set at $p \leq 0.0125$. (*) indicate statistical difference. $p < 0.05$.

responses. Indeed, it has been shown previously that TSLP directly suppressed the production of IL-12 in DCs (32); here, we report that in turn IL-12, in combination with other factors plays a role in controlling the production of TSLP by IECs.

The utilization of a mouse model of food allergy to understand the biological role of IL-12R β 2 in controlling the production of IEC-derived cytokines led to the parallel finding that the suppression of food allergy by IL-12 was indirectly linked to the downregulation of TSLP but did not affect levels of another allergy-critical epithelium-derived cytokine, IL-33. Instead, at the same time oral delivery of IL-12 partially suppressed allergy-associated expression of OX40L. This was rather surprising considering that IL-33 but not TSLP was previously indicated as having a critical, non-redundant role in allergy (25). Thus, our work in agreement with a recent report (33) argued against the concept that the alteration of a single immunoregulatory pathway (i.e., IL-33) is at the core of allergic reactions. Our data point to a synergistic action of TSLP and IL-33 on OX40L expression, as also more recently shown in a rhinovirus-mediated allergic reaction to aeroallergen (34), rather than an exclusive IL-33-mediated effect as previously suggested (25). In summary, taken together these data show that IL-12R β 2 chain functions as an incomplete yet functional IL-12R on IECs and overall further highlight the importance of a finely tuned lymphoepithelial cross talk in the intestine in maintaining health.

ETHICS STATEMENT

Experiments were approved by the University of East Anglia Animal Welfare and Ethical Review Body (AWERB; ref: 70/7583)

REFERENCES

- Neutra MR, Mantis NJ, Kraehenbuhl JP. Collaboration of epithelial cells with organized mucosal lymphoid tissues. *Nat Immunol* (2001) 2:1004–9. doi:10.1038/ni1101-1004
- Rimoldi M, Chieppa M, Salucci V, Avogadri F, Sonzogni A, Sampietro GM, et al. Intestinal homeostasis is regulated by the crosstalk between epithelial cells and dendritic cells. *Nat Immunol* (2005) 6:507–14. doi:10.1038/ni1192
- Zeuthen LH, Fink LN, Frokiaer H. Epithelial cells prime the immune response to an array of gut-derived commensals towards a tolerogenic phenotype through distinct actions of thymic stromal lymphopoietin and transforming growth factor beta. *Immunology* (2008) 123:197–208. doi:10.1111/j.1365-2567.2007.02687.x
- Mowat AM. Anatomical basis of tolerance and immunity to intestinal antigens. *Nat Rev Immunol* (2003) 4:331–41. doi:10.1038/nri1057
- Sansonetti PJ. War and peace at mucosal surfaces. *Nat Rev Immunol* (2004) 4:953–64. doi:10.1038/nri1499
- Man AL, Gicheva N, Regoli M, Rowley G, De Cunto G, Wellner N, et al. CX₃CR1⁺ cell-mediated *Salmonella* exclusion protects the intestinal mucosa during the initial stage of infection. *J Immunol* (2017) 198:335–43. doi:10.4049/jimmunol.1502559
- Man AL, Lodi F, Bertelli E, Regoli M, Pin C, Mulholland F, et al. Macrophage migration inhibitory factor plays a role in the regulation of microfold (M) cell-mediated transport in the gut. *J Immunol* (2008) 181:5673–80. doi:10.4049/jimmunol.181.8.5673
- de Jong R, Altare F, Haagen IA, Elferink DG, Boer T, van Breda Vriesman PJ, et al. Severe mycobacterial and *Salmonella* infections in interleukin-12 receptor-deficient patients. *Science* (1998) 280:1435–8. doi:10.1126/science.280.5368.1435
- Peterson LW, Artis D. Intestinal epithelial cells: regulators of barrier function and immune homeostasis. *Nat Rev Immunol* (2014) 14:141–53. doi:10.1038/nri3608
- Temblay JN, Bertelli E, Arques JL, Regoli M, Nicoletti C. Production of IL-12 by Peyer patch-dendritic cells is critical for the resistance to food allergy. *J Allergy Clin Immunol* (2007) 120:659–65. doi:10.1016/j.jaci.2007.04.044
- Lee SY, Huang CK, Zhang TF, Schofield BH, Burks AW, Bannon GA, et al. Oral administration of IL-12 suppresses anaphylactic reactions in a murine model

and conducted under the guidelines of the Scientific Procedure Animal Act (1986) of the United Kingdom. Also, experimental protocols were approved by the Animal Care and Use Committee of the University of Siena under the “Guiding Principles for Research Involving Animals and Human Beings.”

AUTHOR CONTRIBUTIONS

CN, AN, MG, EB and MR designed the study and wrote the manuscript; MR, AM, NG, AD, KI, AP, ML, JJVB, GM and US performed the experiments, provided technical support and carried out data analysis; CN, AN and EB supervised research.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at <https://www.frontiersin.org/articles/10.3389/fimmu.2018.01177/full#supplementary-material>.

- of peanut hypersensitivity. *Clin Immunol* (2001) 101:220–8. doi:10.1006/clim.2001.5122
12. Fernandez A, Horn N, Wegmann U, Nicoletti C, Gasson MJ, Narbad A. Enhanced secretion of biologically active murine interleukin-12 by *Lactococcus lactis*. *Appl Environ Microbiol* (2009) 75:869–71. doi:10.1128/AEM.01728-08
 13. Sorensen EW, Gerber SA, Frelinger JG, Lord EM. IL-12 suppresses vascular endothelial growth factor receptor 3 expression on tumor vessels by two distinct IFN-gamma-dependent mechanisms. *J Immunol* (2010) 184:1858–66. doi:10.4049/jimmunol.0903210
 14. Peake K, Manning J, Lewis CA, Barr C, Rossi F, Krieger C. Busulfan as a myelosuppressive agent for generating stable high-level bone marrow chimerism in mice. *J Vis Exp* (2015) 98:e52553. doi:10.3791/52553
 15. Wu C, Wang X, Gadina M, O'Shea JJ, Presky DH, Magram J. IL-12 receptor beta 2 (IL-12R beta 2)-deficient mice are defective in IL-12-mediated signaling despite the presence of high affinity IL-12 binding sites. *J Immunol* (2000) 165:6221–8. doi:10.4049/jimmunol.165.11.6221
 16. Li XM, Serebrisky D, Lee SY, Huang CK, Bardina L, Schofield BH, et al. A murine model of peanut anaphylaxis: T- and B-cell responses to a major peanut allergen mimic human responses. *J Allergy Clin Immunol* (2000) 106:150–8. doi:10.1067/mai.2000.107395
 17. Chambers SJ, Wickham MSJ, Regoli M, Bertelli E, Gunning PA, Nicoletti C. Rapid in vivo transport of proteins from allergen across the presensitized gut. *Biochem Biophys Res Commun* (2004) 325:1258–63. doi:10.1016/j.bbrc.2004.10.161
 18. Chambers SJ, Bertelli E, Winterbone MS, Regoli M, Man AL, Nicoletti C. Adoptive transfer of dendritic cells from allergic mice induces specific immunoglobulin E antibody in naïve recipients in absence of antigen challenge without altering the T helper 1/T helper 2 balance. *Immunology* (2004) 112:72–9. doi:10.1111/j.1365-2567.2004.01846.x
 19. Sato T, Clevers H. Primary mouse small intestinal epithelial cell cultures. *Methods Mol Biol* (2013) 945:319–28. doi:10.1007/978-1-62703-125-7_19
 20. Vidal K, Grosjean I, Evillard JB, Gespach C, Kaiserlian D. Immortalization of mouse intestinal epithelial cells by the SV40-large T gene. Phenotypic and immune characterization of the MODE-K cell line. *J Immunol Methods* (1993) 166:63–73. doi:10.1016/0022-1759(93)90329-6
 21. Presky DH, Yang H, Minetti LJ, Chua AO, Nabavi N, Wu CY, et al. A functional interleukin 12 receptor complex is composed of two beta-type cytokine receptor subunits. *Proc Natl Acad Sci U S A* (1996) 93:14002–7. doi:10.1073/pnas.93.24.14002
 22. Verhagen CE, de Boer T, Smits HH, Verreck FA, Wierenga EA, Kurimoto M, et al. Residual type 1 immunity in patients genetically deficient for interleukin 12 receptor beta1 (IL-12Rbeta1): evidence for an IL-12Rbeta1-independent pathway of IL-12 responsiveness in human T cells. *J Exp Med* (2000) 192:517–28. doi:10.1084/jem.192.4.517
 23. Ziegler SF. Thymic stromal lymphopoietin and allergic disease. *J Allergy Clin Immunol* (2012) 130:845–52. doi:10.1016/j.jaci.2012.07.010
 24. Ziegler SF, Artis D. Sensing the outside world: TSLP regulates barrier immunity. *Nat Immunol* (2010) 11:289–93. doi:10.1038/ni.1852
 25. Chu DK, Llop-Guevara A, Walker TD, Flader K, Goncharova S, Boudreau JE, et al. IL-33, but not thymic stromal lymphopoietin or IL-25, is central to mite and peanut allergic sensitization. *J Allergy Clin Immunol* (2013) 131:187–200. doi:10.1016/j.jaci.2012.08.002
 26. Wu C, Ferrante J, Gately MK, Magram J. Characterization of IL-12 receptor beta1 chain (IL-12Rbeta1)-deficient mice: IL-12Rbeta1 is an essential component of the functional mouse IL-12 receptor. *J Immunol* (1997) 159:1658–65.
 27. Bacon CM, Petricoin EF III, Ortaldo JR, Rees RC, Larner AC, Johnston JA, et al. Interleukin 12 induces tyrosine phosphorylation and activation of STAT4 in human lymphocytes. *Proc Natl Acad Sci U S A* (1995) 92:7307–11. doi:10.1073/pnas.92.16.7307
 28. Bacon CM, McVicar DW, Ortaldo JR, Rees RC, O'Shea JJ, Johnston JA. Interleukin 12 (IL-12) induces tyrosine phosphorylation of JAK2 and TYK2: differential use of Janus family tyrosine kinases by IL-2 and IL-12. *J Exp Med* (1995) 181:399–404. doi:10.1084/jem.181.1.399
 29. Wurster AL, Tanaka T, Grusby MJ. The biology of Stat4 and Stat6. *Oncogene* (2000) 19:2577–84. doi:10.1038/sj.onc.1203485
 30. Spadoni I, Iliev ID, Rossi G, Rescigno M. Dendritic cells produce TSLP that limits the differentiation of Th17 cells, fosters Treg development, and protects against colitis. *Mucosal Immunol* (2012) 2:184–93. doi:10.1038/mi.2011.64
 31. Neill DR, Wong SH, Bellosi A, Flynn RJ, Daly M, Langford TK, et al. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature* (2010) 464:1367–70. doi:10.1038/nature08900
 32. Taylor BC, Zaph C, Troy AE, Du Y, Guild KJ, Comeau MR, et al. TSLP regulates intestinal immunity and inflammation in mouse models of helminth infection and colitis. *J Exp Med* (2009) 206:655–66. doi:10.1084/jem.20081499
 33. Khodoun MV, Tomar S, Tocker JE, Wang YH, Finkelman FD. Prevention of food allergy development and suppression of established food allergy by neutralization of thymic stromal lymphopoietin, IL-25, and IL-33. *J Allergy Clin Immunol* (2018) 141:171–9. doi:10.1016/j.jaci.2017.02.046
 34. Mehta AK, Duan W, Doerner AM, Traves SL, Broide DH, Proud D, et al. Rhinovirus infection interferes with induction of tolerance to aeroantigens through OX40 ligand, thymic stromal lymphopoietin, and IL-33. *J Allergy Clin Immunol* (2016) 137:278–88. doi:10.1016/j.jaci.2015.05.007

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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