





# Asia and Europe: So Distant So Close? The Case of *Lipoptena fortisetosa* in Italy

Annalisa Andreani<sup>1</sup>, Annunziata Giangaspero<sup>2\*</sup> , Marianna Marangi<sup>2</sup>, Alessandra Barlaam<sup>2</sup>,  
Maria Paola Ponzetta<sup>1</sup>, Lise Roy<sup>3\*</sup> , Antonio Belcari<sup>1</sup> , Patrizia Sacchetti<sup>1</sup> 

<sup>1</sup>Department of Agriculture, Food, Environment and Forestry (DAGRI), University of Florence, Piazzale delle Cascine 18, 50144, Firenze, Italy;

<sup>2</sup>Department of Agriculture, Food, Natural Resources and Engineering (DAFNE), University of Foggia, Via Napoli 25, 71121 Foggia, Italy;

<sup>3</sup>CEFE, Univ Montpellier, CNRS, EPHE, IRD, Univ Paul, Valéry Montpellier 3, Montpellier, France

**Abstract:** In Europe, 5 *Lipoptena* species have been recorded, including *Lipoptena fortisetosa*. This species, native to Asian countries, was described as a parasite of sika deer and its appearance in Europe dates back to more than 50 years ago. *Lipoptena fortisetosa* has been recently reported in Italy, sharing its hosts with *Lipoptena cervi*. A morpho-molecular approach was developed to determine the phylogenetic interrelationship of Italian and Asian CO1 haplotypes sequenced from *Lipoptena* fly individuals collected in Italy, and their DNA sequences were compared with conspecifics available in GenBank; morphological key-characters (terminalia) of *L. fortisetosa* were compared with the original description. Two haplotypes were recorded from Italy and assigned to *L. cervi* and *L. fortisetosa*, respectively. The latter was part of the monophyletic clade *L. fortisetosa*, along with 2 Central European and 2 Korean haplotypes (100% identical to one of the Korean haplotypes); moreover, Italian *L. fortisetosa* female terminalia were consistent with the original description of Asian individuals. Pending more in-depth investigations, this study provides a first answer to the hypothesis of the recent colonization of Italy by *L. fortisetosa* from Asia as we did not detect any obvious and stable morphological and molecular differences in specimens from the 2 geographical areas. The presence of the sika deer in Europe was retraced and the possible route traveled by the parasite from Asia and the eco-biological factors that may have enhanced its settlement are discussed.

**Key words:** *Lipoptena fortisetosa*, phylogeny, morphology, Italy, Asia

## INTRODUCTION

Deer keds *Lipoptena* spp. (Hippoboscidae, Lipopteninae) are blood-sucking obligate ectoparasites of almost exclusively Cervidae (deer) and Bovidae (cattle, goats, chamois, antelopes, etc.), and can occasionally bite humans [1-3]. Indeed, the *Lipoptena* genus includes about 30 species spread worldwide [1]; most of them occur in the Palearctic region, mainly in continental Europe and Asia, while 8 species are native to far East Asian countries. Five species have been recorded in America, with 4 of them native to this continent [4]. Fragmentary information is available on the species accounted for African countries [5,6] (Supplementary Table S1).

In severe infestations, *Lipoptena* spp. may be responsible for

anemia and skin lesions and may be involved in the transmission of several pathogens [7,8].

In Europe, 5 species of *Lipoptena* have been recorded: *Lipoptena capreoli* Rondani, 1878, *Lipoptena couturieri* Ségué, 1935, *Lipoptena arianae* Maa, 1969, *Lipoptena cervi* (Linnaeus, 1758), and *Lipoptena fortisetosa* Maa, 1965 [9]; however, the presence and the geographical range of the first 3 species need to be confirmed.

*Lipoptena cervi* may be considered the oldest deer ked in Europe as the relationship of this species with wild ungulates dates back to more than 5,000 years ago, being found in the remains of a Late Neolithic human mummy discovered in a glacier in the Alps [10]. It has a wide distribution in Europe and has currently been reported from more than 20 countries [11]. *Lipoptena cervi* has been recorded on *Cervus elaphus*, *Dama dama*, *Alces alces*, *Rupicapra rupicapra*, *Capreolus capreolus*, *Moschus moschiferus* [9] and it has the potential to transmit bacteria e.g., *Bartonella* spp., *Borrelia* spp. [12-14], *Anaplasma* spp., *Ehrlichia* spp., and *Rickettsia* spp., and protozoans, e.g., *Babesia* spp., *Theileria* spp., *Hepatozoon* spp. [15-17].

•Received 2 August 2020, revised 4 October 2020, accepted 13 October 2020.

\*Corresponding authors (annunziata.giangaspero@unifg.it; lise.roy@univ-montp3.fr)

© 2020, Korean Society for Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Lipoptena fortisetosa* was originally recorded in Japan from the sika deer *Cervus nippon* [18-20], and is considered quite restricted to this ungulate, although it has been occasionally collected from a bird *Emberiza spodocephala* [21]. It has also been reported from the Siberian roe deer *Capreolus pygargus* in South Korea [22] and in *Capreolus capreolus* in Kazakhstan [23]. This species was first reported in Europe about 50 years ago when it was found in the Czech Republic [24] and later in the Moscow district in Russia [25]. Afterward, from the 80s' to date, it has been confirmed and/or recorded in 12 countries, i.e., Czech Republic, Poland, Moldavia, Germany, Switzerland, Lithuania, Romania, Austria, Belarus, Slovak Republic, Moscow-district, and Estonia [26]. The range of *L. fortisetosa* has expanded in the southern part of Europe, including Italy, where it has very recently been reported [27,28]. In Europe, *L. fortisetosa* attacks mainly deer [29,30], and occasionally cattle [31], goats, sheep [29,30], dogs [32,33], and humans, as reported in Germany [34], Estonia [26], and Slovakia [35]. *Lipoptena fortisetosa* has been found to mechanically carry pathogens, i.e., *Coxiella*-like bacteria (CLB), *Theileria luwenshumi*, and *Theileria ovis* [36]. In addition, very recently, both *L. cervi* and *L. fortisetosa* specimens from Poland were found positive to *Trypanosoma* DNA [37].

In Europe, *L. fortisetosa* appears to share with *L. cervi* approximately the same ungulate species as the host group (above listed) and roughly the same territory [26]. However, it is unclear whether the native host (i.e., the sika deer *Cervus nippon*) of *L. fortisetosa* played a role in spreading the Asian species to Europe or the parasite propagated independently, as already speculated [38], or to which extent human activities might have helped the ked expansion. *Lipoptena fortisetosa* likely dispersed widely and settled in Europe quite quickly, so that possible changes in genetic constitution compared to the Asian indigenous populations may be hypothesized.

Molecular investigations coupled to morphological analysis, as well as the ecological requirements, help to provide more insight into the perspective of the integrated taxonomy concept [39].

In order to state whether there are sharp differences between Italian and Asian individuals, we developed a morpho-molecular approach. In particular, we investigated the phylogenetic interrelationship of the mitochondrial cytochrome c oxidase subunit 1 (CO1) gene haplotypes of *Lipoptena* individuals sampled in Italy-analyzing them in a wider context together with the DNA sequences of conspecifics available in the NCBI GenBank and morphologically analyzed the Italian popula-

tion of *L. fortisetosa*. We also focused on the observation of some stable key-characters (terminalia) and compared them with the original description of the species, as well as with the indigenous *L. cervi* species, as additional documentation. Finally, the presence of the sika deer in Europe was retraced in order to discuss the possible route traveled by the ectoparasite from Asia and the eco-biological factors that may have enhanced its settlement.

## MATERIAL AND METHODS

### Ethics statement, specimens and processing

All animal handling procedures followed all regional, national, and institutional guidelines.

From a total of 312 *Lipoptena* specimens, previously collected from 3 species of wild ruminants [28], belonging to *Lipoptena cervi* and *Lipoptena fortisetosa*, the following were selected from different host species: 10 flies each from 5 *Cervus elaphus* hosts (total 50 specimens: 30 and 20 specimens belonging to *L. cervi* and *L. fortisetosa*, respectively); 10 flies from 3 *Capreolus capreolus* hosts (total 30 specimens, all belonging to *L. fortisetosa*), 10 flies from 1 *Dama dama* host (total 10 specimens, all belonging to *L. fortisetosa*), and frozen at  $-20^{\circ}\text{C}$ , until DNA extraction. Genomic DNA was extracted individually from the abdomens using the Nucleospin Tissue kit (Macherey-Nagel, Amsterdam, Netherlands) in accordance with the manufacturer's instructions. The extracted DNA was eluted in 50  $\mu\text{l}$  of distilled water and the samples were stored at  $-20^{\circ}\text{C}$ , pending molecular analysis. PCR amplifications were performed in a CFX96 thermal cycler (Bio-Rad, Hercules, California, USA) using 10  $\mu\text{l}$  of Phire Reaction Buffer 5X (Thermo Scientific, Waltham, Massachusetts, USA), 0.4  $\mu\text{l}$  of dNTPs (200  $\mu\text{M}$ ) (Qiagen, Germantown, Maryland, USA), 1  $\mu\text{l}$  of specific primer pairs (10  $\mu\text{M}$ ), 0.4  $\mu\text{l}$  of Phire Hot Start II DNA Polymerase 1 U (Thermo Scientific), and 5  $\mu\text{l}$  (approximately 100  $\mu\text{g}$ ) of genomic DNA per reaction. A blank control (pure water instead of genomic DNA) was included in each PCR run.

An approximately 710-bp gene fragment of CO1 was amplified using primers LCO-1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO-2198 (5'-TAAACTTCAGGGTGACAAAAATCA-3') [40]. The cycling parameters were: 2 min denaturing at  $94^{\circ}\text{C}$ , followed by 35 cycles of 30 sec at  $94^{\circ}\text{C}$ , 30 sec at  $56^{\circ}\text{C}$  and 60 sec at  $70^{\circ}\text{C}$ , and final extension of 7 min at  $70^{\circ}\text{C}$ . PCR products were run on 1.2% agarose gel, and positive samples purified with exonuclease I (EXO I) and thermo-

sensitive alkaline phosphatase (FAST AP) (Fermentas, Waltham, Massachusetts, USA) enzymes, in accordance with the manufacturer's instructions.

PCR products were directly sequenced in both directions using the ABI PRIMS BigDye Terminator v. 3.1 Cycle Sequencing Kit (Applied Biosystems, Foster City, California, USA) with the same primers as the respective PCR reactions, in accordance with the manufacturer's instructions. The sequences obtained were determined using an ABI PRISM 3130 Genetic Analyser (Applied Biosystems), chromatograms were inspected by eye using FinchTV (<https://digitalworldbiology.com/FinchTV>) and primer regions plus bad-quality regions were removed. Once the sequences were cleaned up, each sequence was compared with the *Lipoptena* spp. homologous nucleotide sequences available in the GenBank database using the BLAST program (Basic Local Alignment Search Tool; [https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastn&PAGE\\_TYPE=BlastSearch&LINK\\_L+OC=blasthome](https://blast.ncbi.nlm.nih.gov/Blast.cgi?PROGRAM=blastn&PAGE_TYPE=BlastSearch&LINK_L+OC=blasthome)). The 19 sequences having the highest percent similarity with our sequences and labelled as *Lipoptena* CO1 in Genbank were then sampled and gathered in a FASTA file with our own sequences. The new sequence dataset was aligned using the CLUSTALW implementation of BIOEDIT, version 7.0.5 (<http://www.mbio.ncsu.edu/BioEdit/bioedit.html>) and the alignment adjusted manually, if necessary. Once the sequences were aligned, the absence of stop codons was checked. Phylogenetic analysis of the obtained sequences and homologous sequences from GenBank were performed using the maximum likelihood method in MEGA, version 7.0.9 (<https://www.megasoftware.net>). Bootstrap confidence values for the branching reliability were calculated with 10,000 replicates.

All specimens of *L. fortisetosa* and *L. cervi* intended for molecular analysis were morphologically identified, based on Maa's original description [19] and a recent taxonomic key [9]. Other specimens were processed for Scanning Electron Microscope (SEM) observations, according to the procedures previously described [28] to further examine male and female terminalia. Description of terminalia features follows the terminology and nomenclature reported by Maa and Peterson [41].

## RESULTS

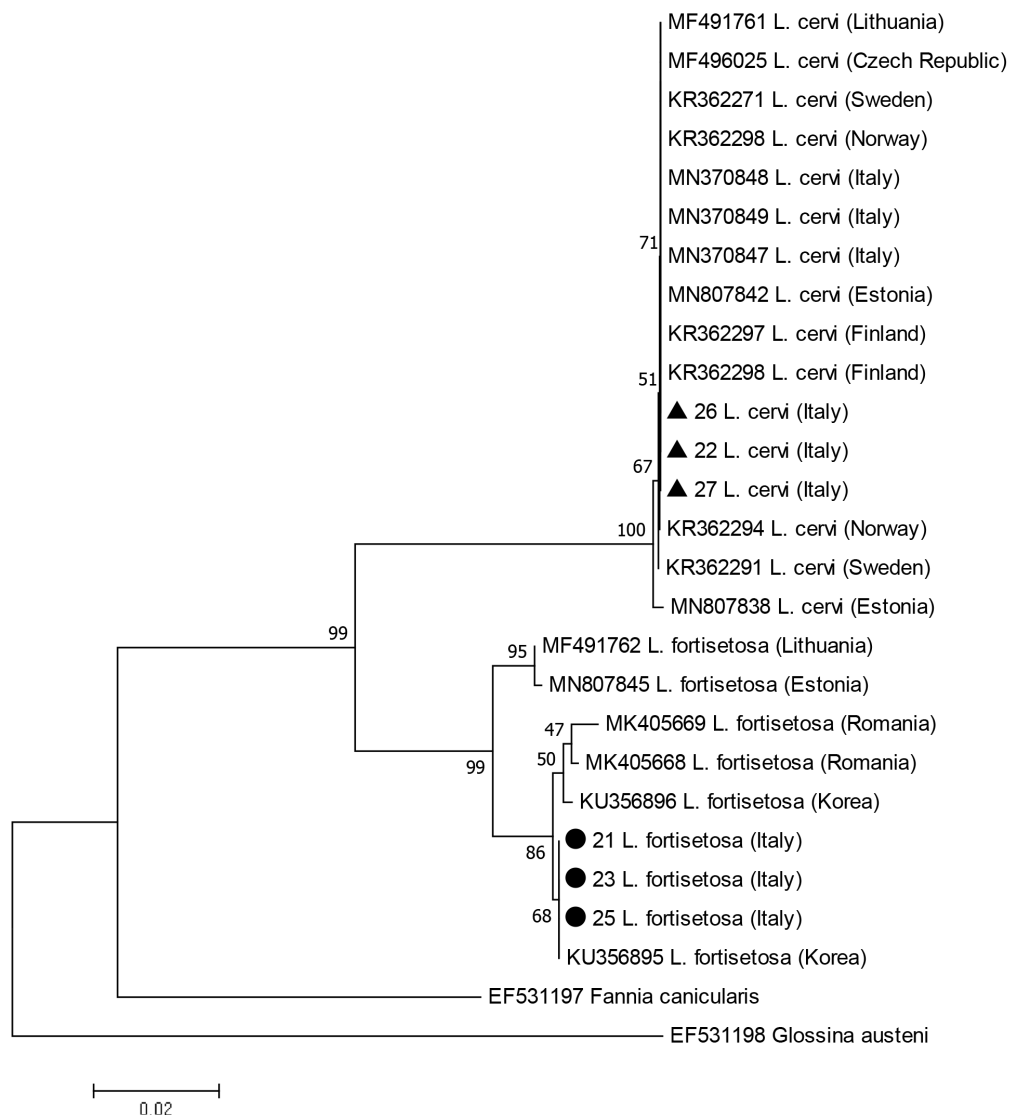
A total 60/90 (66.7%) of the specimens provided good quality PCR fragments and sequences for the CO1 gene. After alignment with the homologous sequences of *Lipoptena* spp. available in GenBank, 2 sets of sequences were identified, one

with the mean percentage of identity of 89% with *L. cervi* and 92% with *L. fortisetosa*, and one with 97% with *L. cervi* and 86% with *L. fortisetosa*. Two haplotypes were recorded from the 60 specimens sequenced in the present study and the phylogenetic analysis confirmed that these 2 haplotypes belonged to *L. cervi* and *L. fortisetosa*, respectively. The data matrix comprised 14 haplotypes of *L. cervi* (one haplotype from the present study, 13 downloaded from GenBank) and 7 haplotypes of *L. fortisetosa* (one haplotype from the present study, 6 downloaded). The genetic distances ranged from 0.081-0.084 for *L. cervi* group sequences and 0.003-0.018 for *L. fortisetosa* group sequences.

While the clade *L. fortisetosa* appears to be monophyletic, the internal 2-subclade structure of this clade is poorly supported (low bootstrap values) as expected within the species. We will thus note essentially the following points: the single haplotype recorded from the Italian *L. fortisetosa* specimens was closely related to 2 Central European haplotypes and 100% identical to one of the 2 haplotypes (KU356895) found in Korea (Fig. 1).

Morphological investigations showed that female terminalia of *L. fortisetosa* are characterized by a typical pregenital sclerite that is peg-like and bears 2 or 3 strong bristles (Fig. 2A, C). The pregenital plate is elongated and lozenge-shaped, and the underlying hypoproct is covered by several bristles interspersed with an area densely hairy. The genital opening is clearly visible between the pregenital sclerite and the pregenital plate (Fig. 2C). Female terminalia of *L. cervi* showed the presence of 3 pregenital sclerites bearing several differently sized bristles (Fig. 2B, D). The central sclerite is bigger and with more numerous setae than the external 2; the pregenital plate shows many series of long setae arranged in the distal portion, while the hypoproct is completely bare. Male terminalia of *L. fortisetosa* (Fig. 2E) consist of 2 well-sclerotized and slender gonopods that guide the aedeagus. This latter is wider in the proximal part and ends with a bilobate tip provided with spines. In *L. cervi*, the gonopods (Fig. 2F) are similar to those of *L. fortisetosa*. However, the aedeagus is membranous in the proximal and lateral parts, while in the middle it is formed by 2 fused and sclerotized strips ending in a ridge tip. Morphological observations of *L. cervi* and *L. fortisetosa* strengthened the strong diversity between these 2 species [28] but, more importantly, demonstrate that the features of *L. fortisetosa* female terminalia are consistent with the original description by Maa [18] (Fig. 3).

Taking into account that terminalia are considered stable

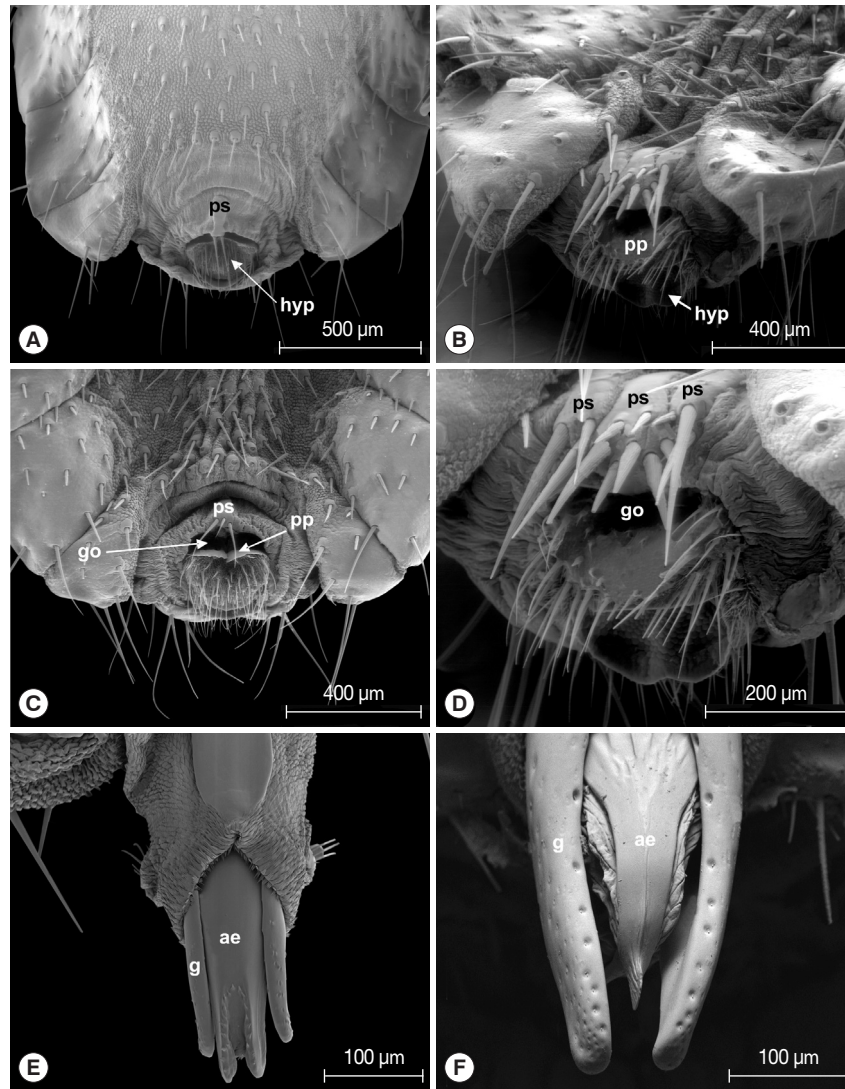


**Fig. 1.** Phylogenetic topology based on the analysis of the maximum likelihood of the partial *CO1* gene sequences from *Lipoptena* individuals from the present study and *Lipoptena* sequences available from GenBank. Labels include accession numbers, species identity and country origin. The 2 haplotypes found from the present study are labelled with a black triangle for *Lipoptena cervi* and a black circle for *Lipoptena fortisetosa*. *Fannia canicularis* and *Glossina austeni* sequences were used as outgroups. The percentage of trees in which the associated individuals clustered together is shown next to the branches.

features that allow a correct morphological identification at species-specific level in Diptera [42], including hippoboscids [19,43], this supports the above confirmation that the boundaries between the 2 species (*L. fortisetosa* and *L. cervi*) are correct based on morphology and molecular data. Therefore, we can conclude that the individuals of which we only have the sequences (GenBank) belong to the same species.

*Lipoptena fortisetosa* has been introduced in Europe probably with *Cervus nippon*, the sika deer [28,33,44], during the last 150 years of restocking of deer in the Continent, apparently,

since 1893, and probably a number of times [44]. Sika deer has successfully settled in the European fauna thanks to its high potential to compete with autochthonous species and readiness to hybridize with native red deer, as demonstrated by the presence of hybrids of sika with red deer in several countries [45-49]. Sika deer (or hybrids) is currently present in 20 European countries, including Italy, where it was recently reported [50]. Spreading through Europe, sika deer has likely carried and disseminated its ectoparasites, including *L. fortisetosa* that is currently recorded in 13 European countries (Fig. 4),



**Fig. 2.** Terminalia of *Lipoptena fortisetosa* (A&C, female; E, male) and *Lipoptena cervi* (B&D, female; F, male). ps, pregenital sclerite; hyp, hypoproct; go, genital opening; pp, pregenital plate; ae, aedeagus; g, gonopod.

Italy included [27].

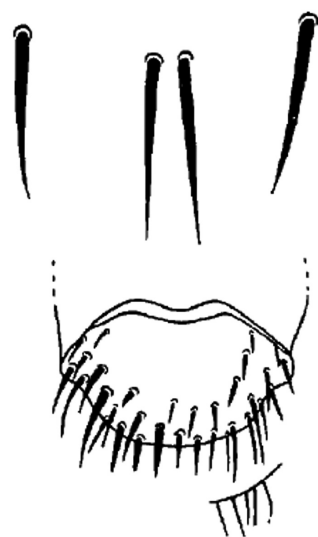
## DISCUSSION

According to the present results, while *L. cervi* confirms previous findings [9], we cannot exclude a scenario depicting a recent colonization(s) of *L. fortisetosa* in Europe. According to the latest studies, the hypothesis of geographically distinct CO1 lineages should be rejected at the Central Europe level [26]. Our study suggests that this hypothesis should also be rejected at a global level. However, we were unable to determine the time of colonization or to identify the actual host that introduced *L. fortisetosa* from Asia to Europe. In fact, this parasite

has been collected from many different cervids, e.g., red deer (*Cervus elaphus*) [27,28], Manchurian elk (*Cervus elaphus xanthopygus*), Maral red deer (*Cervus elaphus maral*), fallow deer (*Dama dama*) [34], Korean water deer (*Hydropotes inermis*) [36], roe deer (*Capreolus capreolus*) [23], Siberian roe deer (*Capreolus pygargus*) [22], both in Europe and in Asia. The possibility of recent colonization by *L. fortisetosa* specimens from cervids purchased for the restocking of deer farms, or in captive hosts in the Oriental region transferred to fenced areas for recreational or conservation purposes, cannot be excluded. We also suggest, as a minor hypothesis, the relatively recent migration to Italy of sika or hybrid individuals from neighboring countries that may have transmitted the ectoparasite joining red

deer groups.

This study doesn't reject the hypothesis of the recent colonization of Italy by *L. fortisetosa* from Asia as no obvious and stable morphological and molecular differences were observed in the populations from the 2 regions. However, such a hypothesis requires further study, including a straightforward analysis



**Fig. 3.** Female terminalia of *Lipoptena fortisetosa*, drawing from Maa [18].

of numerous specimens from Asian and European countries. In line with other authors' suggestions [26], population genetic analyses of Asian and European keds' populations are required to evaluate whether the colonization scenario is likely or not. Moreover, wider investigations aimed at comparing genetic make-up of populations from different countries, host and parasite distribution, together with morphological differentiation, might resolve phylogenetic relationships of this neglected group within a desirable integrated taxonomic framework as applied in other groups of insects [51,52].

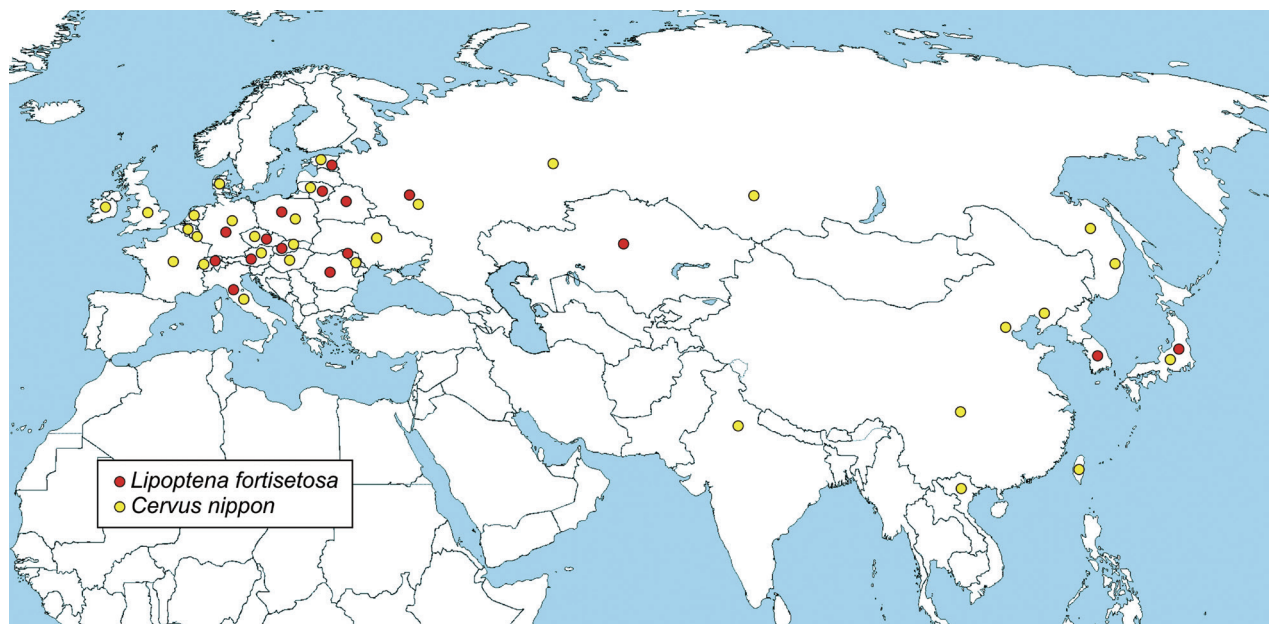
Due to the ability of *L. fortisetosa* to parasitize a wide range of homeothermic animals, there are no obvious limitations to its further expansion. Given such possibility, the potential for transmitting pathogens, and the frequent attacks reported among humans [26], more in-depth investigations are required on *Lipoptena* species with a One Health perspective.

#### ACKNOWLEDGMENT

This research was partially funded by the "Fondazione Cassa di Risparmio di Firenze, Italy".

#### CONFLICT OF INTEREST

The authors don't have any conflicts of interest to declare.



**Fig. 4.** Distribution map of *Lipoptena fortisetosa* (red dot) and *Cervus nippon* (sika deer) (yellow dot). Data from different sources [11,19,23,26,44,49,50].

## REFERENCES

- Dick CW. Checklist of world Hippoboscidae (Diptera: Hippoboscoidea). Chicago, USA. Department of Zoology, Field Museum of Natural History. 2006, pp 1-7.
- Kortet R, Härkönen L, Hokkanen P, Härkönen S, Kaitala A, Kaunisto S, Laaksonen S, Kekäläinen J, Ylönen H. Experiments on the ectoparasitic deer ked that often attacks humans; preferences for body parts, colour and temperature. *Bull Entomol Res* 2010; 100: 279-285. <https://doi.org/10.1017/S0007485309990277>
- Soós Á, Húrka K. Family hippoboscidae. In Soós Á, Papp L eds, *Catalogue of Palaearctic Diptera, Vol 11*. Budapest, Hungary. Akademiai Kiado 1986, pp 215-226.
- Skvarla MJ, Machtinger ET. Deer keds (Diptera: Hippoboscidae: *Lipoptena* and *Neolipoptena*) in the United States and Canada: new state and county records, pathogen records, and an illustrated key to species. *J Med Entomol* 2019; 56: 744-760. <https://doi.org/10.1093/jme/tjy238>
- GBIF.org. GBIF Home Page [Internet]; [cited 2020 September 20]. Available from: <https://www.gbif.org>
- Visagie EJ. A new species of *Lipoptena* (Diptera: Hippoboscidae) from Southern Africa. *Onderstepoort J Vet Res* 1992; 59: 293-302.
- Härkönen S, Laine M, Vornanen M, Reunala T. Deer ked (*Lipoptena cervi*) dermatitis in humans—an increasing nuisance in Finland. *Alces* 2009; 45: 73-79.
- Lazăr M, Iacob OC, Solcan C, Pașca SA, Lazăr R, Boișteanu PC. The first report of massive infestation with *Lipoptena cervi* (Diptera: Hippoboscidae) in roe deer (*Capreolus capreolus*) in Iasi county, N-E of Romania. *Arq Bras Med Vet Zootec* 2017; 69: 293-298. <https://doi.org/10.1590/1678-4162-8612>
- Salveti M, Bianchi A, Marangi M, Barlaam M, Giacomelli S, Bertolotti I, Roy L, Giangaspero A. Deer keds on wild ungulates in northern Italy, with a taxonomic key for the identification of *Lipoptena* spp. of Europe. *Med Vet Entomol* 2020; 34: 74-85. <https://doi.org/10.1111/mve.12411>
- Gothé R, Schöl H. Deer keds (*Lipoptena cervi*) in the accompanying equipment of the late Neolithic human mummy from the Similaun, South Tyrol. *Parasitol Res* 1994; 80: 81-83. <https://doi.org/10.1007/BF00932630>
- Fauna Europaea. All European Animal Species Online [Internet]; [cited 2020 September 20]. Available from: <https://fauna-eu.org>.
- Dehio C, Sauder U, Hiestand R. Isolation of *Bartonella schoenbuchensis* from *Lipoptena cervi*, a blood-sucking arthropod causing deer ked dermatitis. *J Clin Microbiol* 2004; 42: 5320-5323. <https://doi.org/10.1128/JCM.42.11.5320-5323.2004>
- Duodu S, Madslie K, Hjelm E, Molin Y, Paziewska-Harris A, Harris PD, Colquhoun DJ, Ytrehus B. *Bartonella* infections in deer Keds (*Lipoptena cervi*) and moose (*Alces alces*) in Norway. *Appl Environ Microbiol* 2013; 79: 322-327. <https://doi.org/10.1128/AEM.02632-12>
- Szewczyk T, Werszko J, Steiner-Bogdaszewska Ż, Jeżewski W, Lasowski Z, Karbowski G. Molecular detection of *Bartonella* spp. in deer ked (*Lipoptena cervi*) in Poland. *Parasit Vectors* 2017; 10: 487. <https://doi.org/10.1186/s13071-017-2413-0>
- Chomel BB, Boulouis HJ, Breitschwerdt EB, Kasten RW, Vayssier-Taussat M, Birtles RJ, Koehler JE, Dehio C. Ecological fitness and strategies of adaptation of *Bartonella* species to their hosts and vectors. *Vet Res* 2009; 40: 29. <https://doi.org/10.1051/vetres/2009011>
- Hornok S, de la Fuente J, Biró N, Fernández de Mera IG, Meli ML, Elek V, Gönczi E, Meili T, Tánzos B, Farkas R, Lutz H, Hofmann-Lehmann R. First molecular evidence of *Anaplasma ovis* and *Rickettsia* spp. in keds (Diptera: Hippoboscidae) of sheep and wild ruminants. *Vector Borne Zoonotic Dis* 2011; 11: 1319-1321. <https://doi.org/10.1089/vbz.2011.0649>
- Kelsey A, Finch J. Deer ked: a Lyme-carrying ectoparasite on the move. *Cutis* 2018; 102: 121-122.
- Maa TC. A synopsis of the Lipopteninae (Diptera: Hippoboscidae). *J Med Entomol* 1965; 2: 233-248. <https://doi.org/10.1093/jmedent/2.3.233>
- Maa TC. A revised checklist and concise host index of Hippoboscidae (Diptera). *Pacific Insects Monograph* 1969; 20: 261-269.
- Mogi M. A new species of *Lipoptena* (Diptera, Hippoboscidae) from the Japanese deer. *Kontyû* 1975; 43: 387-392.
- Yamauchi T, Tsurumi M, Kataoka N. Distributional records of *Lipoptena* species (Diptera: Hippoboscidae) in Japan and Jeju-do, Korea. *Med Entomol Zoo* 2009; 60: 131-133. <https://doi.org/10.7601/mez.60.131>
- Choi CY, Lee S, Moon KH, Kang CW, Yun YM. New Record of *Lipoptena fortisetosa* (Diptera: Hippoboscidae) collected from Siberian roe deer on Jeju Island, Korea. *J Med Entomol* 2013; 50: 1173-1177. <https://doi.org/10.1603/me12150>
- Edwards SJ, Hood MW, Shaw JH, Rayburn JD, Kirby MD, Hanfman DT, Zidar JA. Index-Catalogue of Medical and Veterinary Zoology. Supplement 21, Part 5: Parasite-Subject Catalogue. Parasites: Arthropoda and Miscellaneous Phyla. Washington DC, USA. USDA Government Printing Office. 1978, pp 246.
- Theodor O. *Lipoptena parvula*, n. sp., eine neue Art aus der Tschechoslowakei (Diptera, Hippoboscidae). *Acta Entomol Mus Natl Pragae* 1965; 37: 275-278.
- Grunin KJ. Hippoboscidae-krovososki. In Bej-Bienko GJ ed. *Opređliti nasekomych evropejskoj czasti SSSR*. T. 5. Nauka, Leningrad. 1970, pp 979-987 (in Russian).
- Kurina O, Kirik H, Õunap H, Õunap E. The northernmost record of a blood-sucking ectoparasite, *Lipoptena fortisetosa* Maa (Diptera: Hippoboscidae), in Estonia. *Biodivers Data J* 2019; 7: e47857. <https://doi.org/10.3897/BDJ.7.e47857>
- Andreani A, Belcari A, Sacchetti P, Ponzetta MP. Occurrence of a new parasite of the ungulates in Italy: *Lipoptena fortisetosa* (Diptera: Hippoboscidae). *Atti del IV Congresso Nazionale di Ecopatologia della Fauna*; 2017 October 11-13. S.I.E.F. Società Italiana di Ecopatologia della Fauna. 2017, pp 36.
- Andreani A, Sacchetti P, Belcari A. Comparative morphology of the deer ked *Lipoptena fortisetosa* first recorded from Italy. *Med Vet Entomol* 2019; 33: 140-153. <https://doi.org/10.1111/mve.12342>
- Büttiker W. Die Lausfliegen der Schweiz (Diptera, Hippoboscidae).

- dae): mit Bestimmungsschlüssel. Neuchâtel, Switzerland. Centre Suisse de cartographie de la faune. 1994, pp 1-117 (in German).
30. Schedl W. Beitrag zur Lausfliegen-Fauna an Säugetieren von Kärnten und anderen Bundesländern Österreichs (Insecta: Diptera, Hippoboscidae). *Linzer biol Beitr* 2018; 50: 1283-1293 (in Deutsch).
  31. Metelitsa AK, Veselkin GA. Parasitism of the louse fly *Lipoptena fortisetosa* on cattle. *Parazitologiiia* 1989; 23: 276-277. [In Russian].
  32. Sokół R, Gałęcki R. Prevalence of keds on city dogs in Central Poland. *Med Vet Entomol* 2017; 31: 114-116. <https://doi.org/10.1111/mve.12209>
  33. Mihalca AD, Păstrav IR, Sándor AD, Deak G, Gherman CM, Sarmași A, Votýpka J. First report of the dog louse fly *Hippobosca longipennis* in Romania. *Med Vet Entomol* 2019; 33: 530-535.
  34. Schumann H, Messner B. Erstnachweis von *Lipoptena fortisetosa* Maa, 1965 in Deutschland (Dipt., Hippoboscidae). *Entomol Nachr Ber* 1993; 37: 247-249.
  35. Oboňa J, Sychra O, Greš S, Heřman P, Manko P, Roháček J, Šestáková A, Šlapák J, Hromada M. A revised annotated checklist of louse flies (Diptera, Hippoboscidae) from Slovakia. *ZooKeys* 2019; 862: 129-152. <https://doi.org/10.3897/zookeys.862.25992>
  36. Lee SH, Kim KI, Kwon OD, Ock Y, Kim T, Choi D, Kwak D. Novel detection of *Coxiella* spp., *Theileria luwenshuni*, and *T. ovis* endosymbionts in deer keds (*Lipoptena fortisetosa*). *PLoS One* 2016; 11: e0156727. <https://doi.org/10.1371/journal.pone.0156727>
  37. Werszko J, Steiner-Bogdaszewska Ż, Jeżewski W, Szewczyk T, Kuryło G, Wolkowycy M, Wróblewski P, Karbowski G. Molecular detection of *Trypanosoma* spp. in *Lipoptena cervi* and *Lipoptena fortisetosa* (Diptera: Hippoboscidae) and their potential role in the transmission of pathogens. *Parasitology*. 2020; 1-26. <https://doi.org/10.1017/S0031182020001584>.
  38. Kowal J, Nosal P, Komaś S, Wajdzik M, Matysek M, Basiaga M. Biodiversity and importance of hippoboscids infection in cervids. *Med Weter* 2016; 72: 745-749.
  39. Schlick-Steiner BC, Steiner FM, Seifert B, Stauffer C, Christian E, Crozier RH. Integrative taxonomy: a multisource approach to exploring biodiversity. *Annu Rev Entomol* 2010; 55: 421-438. <https://doi.org/10.1146/annurev-ento-112408-085432>
  40. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. DNA primers for amplification of mitochondrial cytochrome CO11dase subunit I from diverse metazoan invertebrates. *Mol Mar Biol Biotechnol* 1994; 3: 294-299.
  41. Maa T.C., Peterson BV. Hippoboscidae. In McAlpine JF, Peterson BV, Shewell GE, Teskey HJ, Vockeroth JR, Wood DM eds, *Manual of Nearctic Diptera*, Vol. II. Monograph 28. Ottawa, ON, Canada. Research Branch, Agriculture Canada. 1987; pp 1271-1281.
  42. McAlpine JF. Morphology and terminology – Adults. In McAlpine JF, Peterson BV, Shewell GE, Teskey HJ, Vockeroth JR, Wood DM eds, *Manual of Nearctic Diptera*, Vol. I. Monograph 27. Ottawa, ON, Canada. Research Branch, Agriculture Canada. 1981; pp 9-63.
  43. Bequaert J. A monograph of the Melophaginae, or ked-flies, of sheep, goats, deer and antelopes (Diptera, Hippoboscidae). *Entomol Americana* 1942; 22: 1-220.
  44. Bartoš L. Sika deer in continental Europe. In McCullough DR, Takatsuki S, Kaji K eds, *Sika Deer: Biology and Management of Native and Introduced Populations*. Tokyo, Japan. Springer. 2009, pp 573-594.
  45. Lowe VPW, Gardiner AS. Hybridization between Red deer (*Cervus elaphus*) and Sika deer (*Cervus nippon*) with particular reference to stocks in N.W. England. *J Zool* 1975; 177: 553-566. <https://doi.org/10.1111/j.1469-7998.1975.tb02259.x>
  46. Goodman SJ, Barton NH, Swanson G, Abernethy K, Pemberton JM. Introgression through rare hybridization: a genetic study of a hybrid zone between Red and Sika deer (*Genus Cervus*) in Argyll, Scotland. *Genetics* 1999; 152: 355-371.
  47. Carden RE, Carlin CM, Marnell F, Mcelholm D, Hetherington J, Gammell MP. Distribution and range expansion of deer in Ireland. *Mammal Rev* 2011; 41: 313-325. <https://doi.org/10.1111/j.1365-2907.2010.00170.x>
  48. Biedrzycka A, Solarz W, Okarma H. Hybridization between native and introduced species of deer in Eastern Europe. *J Mammal* 2012; 93: 1331-1341. <https://doi.org/10.1644/11-MAMM-A-022.1>
  49. Ferri M, Fontana R, Lanzi A, Armaroli E, Peloso F, Musarò C, Andina L, Allegri M, Adorni PL, Gelmini L, Barančeková M, Levrini M, De Pietri A, Berti E. Some Sika deer (*Cervus nippon*) recently hunted and spotted free-ranging in the Emilia-Romagna's region (and out of it) question the management of Italian Red deer (*Cervus elaphus*) population. *X Congresso Italiano di Teriologia, Hystrix* 2016; 27 (suppl): 100.
  50. Raganella Pelliccioni E, Riga F, Toso S. *Linee Guida per la gestione degli Ungulati: Cervidi e Bovidi*. Roma, Italy. ISPRA Press. 2013, pp 1-220 (in Italiano)
  51. Kim H, Hoelmer KA, Lee W, Kwon YD, Lee S. Molecular and morphological identification of the soybean aphid and other aphid species on the primary host *Rhamnus davurica* in Asia. *Ann Entomol Soc Am* 2010; 103: 532-543. <https://doi.org/10.1603/AN09166>
  52. Hendrichs J, Vera T, De Meyer M, Clarke AR. Resolving cryptic species complexes of major tephritid pests. *Zookeys* 2015; 540: 5-39. <https://doi.org/10.3897/zookeys.540.9656>