



# The kinetic of calcium silicate hydrate formation from silica and calcium hydroxide nanoparticles

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## ABSTRACT

**Hypothesis:** The mechanism of calcium silicate hydrate (CSH) formation, a relevant component of cement, the largest used material by mankind, is well documented. However, the effects of nano-sized materials on the CSH formation have not yet been evaluated. To this aim, a kinetic study on CSH formation via the “pozzolanic reaction” of nanosilica and calcium hydroxide nanoparticles, and in the presence of hydroxypropyl cellulose (HPC) as hydration regulator, is reported in this paper.

**Experiments:** The reagents were mixed with water and cured at 10, 20, 30 and 40 °C. The reaction kinetics was studied with differential scanning calorimetry (DSC). A Boundary Nucleation and Growth model (BNGM) combined with a diffusion-limited model was used to analyze the data, yielding induction times, reaction rates, activation energies, nucleation and linear growth rates, and the related diffusion coefficients.

**Findings:** The rate constants  $k_B$  and  $k_G$ , which are, respectively, the rate at which the nucleated boundary area transforms, and the rate at which the non-nucleated grains between the boundaries transform, increase with temperature. Their different temperature dependence accounts for the prevailing effect of nucleation over nuclei growth at progressively lower temperatures. The nucleation rate,  $I_B$ , is strongly enhanced when using nano-materials, while the linear growth rate,  $G$ , is limited by the tightly packed structure of the transforming matrix. HPC influences the kinetics between 10 and 30 °C; at 40 °C the temperature effect becomes predominant. HPC delays induction and acceleration periods, increases  $E_d(k_B)$ , and enhances the reaction efficiency during the diffusion regime, by retaining and delivering water over the matrix, thus allowing a higher water consumption in the hydration reaction of CSH.

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## 1. Introduction

Calcium silicate hydrates (CSH) include natural and synthetic phases, whose structure ranges from semi-crystalline to nearly amorphous, with variable stoichiometry [1,2]. The general formula  $(\text{CaO})_x \cdot \text{SiO}_2 \cdot (\text{H}_2\text{O})_y$  implies large variability in CSH composition and properties. Synthetic CSH is commonly obtained from the hydration of calcium silicates in cement pastes, i.e. tricalcium silicate ( $\text{C}_3\text{S}$ ) or  $\beta$ -dicalcium silicate ( $\text{C}_2\text{S}$ ), and represents the main binding phase of cement-based materials, contributing to their mechanical properties and cohesion [2–6]. The Ca/Si ratio in cement pastes usually varies from 0.4 to 3, while the water/solid ratio ranges from 0.3 to 8 [7–10].

The use of fumed silica was found to accelerate the early hydration of cement and to increase the amount of CSH [11–13]. CSH can be also obtained from the reaction of Ca salts with alkali silicates or  $\text{SiO}_2$ , in the presence of water [2,14–18]. For instance, silica-lime reaction usually yields CSH with Ca/Si ratio of 1.5, while higher ratios can be obtained from  $\text{C}_3\text{S}$  [10,19].

Smaller silica particles, such as silica fume or silica nanoparticles, can be mixed at room temperature with calcium oxide or hydroxide to obtain CSH phases via the “pozzolanic reaction”, i.e. the reaction of finely dispersed siliceous/aluminous materials with calcium hydroxide in the presence of water [7,20,21], to form compounds with cementitious properties [22]. Lin *et al.* studied  $\text{Ca}(\text{OH})_2$ -activated nano- $\text{SiO}_2$  cements as bioactive cements for bone regeneration, and found shorter setting times and lower heat release than pure  $\text{C}_3\text{S}$  with the same reaction stoichiometry [23]. The process starts with the break of the siloxane bonds due to alkaline activation. Thereafter,  $\text{Ca}^{2+}$  ions offset the charge imbalance by bonding to silanol groups forming the CSH gel. The ability of such materials to set in humid and wet environments (wa-

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ter, saliva, blood and other fluids) makes them also useful for dental biomedical applications [24]. Recently, Daniele *et al.* investigated the interactions of silica fume with  $\text{Ca}(\text{OH})_2$  aqueous suspensions stabilized by a non-ionic surfactant, which leads to the formation of CSH phases with lower energy consumption and  $\text{CO}_2$  emission with respect to traditional products, i.e. ordinary Portland cements [25].

The cementing properties of CSH have been also recently exploited in the field of cultural heritage conservation, where consolidation of carbonate-based materials, including *frescoes* and stones, can be achieved with alkaline earth metal hydroxides nanoparticles [26,27]. In a recent study, we developed a system based on nanosilica and calcium hydroxide nanoparticles was developed for the consolidation of adobe, an earthen powdery substrate of archaeological and architectural interest [28]. Adobe bricks, obtained by mixing earth with water and drying the mixture under sunlight, are particularly susceptible to wind and water erosion, and degrade easily. The “in situ” formation of CSH phases within the porosities of the treated adobe material, due to the reaction of nanosilica with  $\text{Ca}(\text{OH})_2$  nanoparticles, granted the consolidation of the adobe bricks. Hydroxypropyl cellulose (HPC), included in the consolidating mixture, performed three main actions: it acts as a viscosity-modifier, provides flexural strength, and reduces hygroscopic shrinkage during drying [28]. Following these results, we decided to study the kinetics of CSH formation starting from nanosized reactants, also in the presence of HPC. Several studies have been carried out on the mechanism of CSH formation over the years [6,11,24,29–33]. The most recent advancement in the field includes the quantitative monitoring of the hydration process in terms of transient local molecular composition using solid-state nuclear magnetic resonance [34] and the study of the distribution of hydration products in high spatial, temporal, and spectral resolution using Raman microspectroscopy [35]. However, to the best of our knowledge, this is the first kinetic study of the reaction of silica and calcium hydroxide nanoparticles. It is worth noting that, in modern cement chemistry, cellulose ethers (HCP) are commonly used to modify the rheology of the pastes. The presence of these organic polymers is reported to influence the initial phase of the hydration and its temperature dependence, without affecting the kinetic laws describing the process [32,33,36–39].

CSH formation over time was studied through a method based on Differential Scanning Calorimetry (DSC), which allows to monitor the hydration process of cement [6]. This approach quantifies the unreacted water at different hydration times through the integration of the melting peak measured by DSC, which allows to obtain the Free Water Index (FWI) [6,36,37]. Early stages of the kinetics were described by using the Boundary Nucleation and Growth Model (BNGM), originally proposed by Thomas for the study of the formation of CSH from pure  $\text{C}_3\text{S}$  [40], and recently used to study the carbonation kinetics of calcium hydroxide nanoparticles [41]. The BNGM is a geometry-controlled kinetic model that takes into account nucleation and growth processes occurring at the interface of reacting grains. In cement pastes, hydrate phases form first on the surface of the particles, then grow along the surface, eventually covering the reacting grains, and outward into the pore space, until the reacted regions coalesce, causing the paste to set. This model allows to separately evaluate the contribution of boundary CSH nucleation and growth, and accounts for the important effect of reactants’ surface area on the reaction kinetics. Later stages of the CSH formation are here described by a diffusion-limited model.

The reaction rate constants were obtained at four different temperatures, with and without HPC, and the activation energy, the nucleation rate and the linear growth rate were calculated. Fourier Transform Infrared (FT-IR) Spectroscopy, Thermogravimetric Analysis (TGA) and Scanning Electron Microscopy (SEM) were also performed to gain information on the composition and morphology of the new forming phase.

## 2. Materials and methods

### 2.1. Materials

Metal granular calcium (Aldrich, 99%) and ethanol absolute (Sigma-Aldrich, 99.8%) were used for the synthesis of calcium hydroxide nanoparticles. Monodisperse spheres of amorphous  $\text{SiO}_2$  (Levasil CS40-213 - Akzo Nobel Chemicals, 40 wt% in water, 0.2 wt% of  $\text{Na}_2\text{O}$  as stabilizer, pH = 9) were used. Hydroxypropyl cellulose (Klucel®-G - Phase Restauro, technical grade, MW: 370,000 Da, moles of substitutions: 2–4.1) was used as additive for the preparation of CSH samples. Water was purified by a Millipore Milli-Q UV system (resistivity > 18  $\text{M}\Omega\cdot\text{cm}$ ). KBr (FT-IR grade, Merck) for FT-IR analysis was used as received.

### 2.2. Samples’ preparation

Calcium hydroxide nanoparticles in ethanol, with 100–200 nm diameter and a surface area of 36  $\text{m}^2/\text{g}$  were prepared by a solvothermal synthesis [42,43]. Calcium hydroxide nanoparticles and silica nanoparticles were dried overnight at 130 °C, under  $\text{N}_2$  flow. After drying, a small amount of calcium carbonate in the dried calcium hydroxide particles was assessed with TGA, to evaluate precisely the weight of reacting Ca in the powder. An aqueous solution of HPC was prepared at a concentration of 1 wt% by stirring the mixture overnight. Calcium silicate hydrate (CSH) was prepared by mixing the two powders with a reacting Ca/Si ratio of 0.9 by mass, and then adding the HPC solution or pure water (for samples with or without the cellulosic additive, respectively), to obtain a paste with a 0.8 water/solid ratio by mass. Samples’ pH is about 12, regardless of the composition. Eight samples were prepared, four without HPC and four with HPC. The former samples are labelled with the curing temperature (e.g. ‘10’), the latter ones with the curing temperature followed by ‘\_HPC’ (e.g. ‘10\_HPC’).

### 2.3. Methods

Differential scanning calorimetry (DSC) measurements were performed with a DSC-Q2000 from TA Instruments (New Castle, USA). About 40 mg of the paste was placed in a steel pan (diameter 7.4 mm, capacity 60  $\mu\text{l}$ ) and sealed with a cover equipped with a neoprene O-ring to avoid water evaporation and carbonation of calcium hydroxide nanoparticles. The remaining part of the paste was placed in a sealed Eppendorf tube. Each sample was maintained at constant temperature in a dry block heater (10 °C, 20 °C, 30 °C, or 40 °C) and DSC pans were periodically analyzed. The measurements were carried out with the following temperature program: isothermal for 4 min at –60 °C, ramp from –60 °C to +50 °C at 5 °C/min.

The melting enthalpy of water was obtained from the endothermic peak between –35 and +20 °C. The area of the peak decreases over time, as unreacted water is gradually consumed by the reaction. The time evolution of the melting peak is reported in Fig. 1.

The melting enthalpy of water was used to calculate the Free Water Index (FWI) according to the formula [6,36]:

$$FWI = \frac{\Delta H_{\text{exp}}}{\Phi_w \Delta H_{\text{theor}}} \quad (1)$$

where  $\Delta H_{\text{exp}}$  is the experimental melting enthalpy,  $\Phi_w$  is the original weight fraction of water in the sample, and  $\Delta H_{\text{theor}}$  is the theoretical value of the melting enthalpy of water (333.4 J/g). Changes in FWI were plotted against reaction time and the curves were fitted using the boundary nucleation and growth model (BNGM) for early stages, combined with a diffusion-limited model for the later stages of the reaction.

BNGM was originally developed by Cahn for metal phases transformations [44] and later used by Thomas to describe the hydration of tricalcium silicate grains [40]. A detailed treatment of the geometrical de-

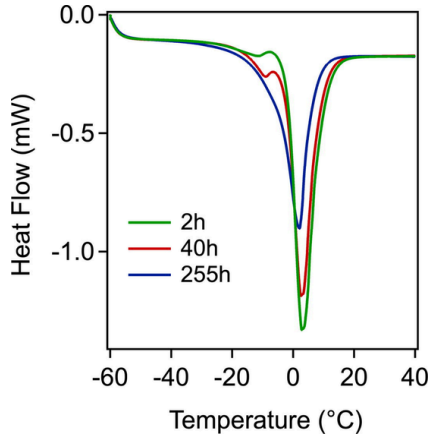


Fig. 1. Heat flow of '10\_HPC' sample at different reaction times.

rivation of this model can be found elsewhere [32,40,44–48]. Here only the basic concepts of the BNGM are reported. The derivation considers an untransformed volume containing a single planar boundary and assumes that the transformed phase only nucleates at spatially random locations on this boundary. Then, a single growing spherical region of nucleated product which intersects a plane parallel to the boundary (at a perpendicular distance  $y$ ) is considered. The integration of the area fraction of intersection, over all values of the perpendicular distance  $y$  between the plane and the boundary, gives the volume fraction of transformed phase originating from nuclei on the same grain boundary,  $X$ , which can be expressed as follows:

$$X = 1 - \exp[-2O_V^B \int_0^{Gt} (1 - \exp(-Y^e)) dy] \quad (2)$$

where  $O_V^B$  is the total area of the grain boundaries (randomly distributed in the original untransformed volume) per unit volume,  $G$  is the linear growth rate,  $t$  is the time since the start of the transformation,  $y$  is the perpendicular distance of a plane parallel to the boundary from the transforming boundary (i.e.,  $Gt$ ),  $Y^e$  is the extended area fraction of the intersection between the plane at distance  $y$  from the boundary and all regions nucleated on the grain boundary.  $Y^e$  can be mathematically described as:

$$Y^e = \frac{\pi I_B G^2 t^3}{3} \begin{cases} \left[1 - \frac{3y^2}{G^2 t^2} + \frac{2y^3}{G^3 t^3}\right] & \text{if } t > y/G \\ 0 & \text{if } t < y/G \end{cases} \quad (3)$$

where  $I_B$  is the nucleation rate per unit area of untransformed boundary.

The volume of the transformed phase depends on three covariant parameters ( $G$ ,  $I_B$ , and  $O_V^B$ ) such that Eq. (2) has only two degrees of freedom, identified by Thomas as two independent rate constants, named  $k_B$  and  $k_G$ . They are defined respectively as the rate at which the nucleated boundary area transforms, and the rate at which the non-nucleated grains between the boundaries transform (i.e. the rate at which the porosities are filled with reaction products) [40]. The linear growth rate,  $G$ , and the nucleation rate,  $I_B$ , are obtained from  $k_B$  and  $k_G$  by the following relationships:

$$k_B = (I_B O_V^B)^{1/4} G^{3/4} \quad (4)$$

$$k_G = O_V^B G \quad (5)$$

Eq. (3) can be transformed by performing a change of variable from  $y = Gt$  to  $z = y/G$ , [40]:

$$Y^e = \frac{\pi k_B^4}{3 k_G} t^3 \left[1 - \frac{3z^2}{t^2} + \frac{2z^3}{t^3}\right] \quad \text{if } t > z \quad (6)$$

and Eq. (2) becomes:

$$X = 1 - \exp[-2k_G \int_0^t (1 - \exp(-Y^e)) dz] \quad (7)$$

Equations (6) and (7) can now be solved, and the parameters  $k_B$  and  $k_G$  can be directly accessed. In fact, by introducing FWI into Eq. (7) [5], FWI curves versus time can be fitted using the BNGM:

$$FWI(t < t_d) = FWI_d + (1 - FWI_d) \exp[-2k_G \int_0^t (1 - \exp(-Y^e)) dz] \quad (8)$$

where  $FWI_d$  is the fraction of unreacted water still present at the beginning of the diffusion period, which starts at time  $t_d$ .

Fitting the time evolution of FWI using Eq. (6) and Eq. (8) returns the independent parameters  $k_B$ ,  $k_G$  and  $FWI_d$ . The values of the linear growth rate,  $G$ , and of the nucleation rate,  $I_B$ , can be then calculated through Eq. (4) and Eq. (5), using  $k_B$ ,  $k_G$ , and  $O_V^B$ . The value of  $O_V^B$  in our system is  $151 \mu\text{m}^{-1}$  and was obtained dividing the surface area of the dry silica nanoparticles powder ( $100 \text{ m}^2/\text{g}$ ) by the volume occupied by the products after complete reaction ( $0.662 \text{ cm}^3/\text{g}$ ), as reported elsewhere [40].

It is important to note that the BNG model is only valid for  $t < t_d$ . For  $t > t_d$ , the well-known diffusion-limited model is used to describe the reaction kinetics. The rate-limiting process at later stages of the reaction is the diffusion of the water through the layer of CSH surrounding the unreacted particles. Therefore, when  $t > t_d$ , the equation that describes the time evolution of FWI is:

$$FWI(t > t_d) = \left\{ FWI_d^{1/3} - \frac{(2D^2)}{\langle R \rangle} (t - t_d)^{1/2} \right\}^3 \quad (9)$$

where  $D$  is the diffusion constant, and  $\langle R \rangle$  is the average size of the calcium hydroxide nanoparticles.

In this regime, the reaction rate depends on the ability of water to diffuse through calcium hydroxide nanoparticles, whose dissolution triggers the process, i.e. the break of the siloxane groups of silica nanoparticles in alkaline environment, and the subsequent bonding with  $\text{Ca}^{2+}$  ions to form CSH [24].

BNGM equations combined with the diffusion-limited model have been therefore implemented and numerically solved using the software Igor Pro, version 6.2 (Wavemetrics Inc.).

SEM analyses were performed on samples in 1–50 days window from their preparation, to gather information on the morphology of the new forming phase. To this purpose, each sample was removed from the sealed Eppendorf tube and lyophilized prior to deposition on the stub, with the aim of hampering further hydration of the paste. For the observations, a field emission gun scanning electron microscope (FEG-SEM), SIGMA (Carl Zeiss, Germany), was used, with an acceleration potential of 2 kV and a working distance of  $\sim 2 \text{ mm}$ .

Fourier Transform Infrared (FT-IR) Spectroscopy and Thermogravimetric Analysis (TGA) were employed to get further insights on the formation of CSH: FT-IR was performed after 1 day and 50 days from samples' preparation, on the same lyophilized specimens used for SEM analyses. The analysis was performed using KBr pellets prepared with 0.01% of the collected samples, using a BIO-RAD FTS-40 spectrometer (Hercules, USA) using the following parameters: transmission mode,  $4000\text{--}400 \text{ cm}^{-1}$ , 64 scans, delay time of 300 s. The spectral data were elaborated using the Win-Ir software. TGA was carried out on the same powders using an SDT Q600 TA Instrument (New Castle, USA), operating between 25 and  $800 \text{ }^\circ\text{C}$ , at a heating rate of  $10 \text{ }^\circ\text{C}/\text{min}$ , under nitrogen flow ( $100 \text{ mL}/\text{min}$ ). For each measurement, about 5 mg of sample were placed inside an alumina pan.

The specific surface area of nanosilica and calcium hydroxide nanoparticles was determined by Brunauer-Emmett-Teller (BET) analysis of  $N_2$  isotherms, obtained with a Coulter SA 3100 Surface area analyzer (Brea, USA), with an experimental error of about 5%. From the specific surface area, the average size of nanosilica was estimated by calculating the equivalent spherical diameter, or BET particle diameter ( $d_{BET}$ ), from the equation:  $d_{BET} = 6/(q \cdot S_w)$ , where  $q$  is the density of silica ( $2.65 \text{ g/cm}^3$ ) and  $S_w$  is the specific surface area [49]. The calculated  $d_{BET}$  of nanosilica is 23 nm, which is in good accordance with the values reported in the literature, obtained from dynamic light scattering measurements [50]. Therefore, silica particles are 5–10 times smaller than calcium hydroxide nanoparticles.

### 3. Results and discussion

#### 3.1. FT-IR spectroscopy and thermogravimetric analysis

FT-IR spectroscopy was performed on samples after 1 day and 50 days from their preparation. Fig. 2 shows a comparison between samples cured at  $20^\circ\text{C}$ , without and with HPC.

In the FT-IR spectra of all the prepared samples, the characteristic signals of silicon-based compounds are present: in particular, the bands located at about  $1100 \text{ cm}^{-1}$  and  $800 \text{ cm}^{-1}$  are due to the internal Si-O-Si asymmetric and symmetric stretching, respectively; the band at  $950 \text{ cm}^{-1}$  is related to the Si-O stretching of surface Si-OH groups, and the band at  $450 \text{ cm}^{-1}$  is ascribed to the Si-O-Si asymmetric bending. It is interesting to note that, after 50 days, an inversion in the intensity of signals located at about  $1100 \text{ cm}^{-1}$  (filled triangle in Fig. 2) and  $950 \text{ cm}^{-1}$  (empty triangle) occurs. This phenomenon is due to the transformation of the siloxane groups in silanols, i.e. silica depolymerization, and is a fingerprint of CSH formation [8,23,24,51]. In fact, the process starts with the dissolution of  $\text{Ca}(\text{OH})_2$  in water, followed by the break of the Si-O-Si covalent bonds by the released OH groups; afterwards,  $\text{Ca}^{2+}$

ions bridge to the depolymerized silica and form the basic unit of CSH [24].

The narrow peak centered at  $3650 \text{ cm}^{-1}$  (filled star in Fig. 2), due to  $\text{Ca}(\text{OH})_2$  [52], shows a decrease after 50 days of curing at  $20^\circ\text{C}$ , confirming the consumption of the reactant due to the formation of CSH. The band at  $1350\text{--}1550 \text{ cm}^{-1}$  (asymmetric  $\text{CO}_3$  stretching,  $\nu_3$ ), the peak at  $876 \text{ cm}^{-1}$  (asymmetric  $\text{CO}_3$  bending,  $\nu_2$ ), and the peak at  $713 \text{ cm}^{-1}$  (symmetric  $\text{CO}_3$  bending,  $\nu_4$  (calcite)) are due to the presence of small amount of calcium carbonate [53–55] in the dried nanoparticles, which was formed prior to the preparation of the pastes (see section 2.2) and did not affect the hydration reaction.

The broad band centered at  $3400 \text{ cm}^{-1}$  and the small band at  $1650 \text{ cm}^{-1}$ , related to OH stretching and bending, can be ascribed to water, residual OH groups of silica, and HPC, when present.

Interestingly, the thermogravimetric analyses performed on the same samples (Fig. 3) show that the formation of CSH has already started after 24 h from the samples preparation, as shown by the mass loss up to  $380^\circ\text{C}$ , which is due to the decomposition of hydrate phases [37,56,57]. It is worth noting that the amount of CSH is significantly higher after 50 days from samples' preparation. The well-defined mass loss between  $400$  and  $450^\circ\text{C}$  can be ascribed to calcium hydroxide, which is not completely consumed after 50 days of curing, as also shown by the FTIR analysis. The presence of HPC seems to produce a slight increase on the decomposition temperature of both the aforementioned phases, without influencing the chemical nature of the newly formed phases [37].

#### 3.2. Differential scanning calorimetry

Experimental data obtained from FT-IR and TGA measurements clearly show the formation of CSH starting from nanosilica and calcium hydroxide nanoparticles in the selected curing condition. The kinetics of this process was therefore studied with DSC. The FWI values of the samples were plotted against the reaction time, and the data were fitted by combining the BNGM (8) and the diffusion-limited model (9). The

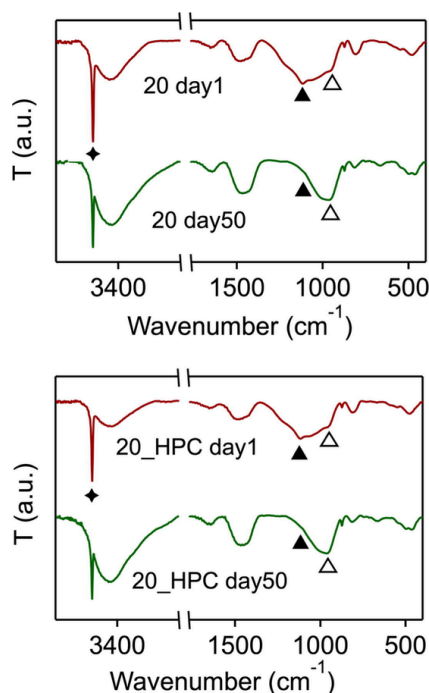


Fig. 2. FT-IR spectra collected after 1 day (red line) and 50 days (green line) on sample '20' (upper panel) and sample '20\_HPC' (lower panel). Markers refer to calcium hydroxide (filled star), siloxane groups (filled triangles) and silanols (empty triangles), respectively. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

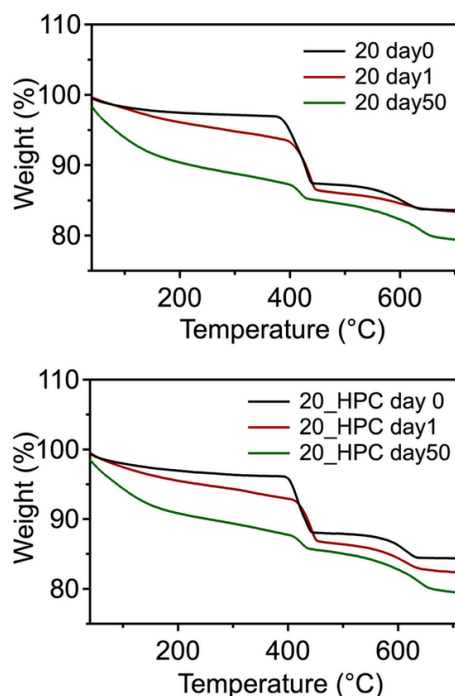


Fig. 3. TGA curves collected on pastes before curing (black line), and after 1 day (red line) and 50 days (green line) on sample '20' (upper panel) and sample '20\_HPC' (lower panel). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)



obtained parameters are reported in Table 1. Fig. 4 shows the best results for the samples at four temperatures (10 °C, 20 °C, 30 °C, and 40 °C), without and with HPC (blue and red markers, respectively). All the curves are characterized by an induction period, where FWI is almost constant, and an acceleration period, where water consumption increases. The induction time,  $t_i$ , is obtained as the intersection of the lines describing the induction and the acceleration periods. The nucleation and growth phase ends when  $t_d$  is reached; after that, the process slows down, and the diffusion regime become predominant.

**Table 1**

Fitting parameters obtained by combining the BNGM and the diffusion-limited models. The standard deviations in least significant digits are reported between brackets.

Sample name	$t_i$ (h)*	$t_d$ (h)	$FWI_d$	$k_B$ (h <sup>-1</sup> )	$k_G$ (h <sup>-1</sup> )	$k_B/k_G$	$D$ (10 <sup>-21</sup> m <sup>2</sup> h <sup>-1</sup> )
10	10 (1)	542 (3)	0.73 (1)	0.033 (3)	0.0061 (5)	5.41	0.4(3)
10_HPC	18 (1)	646 (3)	0.79 (1)	0.022 (2)	0.0053 (6)	4.15	1.1(5)
20	8(1)	245 (3)	0.82 (1)	0.041 (2)	0.012(1)	3.42	0.3(2)
20_HPC	8(1)	497 (3)	0.78 (1)	0.032 (2)	0.010(1)	3.20	0.9(5)
30	4(1)	69(2)	0.80 (1)	0.072 (3)	0.033(2)	2.18	1.3(4)
30_HPC	4(1)	150 (3)	0.77 (1)	0.067 (3)	0.028(3)	2.39	3.9(8)
40	< 4	66(2)	0.81 (1)	0.110 (3)	0.067(7)	1.64	1.8(5)
40_HPC	< 4	64(2)	0.82 (1)	0.106 (5)	0.057(6)	1.86	2.6(5)

\*the induction time  $t_i$  is obtained as the junction point of the lines describing the induction and the acceleration periods.

For all samples, the induction period is reduced from 10 h (samples cured at 10 °C) to less than 4 h (samples cured at 40 °C). The acceleration period is also reduced with increasing temperature, i.e., from 3 to 4 weeks for samples cured at 10 °C to 2–3 days for those cured at 40 °C.

The presence of HPC delays the start of the acceleration period only at 10 °C, where  $t_i$  is 1.5–2 times higher than that of the same system without the cellulosic additive.

The acceleration period is extended by the presence of HPC, with exception of the highest temperature considered (40 °C), where this effect is negligible. These findings are in agreement with literature data about the hydration kinetics of C<sub>3</sub>S, where the presence of additives generally increases the duration of the induction and acceleration steps [37,39,58].

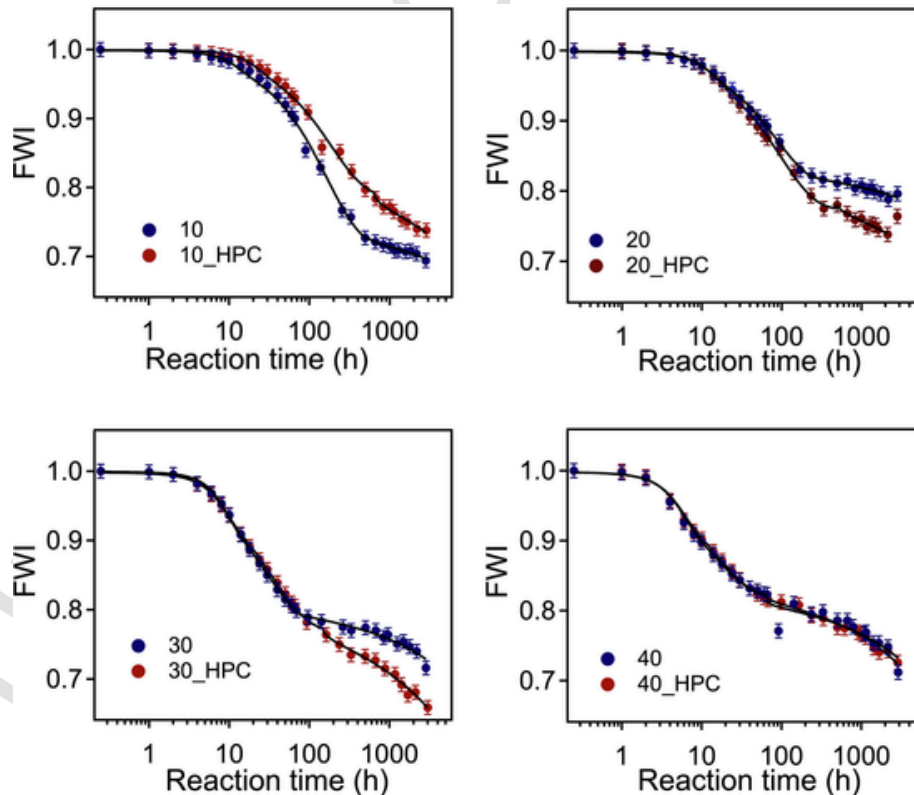
### 3.2.1. BNGM – $k_B$ , $k_G$ , and activation energies

The BNG model returned the rate constants  $k_B$ , the rate at which the nucleated boundary area transforms, and  $k_G$ , the rate at which the non-nucleated grains between the boundaries transform (i.e., the rate at which the porosities are filled with reaction products).

Regardless of the system composition,  $k_B$  and  $k_G$  increase with increasing temperature, while the presence of HPC leads to a decrease of both rate constants for all the curing temperatures investigated (see Table 1).

Two limiting cases can be described by the  $k_B/k_G$  ratio [40]: for  $k_B/k_G \gg 1$ , the boundary region is densely populated with nuclei that completely react in the process. On the contrary, when  $k_B/k_G \ll 1$ , the internal boundaries are sparsely populated with nuclei and transform at the same rate as the whole system. The latter case approaches a ‘spatially random nucleation’, which can be well described with the standard Avrami equation [59].

In our case, at 40 °C, neither nucleation nor growth is the dominant contribution to the process; at progressively lower temperatures the  $k_B/k_G$  ratio increases: in fact, both  $k_B$  and  $k_G$  rate constants decrease, but



**Fig. 4.** Kinetics of CSH formation from Ca(OH)<sub>2</sub> nanoparticles and SiO<sub>2</sub> nanoparticles without HPC (blue markers) and with HPC (red markers). The BNGM fittings combined with diffusion-limited model are reported on each data set as continuous black lines. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

it is worth noting that the decrease of  $k_G$ , which is almost halved with every curing temperature step, is significantly faster than the decrease of  $k_B$ . In other terms, with decreasing temperature, a progressive increase in the contribution of nucleation, which is mainly described by the  $k_B$  term, takes place. This is clearly shown by the Arrhenius plot,  $\ln(k)$  versus  $1/T$ , which describes the temperature dependence of the two rate constants (Fig. 5).

In Fig. 5, the blue markers are used for systems without HPC, while the red markers are used for systems containing the cellulosic additive. All the Arrhenius plots are linear across the investigated temperature range, with a  $R^2 \geq 0.97$ , as indicated in Table 2. The higher dependence on temperature of  $k_G$  with respect to  $k_B$ , shown by the Arrhenius plots, is confirmed by the calculated activation energies ( $E_a$ ) reported in Table 2.

The lower temperature dependence of  $k_B$  can be explained by taking into account the high surface area of reacting nanoparticles: in fact, the high extension of boundaries available for nucleation probably makes the effect of curing conditions, i.e., temperature, less relevant in the initial phase of the reaction.

Interestingly, HPC increases the temperature dependence of  $k_B$ , as shown by the increase in  $E_a(k_B)$ . This might be explained by the absorption of the cellulosic additive on the reacting nanoparticles' surface, which decreased the amount of available boundary area, that results in

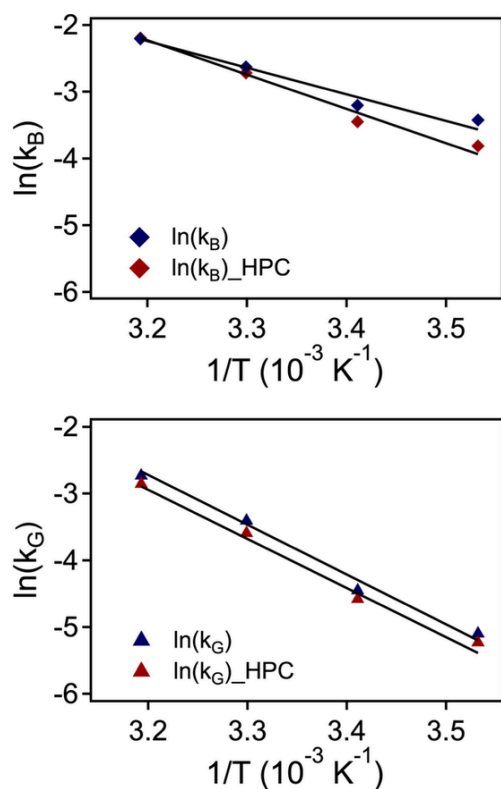


Fig. 5. Arrhenius plot showing the temperature dependence of the rate constants,  $k_B$  (upper panel) and  $k_G$  (lower panel), without HPC (blue markers) and with HPC (red markers). The experimental data were fitted with linear regressions (black lines). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Table 2

$R^2$  of the Arrhenius plots and activation energies calculated for  $k_B$  and  $k_G$ .

	$R^2(k_B)$	$R^2(k_G)$	$E_a(k_B)$ (kJ/mol)	$E_a(k_G)$ (kJ/mol)
without HPC	0.97	0.99	31(4)	60(4)
with HPC	0.98	0.99	41(4)	60(4)

a significantly increased effect of the environmental conditions on the rate at which the nucleated boundary area transforms.

On the contrary, HPC does not affect  $E_a(k_G)$ . In other terms, the presence of the cellulosic additive seems not to influence the rate at which the porosities are filled with reaction products, which is mainly described by the  $k_G$  term.

### 3.2.2. BNGM – $I_B$ and $G$

Table 3 shows the values of  $I_B$  (nucleation rate) and  $G$  (linear growth rate) for each sample, calculated from  $k_B$  and  $k_G$ , using 4 and 5, in which the total area of the grain boundaries per unit volume,  $O_V^B$ , is introduced.

The trend of  $G$  reflects that of  $k_G$ , i.e., it increases with increasing temperature and is reduced by the presence of HPC regardless of the temperature. On the contrary,  $I_B$  decreases with increasing temperatures.

It is interesting to note that the  $I_B$  values of our systems are orders of magnitude higher than those reported in the literature for CSH formation in cement pastes, which generally range from  $10^{-4}$  to  $10^{-5}$  [32,39,40,58]. This is due to the fact that both silica and calcium hydroxide nanoparticles' have a high surface area, respectively of  $100 \text{ m}^2/\text{g}$  and  $36 \text{ m}^2/\text{g}$ , which result in an impressive increase in the number of nuclei in the system. A scheme of the time evolution of the formation of CSH due to the reaction of silica and calcium hydroxide nanoparticles is shown in Fig. 6.

On the contrary, the linear growth rates obtained for our systems are significantly lower than those reported in literature for CSH formation in cement [32,39,40,58]. Again, this might be explained by the nanosized particles used as reactants, which probably arrange in a packed structure that limits the outward growth into the pore space, resulting in low  $G$  values.

Overall, the results obtained using the BNGM point out the combined effect of nanoparticles, curing conditions and HPC on the kinetics of CSH formation from nanosilica and calcium hydroxide nanoparticles. To summarize, HPC extends the induction period, at  $10^\circ\text{C}$ , and the acceleration period, between  $10$  and  $30^\circ\text{C}$ . At  $40^\circ\text{C}$ , temperature seems to be the leading factor and the effect of the additive appears negligible. This might be explained by the fact that at about  $40^\circ\text{C}$  HPC displays a cloud point [60], which leads to the precipitation of the polymer that is not anymore available to interact with the reactants. HPC has also a major influence on the early stages of the reaction, as it probably reduces the available surface area of reacting nanoparticles, and hence modifies the temperature dependence of the rate constant  $k_B$ . The obtained nucleation rates, which are significantly higher than those measured on cement pastes, can be ascribed to the nanosized materials, whose high surface area favors CSH formation. On the contrary, low linear growth rate are due to a tightly packed structure created by nanoparticles in which the growth of nucleated material in the pore space is partially hampered.

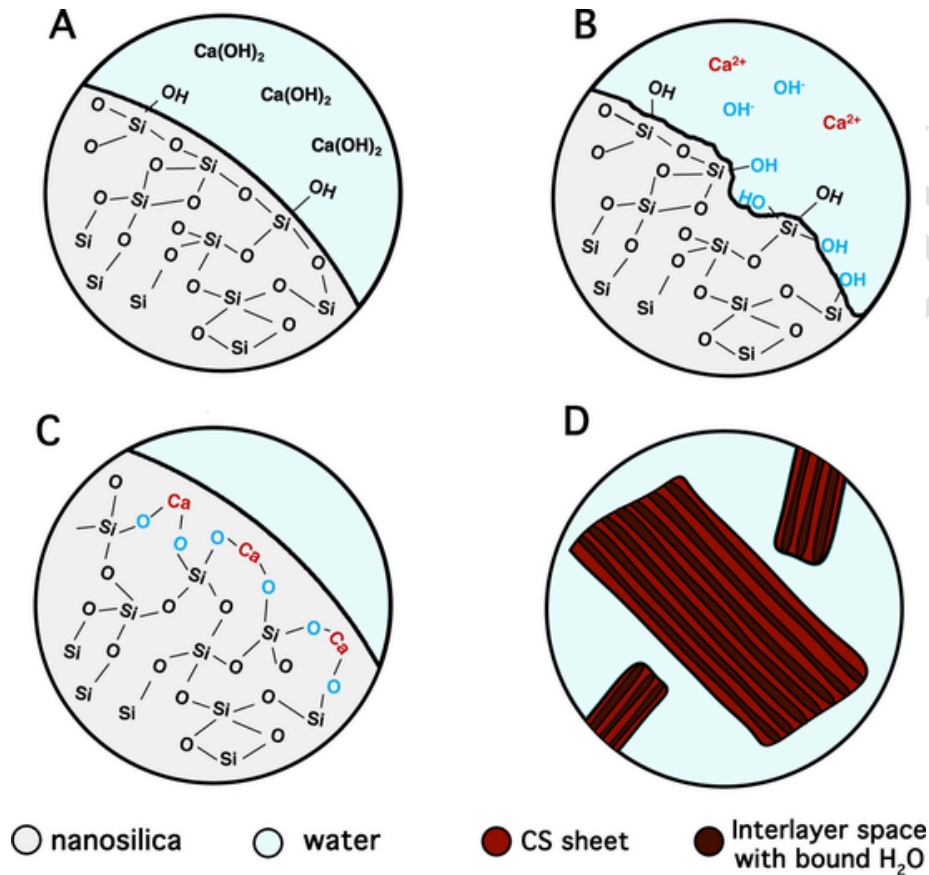
### 3.2.3. Diffusion period

The beginning of the diffusion period is delayed by the decrease in curing temperature and by the presence of HPC. The diffusion coeffi-

Table 3

Nucleation rate,  $I_B$ , and linear growth rate,  $G$ , obtained from the BNG model.

Sample name	$I_B (\mu\text{m}^{-2} \cdot \text{h}^{-1})$	$G (\mu\text{m}/\text{h})$
10	$11(6) \cdot 10^4$	$40(3) \cdot 10^{-6}$
10_HPC	$35(27) \cdot 10^3$	$35(4) \cdot 10^{-6}$
20	$39(21) \cdot 10^3$	$77(8) \cdot 10^{-6}$
20_HPC	$22(10) \cdot 10^3$	$68(8) \cdot 10^{-6}$
30	$17(6) \cdot 10^3$	$22(1) \cdot 10^{-5}$
30_HPC	$21(11) \cdot 10^3$	$18(2) \cdot 10^{-5}$
40	$12(5) \cdot 10^3$	$43(5) \cdot 10^{-5}$
40_HPC	$18(9) \cdot 10^3$	$38(4) \cdot 10^{-5}$



**Fig. 6.** Time evolution of CHS formation. Panel A shows a portion of a grain of nanosilica immersed in the reaction media, together with alkaline nanoparticles. The reaction starts with the dissolution of  $\text{Ca}(\text{OH})_2$  in water followed by a depolymerization of silica, to form silanol groups (panel B). Afterwards,  $\text{Ca}^{2+}$  ions bridge to the depolymerized silica and form the basic unit of CSH (panel C). The use of nanosilica, whose surface area is  $100 \text{ m}^2/\text{g}$ , significantly increase the rate of the process. Panel D shows the system at the nanoscale: the primary units of CSH are polydisperse multilayer disks with calcium silicate (CS) sheets and interlayer space with bound water. These primary units organize themselves to form fractal structures below the micrometer length-scale, as reported in the literature [33].

cient, D, (last column of Table 1), obtained from fitting the later stages of the kinetics, is related to the easiness for free water to reach the unreacted nanoparticles. As indicated above, CSH formation occurs through the break of the siloxane groups in alkaline environment, followed by the bridging of  $\text{Ca}^{2+}$  ions to the depolymerized silica [24]. Therefore, the diffusion regime starts when water needs to diffuse through the growing CSH matrix to reach unreacted hydroxide particles. Due to the fact that reactants must move through the nanopores within a continuous product layer around the unreacted particles [39,40], diffusion rates in cement are significantly smaller than those measured in bulk solutions. Moreover, the diffusion coefficients, obtained starting from nanosized reactants, are smaller than those commonly obtained in standard cement pastes ( $10^{-21}$  vs  $10^{-15} \text{ m}^2/\text{h}$ ). This is probably due to nanoparticles that create a denser-packed matrix with respect to micron-sized grains included in standard pastes, leading to a less porous structure, in which the diffusion of water is very low. Finally, it is worth noting that D values tend to increase with increasing temperature, which can be explained by the increased mobility of water in such environmental conditions.

Interestingly, the presence of HPC leads to the same overall effect, significantly increasing the D values up to a curing temperature of  $30^\circ\text{C}$ . In cement chemistry, cellulosic additives act as regulators of water release, due to their carrying capacity and hydrophilicity, and water results homogeneously distributed and more prone to react [37,39,58, 61]. Here a similar effect took place: the presence of HPC leads to a more efficient diffusion of water in the transforming gel matrix, resulting in a higher water consumption in the later stages of the reaction, which starts from  $t_d$ . It is worth noting that at  $40^\circ\text{C}$ , the presence of cel-

lulose only slightly enhances the D parameter, probably because temperature becomes the driving force for the CSH formation as a result of the lower solubility of HPC at higher temperature.

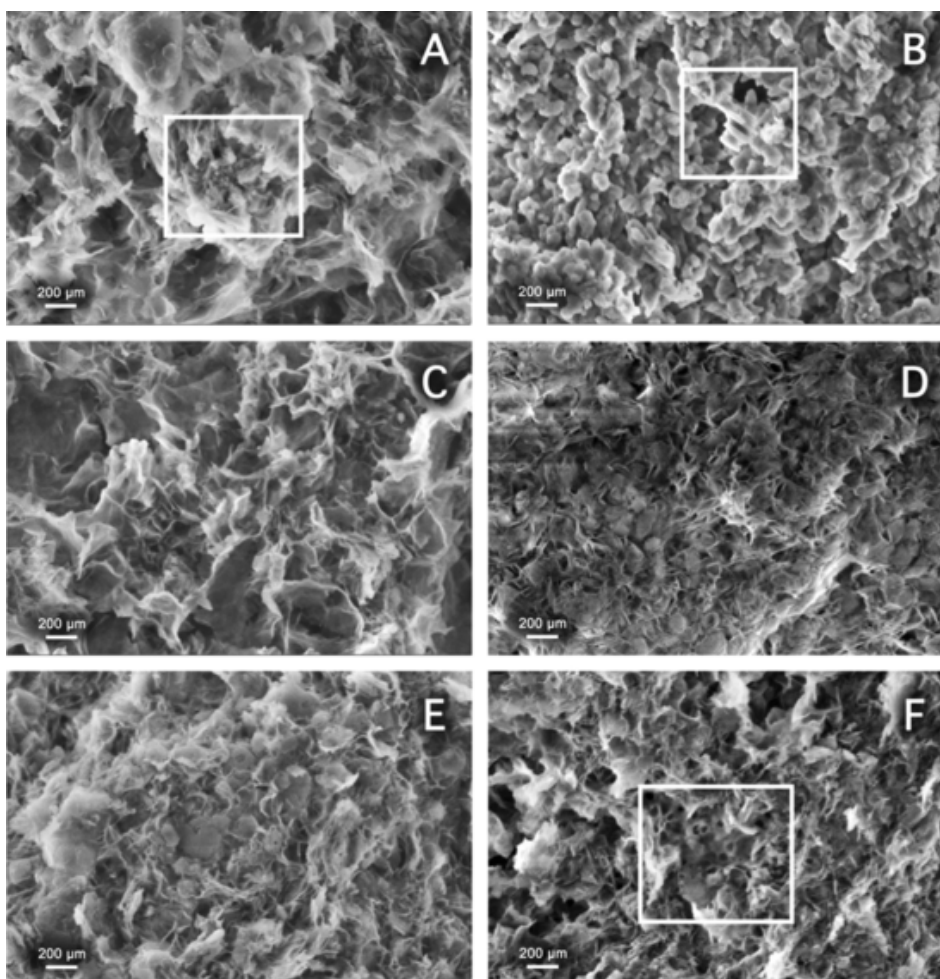
### 3.3. Scanning Electron Microscopy

Fig. 7 reports SEM analysis after 1 day and 50 days from sample preparation. Fig. 7 A-D show samples cured at  $20^\circ\text{C}$ , without cellulose (Fig. 7A and 7C, after 1 day and 50 days, respectively) and with cellulose (Fig. 7B and 7D, after 1 day and 50 days, respectively), while Fig. 7E and 7F show the samples cured at  $30^\circ\text{C}$ , without and with cellulose, respectively.

After 1 day (acceleration phase), unreacted nanoparticles are still present, as shown in Fig. 7A (see the white box highlighting a group of  $\text{SiO}_2$  spheres) and Fig. 7B (see the white box highlighting stacked platelets of  $\text{Ca}(\text{OH})_2$ ). After 50 days, CSH is the dominant phase, with the typical morphology of crumpled foils that characterizes alkaline activated silica [9,23]. In the presence of HPC (Fig. 7D), the classical CSH porous network displayed in Fig. 7C is partially covered up, due to the new forming phase embedded in the cellulosic matrix. At this stage, unreacted nanoparticles of silica and calcium hydroxide are still present, in agreement with the fact that the reaction is in the diffusion regime and that the FWI is still decreasing (see Fig. 4).

A similar effect is visible in samples cured at  $30^\circ\text{C}$ , after 50 days (Fig. 7E and 7F). Interestingly, the footprint left by the reacted silica spheres in the matrix is clearly shown in Fig. 7F, as highlighted by a white box in the center of the image.





**Fig. 7.** SEM images (Mag = 100 kX) of (A) sample '20' after 1 day (WD = 2 mm), (B) sample '20\_HPC' after 1 day (WD = 2 mm), (C) sample '20' after 50 days (WD = 2.9 mm), (D) sample '20\_HPC' after 50 days (WD = 2.6 mm), (E) sample '30' after 50 days (WD = 2.6 mm), (F) sample '30\_HPC' after 50 days (WD = 2.7 mm).

#### 4. Conclusions

CSH was obtained starting from nanosilica and calcium hydroxide nanoparticles, and in the presence of hydroxypropyl cellulose, at different curing temperature. FT-IR and TGA measurements confirmed the formation of CSH, which has the typical morphology of crumpled foils that characterizes alkaline activated silica, as shown by SEM pictures. The reaction kinetics of CSH formation was studied by means of DSC, applying the BNGM combined with a diffusion-limited model.

The rate constants  $k_B$  and  $k_G$ , obtained from the BNGM, increase with temperature up to 40 °C when none of the two contributions is predominant. The contribution of nucleation prevails with decreasing temperature, according to the higher dependence on temperature of  $k_G$  with respect to  $k_B$ . The lower temperature dependence of  $k_B$  can be explained by taking into account the high surface area of reacting nanoparticles: in fact, the high extension of boundaries available for nucleation probably makes the effect of curing conditions, i.e., temperature, less relevant in the initial phase of the reaction.

The addition of a cellulosic additives delays the induction and acceleration periods, especially at low temperatures [37,39,58]. The presence of HPC decreases the rate at which the nucleated boundary area transforms ( $k_B$ ), since it reduces the available surface area of the transforming nanoparticles. On the contrary, the rate at which the porosities are filled with the reaction products ( $k_G$ ) is not significantly affected by the presence of the additive. Overall, HPC increases the temperature dependence of  $k_B$  ( $E_a = 41 \pm 4$  kJ/mol), but has no impact on  $k_G$ .

The use of nanosized reagents significantly influences the reaction kinetics in terms of  $I_B$  and  $G$  with respect to standard cement pastes [32, 39,40,58]. In fact,  $I_B$  values are several orders of magnitude higher than those reported in the literature. *This effect can be explained by taking into account the higher reactivity of nanomaterials, which enhances the nucleation rate.* On the contrary, nanoparticles generate a packed structure limiting the outward growth of the reacting species into the pore space between the particles, because of steric hindrance, accounting for smaller  $G$  values in the BNGM kinetic model.

During the diffusion regime, which starts at  $t_d$ , the presence of HPC increases the water consumption, specially between 10 and 30 °C. This is due to the fact that HPC is able to retain and distribute in a controlled way water over the whole matrix, as reported for cement pastes [37,39, 58,61]. At 40 °C the effect is no longer detectable, probably due to the precipitation of HCP aggregates, which makes temperature the driving force for CSH formation, regardless of the composition of the system. Overall, the addition of silica nanoparticle to calcium hydroxide nanoparticles consistently improves, as compared to the classic CSH occurring in the micrometer size cement powders, the nucleation process allowing the control and the tuning of CSH formation.

In modern cement chemistry, the reaction of magnesium oxides with silica and water is exploited to produce an environmentally convenient alternative to standard binders based on CSH [62,63]. Future developments of our research foresee the study of the kinetic of magnesium silicate hydrate (MSH) formation in the presence of several additives, including cellulose derivatives and phosphates.



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## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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