A severe outbreak of influenza A(H1N1)pdm09 infection in seven children (median age: 52 months) occurred between December 2023 and January 2024 in Tuscany, Italy. Clinical presentation ranged from milder encephalopathy to acute necrotizing encephalopathy (ANE) with coma and multiorgan failure; one child died. This report raises awareness for clinicians to identify and treat early acute encephalopathy caused by H1N1 influenza and serves as a reminder of severe presentations of influenza in young children and the importance of vaccination.

Influenza infection in young children can present with severe complications, especially during the peak season [1]. The risk of severe outcome can be exacerbated by low rates of influenza vaccination in children [2]. Here, we describe an outbreak of influenza A(H1N1)pdm09 with severe neurological involvement in seven children admitted between December 2023 and January 2024 to Meyer Children's Hospital (Florence, Italy), a tertiary care referral centre in Tuscany with a catchment area of ca 600,000 children.

Clinical characteristics

All seven patients (median age: 52 months (interquartile range (IQR): 25–70), n = 5 males) presented with febrile illness and encephalopathy. Five children were previously healthy, while two had experienced seizures in the past. Four patients were directly admitted to the paediatric intensive care unit (PICU), and three were admitted to the hospital wards. Three PICU patients were diagnosed with acute necrotizing encephalopathy (ANE) [3]. Clinical characteristics are summarised in Table 1 and brain magnetic resonance imaging (MRI) findings are shown in Figure 1.

None of the children had been vaccinated for influenza. Six of seven patients received oral oseltamivir [4] for 5–12 days upon admission.

Immunomodulatory treatment and outcome in severe cases

A previously healthy child (PICU 1) was transferred to our PICU on the fourth day of fever because of liver failure and severe coagulopathy with spontaneous gastrointestinal bleeding and multiorgan failure with rapid neurological deterioration. The patient was given high dose methylprednisolone (30 mg/kg once daily) and received only two doses before the child died on the fifth day of illness.

A previously healthy child (PICU 2) was transferred to our PICU because of convulsive febrile status epilepticus. The child was intubated and put on a midazolam drip to treat ictal-interictal continuum/non-convulsive status epilepticus. Immunomodulation started at 48 h of illness and consisted of dexamethasone 1 mg/kg/day for 2 days, high dose methylprednisolone (30 mg/kg once daily) for 5 days, intravenous immunoglobulin (IVIg) 1 g/kg/day for 3 days, anakinra 10 mg/kg/day for 7 days, and one dose of tocilizumab 9 mg/kg. The child was extubated on day 7 of mechanical ventilation and required gastrostomy tube placement, but no tracheostomy was necessary. Neurological examination on
<table>
<thead>
<tr>
<th>Patient</th>
<th>Age (months)</th>
<th>Presentation (days after fever onset)</th>
<th>Video EEG (days after fever onset)</th>
<th>Brain MRI (days after fever onset)</th>
<th>Antiseizure medications</th>
<th>Immunomodulator (days of treatment)</th>
<th>Oseltamivir (days of fever when started)</th>
<th>Relevant laboratory results</th>
<th>Outcome (days after fever onset)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PICU 1</td>
<td>52</td>
<td>Coagulopathy (2); multiorgan failure (3); encephalopathy (4)</td>
<td>Moderate slow (4); suppression (3)</td>
<td>Diffuse basal ganglia, thalami, brainstem injury (4)</td>
<td>None</td>
<td>MTP (2)</td>
<td>Yes (4)</td>
<td>PLT: 231x10^9/L, AST: 11, 292 IU/L, ALT: 4, 398 IU/L, Cr: 3.49 mg/dL, INR: 2.89, aPTT: 106 s, CRP: 3.74 mg/dL, PCT: 290.0 ng/mL, Fib: 128 mg/dL</td>
<td>Death (5)</td>
</tr>
<tr>
<td>PICU 2</td>
<td>11</td>
<td>Febrile status epilepticus (5)</td>
<td>Ictal-ictal continuum (1–3); burst suppression (4–7)</td>
<td>Negative (2); diffuse white matter, thalami injury (4)</td>
<td>DZP, MDZ, LCM, PHT</td>
<td>DEX (1), MTP (5), IVIg (5), ANK (7), TOC (1)</td>
<td>Yes (1)</td>
<td>AST: 2,968 IU/L, ALT: 2,212 IU/L, Fib: 166 mg/dL, DD: 24,382 ug/L, INR: 2.05, aPTT 53 s, CRP: 1.81 mg/dL, PCT: 42.6 ng/mL, CSF: 4x1/ul, Glu: 105 mg/dL, TP: 35 mg/dL</td>
<td>Gastrostomy tube (no tracheostomy), minimally conscious state, autonomic storms, spastic quadriplegia (63)</td>
</tr>
<tr>
<td>PICU 3</td>
<td>77</td>
<td>Encephalopathy (1)</td>
<td>Mild slow (1–2); frontal discharges (6)</td>
<td>Bilateral temporal, thalami, pons injury (1); evolution of injury (7)</td>
<td>CBZ</td>
<td>MTP (5), IVIg (3), TOC (1), PLEX (3)</td>
<td>Yes (1)</td>
<td>AST: 64 IU/L, ALT: 45 IU/L, CRP: 6.06 mg/dL, PCT: 51.7 mg/mL, CSF WBC: 352x1/ul, Glu: 58 mg/dL, TP: 79 mg/dL</td>
<td>Mild dyspraxia (48)</td>
</tr>
<tr>
<td>PICU 4</td>
<td>86</td>
<td>Complex febrile seizures (3)</td>
<td>Moderate slow (3–4)</td>
<td>N/A</td>
<td>MDZ, PHT, CLB</td>
<td>None</td>
<td>Yes (3)</td>
<td>N/A</td>
<td>Full recovery (8)</td>
</tr>
<tr>
<td>Wards 1</td>
<td>63</td>
<td>Complex febrile seizures (2)</td>
<td>Mild slow (1)</td>
<td>Mild bilateral post white matter hyper (4)</td>
<td>CLO</td>
<td>None</td>
<td>Yes (2)</td>
<td>N/A</td>
<td>Full recovery (6)</td>
</tr>
<tr>
<td>Wards 2</td>
<td>16</td>
<td>Cough (1); febrile status epilepticus (2)</td>
<td>Focal bilateral occipital slow (3); normal (8)</td>
<td>Negative (4)</td>
<td>DZP, MDZ, PHT</td>
<td>DEX (3)</td>
<td>Yes (2)</td>
<td>CSF WBC: 1x1/ul, Glu: 42 mg/dL, TP: 11 mg/dL</td>
<td>Full recovery (6)</td>
</tr>
<tr>
<td>Wards 3</td>
<td>34</td>
<td>Complex febrile seizures (3)</td>
<td>Mild encephalopathy (4); normal (9)</td>
<td>Negative (11)</td>
<td>None</td>
<td>None</td>
<td>No</td>
<td>CSF WBC: 1x1/ul, Glu: 50 mg/dL, TP: 14 mg/dL</td>
<td>Full recovery (7)</td>
</tr>
</tbody>
</table>

aPTT: activated partial thromboplastin time; ALT: alanine aminotransferase; ANK: anakinra; AST: aspartate aminotransferase; CBZ: carbamazepine; CLB: clonazepam; CLO: clonazepam; Cr: creatinine; CRP: C-reactive protein; CSF: cerebrospinal fluid; DD: d-dimer; DEX: dexamethasone; DZP: diazepam; EEG: electroencephalogram; Fib: fibrinogen; Glu: glucose; INR: international normalised ratio; IVIg: intravenous immunoglobulin; LCM: lacosamide; MDZ: midazolam; MRI: magnetic resonance imaging; MTP: methylprednisolone; N/A: not applicable; PCT: procalcitonin; PHT: phenytoin; PLEX: plasma exchange; PLT: platelets; s: seconds; TOC: tocilizumab; TP: total protein; WBC: white blood cell.

Normal ranges for the laboratory results are as follows: ALT (5–19 IU/L), aPTT (29–38 s), AST (5–41 IU/L), Cr (0.30–0.50 mg/dL), CRP (0–0.50 mg/dL), CSF Glu (50–80 mg/dL), CSF TP (20–50 mg/dL), CSF WBC (1x1/ul), DD (500 ug/mL), Fib (200–400 mg/dL), INR (0.92–1.14), PCT (< 0.5 ng/mL), PLT (210–590x10^9/L), Fib: 200–400 mg/dL, INR: 0.92–1.14, PCT: < 0.5 ng/mL, PLT: 210–590x10^9/L.

*Traumatic tap from needle insertion, with bleeding into the subarachnoid space, artificially increasing the white blood cell count.*
the 63rd day of illness showed a severe outcome with minimally conscious state, autonomic storms, and severe dystonic spastic quadriplegia.

A previously healthy child (PICU 3) was admitted to our PICU because of encephalopathy after 2 days of fever. All immunomodulatory drugs were started within 24 h of illness and consisted of high dose methylprednisolone (30 mg/kg once daily) for 5 days, followed by an oral taper over 1 month, IVIg 1 g/kg/day for 3 days, plasma exchange every other day for 3 sessions, and tocilizumab (9 mg/kg/dose) once weekly for 1 month. The child was extubated after 6 days of mechanical ventilation. Neurological examination on the 48th day of illness showed mild dyspraxia as the only manifestation, highlighting good clinical outcome.

The fourth child (PICU 4) admitted to the PICU was extubated and transferred out of the PICU in less than 48 h. No immunomodulation was pursued. The child made a full recovery.

Two patients admitted to the wards did not receive immunomodulatory treatment; the third patient was given dexamethasone 1 mg/kg/day for 3 days. All three children experienced full recovery before discharge.

Genotyping and epidemiological data

According to our institutional surveillance protocol, all children admitted with fever and respiratory or neurological symptoms are tested for viral infection. A nasopharyngeal swab taken from all patients tested positive for influenza A(H1N1)pdm09. Viral PCR was negative in the cerebrospinal fluid (CSF) of the four patients who underwent a lumbar puncture. Patients were tested for other respiratory viruses: influenza A(H1N1), influenza A(H3N2), influenza B, respiratory syncytial virus (RSV), rhinovirus, adenovirus, parainfluenza virus types 1–3, metapneumovirus, SARS-CoV-2 and bocavirus; no co-infections were identified.

Sequencing of the influenza A(H1N1)pdm09 strains from swab samples collected from the three most severe PICU patients showed that all strains belonged to clade 6B.1A.5a.2a. We did not identify any amino acid substitutions of particular interest in patients with ANE (Table 2).

Epidemiological data recorded at our institution from 1 November 2023 to 20 March 2024 for children admitted with fever and respiratory or neurological symptoms revealed 251 A(H1N1)pdm09-positive nasopharyngeal swabs (Figure 2), of which 226 were admitted during the study period between week 49 2023 and week 4 2024.

Discussion

In the post-COVID-19 pandemic seasons, influenza-like illness (ILI) incidence has increased in the European Union/European Economic Area (EU/EEA) [5]. In Italy, during the 2022/23 season, ILI peaked early, during
the week ending with 27 November 2022, driven predominantly by influenza A (96.3% H3N2) and RSV [6]. The 2023/24 season has been characterised by high intensity activity, a later ILI peak between week 49 2023 and weeks 2–3 2024 and a dominance of A(H1N1)pdm09 viruses. These observations reflect the ILI peak identified through molecular surveillance in our institution, with A(H1N1)pdm09 reaching 92.4% of circulating viruses during week 48 [7]. The 6B.1A.5a.2a clade starting from week 40 of 2023 was the most prevalent type A virus [5]. At the beginning of January 2024, the United States (US) Centers for Disease Control and Prevention (CDC) raised an alert on severe influenza illness among children in the US, mainly by influenza A viruses. However, no subtyping was performed, and no description of symptoms or neurological involvement is available [8].

Paediatric mortality data in Italy for the 2023/24 season are not yet available. Our institutional data show that influenza cases with severe neurological involvement comprise 3% (7/226) of all children admitted during the study period for fever associated with respiratory or neurological symptoms. Because our institution used a different surveillance protocol after the COVID-19 pandemic, we are not able to compare the present data with pre-pandemic data concerning neurological involvement in children admitted for ILI. However, no case of ANE was recorded at least since 2017.

Based on a comparison between the genotype of identified virus and PB2A/Victoria/4897/2022(H1N1) available on the NEXCLADE dataset (https://clades.nextstrain.org), we did not identify any amino acid substitutions of particular interest (in PB2, HA or in other genes) in our cases of ANE. The identified mutations do not belong to the category of mutations known to alter the virulence of the virus, cause strong drug resistance or reverse the effects of the premature STOP codon in the PB1-F2 gene of pandemic H1N1 (https://flusurver.bii.a-star.edu.sg).

Acute necrotizing encephalopathy is a rare and very severe neurological disorder characterised by inflammation and necrosis in the brain. The global incidence of ANE is unknown, and most cases have been described in Asia, with a peak between age 18 months and 6 years [3]. The pathophysiology would result from a ‘cytokine storm’ in response to the viral infection, with increased vascular permeability, neuronal cytotoxicity and eventually neuronal cell death [3]. There are no data to suggest that the influenza A(H1N1) virus has a direct neurotoxicity effect based on findings from pathological and cerebrospinal fluid investigations [3] that we observed in our patients. There is no consensus on the best treatment strategy for ANE [9]. Anecdotally, different drugs, including high-dose steroids [10], plasma exchange [11], anakinra [12] and tocilizumab [13] have been used. Of our three most severe cases, the best outcome was observed in the child who received aggressive immunomodulation within 24 h of the onset of illness. Further studies are needed to define optimal treatment strategies based on higher level of evidence.

Interim 2023/24 A(H1N1)pdm09 influenza vaccine effectiveness in children was high (62–85%) in Canada [14] and Europe [15]. Although the risk for unfavourable outcomes in children with influenza is known, influenza vaccination coverage in children in Italy remains low (less than 10% in children under 4 years nationwide; ca 12% in the Tuscany region). The Italian Ministry of Health, following recommendations from the World Health Organization, recommends influenza vaccination in all children aged 6 months to 6 years. A poor influenza vaccine uptake could be the consequence of parental beliefs that influenza is a not-serious illness [16] and paediatricians’ belief that influenza vaccine has low effectiveness against mild to moderate illness [17]. However, influenza vaccine reduces the risk of critical illness in children [2], even in seasons when vaccine-mismatched influenza viruses circulate.

**Conclusion**

In summary, we observed an outbreak of life-threatening influenza A(H1N1)pdm09 illness in young children, resulting in a range of outcomes including severe neurological sequelae and death. Overall, these observations amounted to 3% of all patients hospitalised for fever and respiratory or neurological symptoms at our
Figure 2
Influenza A(H1N1)pdm09 institutional surveillance data, Meyer Children’s Hospital, Florence, Italy, 2023/2024 season

A. Total A(H1N1)pdm09 cases at Meyer Children’s Hospital

B. Distribution of A(H1N1)pdm09 cases in Italy vs Meyer Children’s Hospital

Ethical statement
For this report, we obtained written informed consent from the legal guardians and followed the Declaration of Helsinki guidance.

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Data availability
Data will be made available upon reasonable request by accredited researchers.

Use of artificial intelligence tools
None declared.

Conflict of interest
None declared.

Authors’ contributions
Luca Bartolini, MD, FAAN, FAES: study design, data collection and analysis/interpretation, manuscript writing and reviewing for intellectual content. Silvia Ricci, MD: data interpretation, manuscript writing and reviewing for intellectual content. Chiara Azzari, MD, PhD: data interpretation, manuscript writing and reviewing for intellectual content. Maria Moriondo, PhD: data analysis and interpretation, manuscript writing and reviewing for intellectual content. Francesco Nieddu, PhD: data analysis and interpretation, manuscript writing and reviewing for intellectual content. Manuela L’Erario, MD: data collection, manuscript writing and reviewing for intellectual content. Zaccaria Ricci, MD: data collection, manuscript writing and reviewing for intellectual content. Gabriele Simonini, MD: data interpretation, manuscript writing and reviewing for intellectual content. Marzia Mortilla, MD: data interpretation, manuscript writing and reviewing for intellectual content. Giuseppe Indolfi, MD: data interpretation, manuscript writing and reviewing for intellectual content. Carlotta Montagnani, MD: data interpretation, manuscript writing and reviewing for intellectual content. Elena Chiappini, MD: data interpretation, manuscript writing and reviewing for intellectual content. Luisa Galli, MD: supervision of research conduct, data interpretation, manuscript writing and reviewing for intellectual content. Renzo Guerrini, MD, FRCP, FAES: supervision of research conduct, data interpretation, manuscript writing and reviewing for intellectual content.

References


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