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PER2 C111G polymorphism, cognitive reserve and cognition in subjective cognitive decline and mild cognitive impairment: a 10-year follow-up study

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Background and purpose: *CLOCK* and *PER2* genes have been implicated in sleep–wake cycle alterations and neurodegenerative diseases. Our aim was to evaluate the effect of *CLOCK* T3111C and *PER2* C111G on cognitive functioning in subjective cognitive decline (SCD) patients and mild cognitive impairment (MCI) patients at the baseline of a longitudinal study, and the effect of these two polymorphisms on the progression to Alzheimer's disease (AD) of the two groups.

Methods: Sixty-eight subjects (41 SCD and 27 MCI) who underwent clinical evaluation, neuropsychological assessment, *CLOCK* and *PER2* genotyping at baseline and neuropsychological follow-up every 2 years for a mean time of 10 years were included. Subjects who developed AD (SCD-c and MCI-c) and non-converters (SCD-nc, MCI-nc) were considered.

Results: *CLOCK* T3111C was detected in 47% of cases (21 SCD, 11 MCI) and *PER2* C111G in 19% of cases (eight SCD and five MCI). *PER2* G carriers presented lower premorbid intelligence score (P = 0.049), fewer years of education (P = 0.007) and a lower frequency of family history of AD (P = 0.04) than G non-carriers. MCI *PER2* G carriers had worse performance in tests assessing memory, executive function, language and visuospatial abilities at baseline. During follow-up, two SCD and 15 MCI subjects progressed to AD: both of the SCD-c subjects presented the *PER2* G allele, while none of the SCD *PER2* G non-carriers converted to AD (P = 0.003).

Conclusion: *PER2* seems to have a role in cognitive reserve and cognition in SCD and MCI patients. Nevertheless, further studies are needed to assess the role of *PER2* C111G on the risk of progression to AD.

Introduction

Alzheimer's disease (AD) presents a presymptomatic period lasting from several years to decades [1]. Early stages of AD have been identified [2–4]: mild cognitive impairment (MCI) describes subjects with objective cognitive impairment without impact on instrumental activities of daily living [5] and it is considered

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transitional between a normal cognitive state and dementia; subjective cognitive decline (SCD) is defined as a self-experienced persistent decline in cognitive capacity in comparison with the subject's previously normal status, during which the subject has normal age-, sex- and education-adjusted performance on standardized cognitive tests [6]. Studies of patients with SCD have described the evidence of amyloid load by biomarker positivity similar to that seen in AD patients [7,8]. Recent meta-analyses suggested that older people with SCD are twice as likely to develop dementia as individuals without it [9,10], leading to the hypothesis that self-perceptions of change in cognition may be considered as a very early stage 4681331, 2021, 1, Downloaded from https://onlinelibrary.wiley.com/doi/10.1111/ene.14518 by Universita Di Firenze Sistema, Wiley Online Library on [09/01/2023], See the Terms and Conditions (https://onlinelibrary.wiley.com/derns

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of AD before the detection threshold of current neuropsychological instruments is reached [11].

Amongst the symptoms of AD, sleep and circadian rhythm alterations are very common and several studies have suggested that sleep–wake cycle disturbances are associated with an increased risk of dementia and cognitive decline [12,13]. Sleep-related abnormalities have also been described in community-dwelling elderly people with subjective cognitive complaints [14] and in MCI patients [15]. Moreover, in preclinical AD, amyloid deposition seems to be associated with lower sleep quality [16]. It has been proposed that sleep–wake cycle alterations probably contribute to disease pathogenesis [17]; however, this role has to be confirmed.

The circadian system is hierarchically organized, with the central 'master' clock in the suprachiasmatic nucleus (SCN) of the hypothalamus, which synchronizes the peripheral clocks distributed in different brain regions and throughout the body, in order to coordinate physiology and behavior rhythm within a 24-h period. Both in central and in peripheral clocks, cells use a complex 'molecular machinery' composed of transcriptional-translational feedback loops to drive clock-controlled gene expression. The positive transcriptional limb of this loop is composed of the transcription factors BMAL1 (Brain Muscle ARNT-Like 1) and CLOCK, which heterodimerize and bind E-box motifs to drive the circadian transcription of clock-controlled genes, including PER1, PER2, PER3 and CRY (cryptochrome), which are part of the negative regulatory limb of the molecular machinery [18].

Current research is focusing on polymorphisms of CLOCK and PER genes to better understand their role in sleep-wake cycle alterations, psychiatric disturbances and neurodegeneration. CLOCK T3111C (rs1801260) has recently been related to the quality of aging [19] and has been reportedly associated with the circadian phenotype of eveningness (i.e. individual preference for a later sleep schedule and presenting later peaks of alertness and performance) [20]. Also PER variants have been associated with alterations of circadian phenotypes: a correlation between PER2 C111G (rs2304672) and morningness (i.e. individual preference for an earlier sleep schedule and presenting earlier diurnal peaks of alertness and performance) have been highlighted [21]; however, a recent study in a healthy Italian population did not confirm this result and did not support a role of this polymorphism in chronotypes [22]. The role of CLOCK T3111C and PER2 C111G on cognitive function has not yet been studied, in particular in early stages of cognitive decline, nor has the role of these polymorphisms on the risk of development of AD been

explored so far. The aim of our study was to evaluate the effect of *CLOCK* T3111C and *PER2* C111G polymorphisms on cognitive function and progression to AD in a population of SCD and MCI patients.

Materials and methods

Participants and clinical assessment

As part of a longitudinal, clinical-neuropsychologicalgenetic survey on SCD and MCI, 74 consecutive spontaneous subjects who self-referred to the Center for Alzheimer's Disease and Adult Cognitive Disorders of Careggi Hospital in Florence between April 1996 and May 2014 were included. All participants underwent a comprehensive family and clinical history, general and neurological examination, extensive neuropsychological investigation, estimation of premorbid intelligence and assessment of depression. A positive family history was defined as one or more first-degree relatives with documented cognitive decline. Sleep quality was assessed according to anamnestic data: those subjects who had difficulty falling asleep or woke up early or experienced frequent sleep interruptions were considered 'poor sleepers'; patients who did not report sleep disturbances were classified as 'good sleepers'. Inclusion criteria were (i) complaining of cognitive decline with a duration of ≥ 6 months; (ii) normal functioning on the activities of daily living and the instrumental activities of daily living scales [23]: (iii) unsatisfied criteria for dementia at baseline [24,25]; (iv) attainment of the clinical endpoint, i.e. conversion to AD, according to the National Institute on Aging and Alzheimer's Association (NIA-AA) criteria [25] during follow-up, regardless of follow-up duration; (v) a follow-up time of more than 2 years from the baseline visit for those patients who did not develop AD. Exclusion criteria were (i) history of head injury, current neurological and/or systemic disease, symptoms of psychosis, major depression, alcoholism or other substance abuse; (ii) complete data loss of patients' follow-up; (iii) progression to dementia other than AD.

From the initial sample, six subjects were excluded: two patients had a follow-up shorter than 2 years; two were diagnosed with psychiatric disturbances and one with frontotemporal dementia [26]; one patient received a diagnosis of vascular dementia [27]. Ultimately 68 subjects were included. All of them underwent apolipoprotein E (ApoE), *CLOCK* and *PER2* genotyping. This sample was divided into two groups: 41 subjects classified as SCD, according to the terminology proposed by the Subjective Cognitive Decline Initiative Working Group [6] (i.e. the presence of a self-experienced persistent decline in cognitive capacities with normal performance on standardized cognitive tests); 27 subjects classified as MCI according to NIA-AA criteria for the diagnosis of MCI [3] (i.e. evidence of lower performance in one or more cognitive domains with preserved independence of function in daily life). All patients underwent clinical and neuropsychological follow-up every 12 or 24 months.

On the basis of progression from SCD to AD during the follow-up, SCD subjects were classified respectively into SCD-converters (SCD-c) and SCD-nonconverters (SCD-nc). In the same way, MCI subjects were classified as MCI-converters (MCI-c) and MCInon-converters (MCI-nc).

The local ethics committee approved the protocol of the study. All participants gave written informed consent to participate.

Neuropsychological assessment

All subjects were evaluated by means of an extensive neuropsychological battery standardized and described in further detail elsewhere [28]. The battery consisted of global measurements (Mini-Mental State Examination, MMSE), tasks exploring verbal and spatial short-term memory (digit span; Corsi tapping test), verbal longterm memory (five words and paired words acquisition; recall after 10 min; recall after 24 h; Babcock Short Story immediate and delayed recall) and language (token test; category fluency task) [28]. Visuospatial abilities were also evaluated by the Rev-Osterrieth Complex Figure copy and visuospatial long-term memory was assessed by means of recall of the Rey-Osterrieth Complex Figure test [29]; attention/executive function was explored by means of dual task [30], phonemic fluency test [31] and trail making test [32]. Everyday memory was assessed by means of the Rivermead Behavioral Memory Test [33]. All raw test scores were adjusted for age, education and gender according to the correction factor reported in validation studies for the Italian population [28-33]. Premorbid intelligence was estimated by the Test di Intelligenza Breve (TIB) [34], an Italian version of the National Adult Reading Test [35]. The presence and severity of depressive symptoms were evaluated by means of the 22-item Hamilton Depression Rating Scale (HDRS) [36].

Apolipoprotein E ϵ 4, *CLOCK* T3111C and *PER2* C111G genotyping

A standard automated method (QIAcube; QIAGEN, Hilden, Germany) was used to isolate DNA from peripheral blood samples. ApoE genotypes were investigated by high resolution melting analysis [37]. Two sets of polymerase chain reaction primers were designed to amplify the regions encompassing rs7412 (NC_000019.9: g.45412079C>T) and rs429358 (NC_000019.9: g.45411941T>C). The samples with known ApoE genotypes, which had been validated by DNA sequencing, were used as standard references. The ApoE genotype was coded as ApoE ε 4– (no ApoE ε 4 alleles) and ApoE ε 4+ (presence of one or two ApoE ε 4 alleles).

The analyses of *CLOCK* and *PER2* were performed using high resolution melting analysis in order to detect the 3111T/C *CLOCK* polymorphism using primers as reported [38] and the *PER2* C111G polymorphism with the following primers: forward 5'-ACAGAAAGAGTCAAATGGGTGC-3', reverse 5'-TGTCCACATCTTCCTGCAGT-3' with annealing temperature 60°C.

Statistical analysis

Patient groups were characterized using mean and standard deviation (SD). Scores on cognitive tests were reported as z-scores (z-scores were calculated as the raw score of the patient minus the mean score of the Italian general population divided by the SD of the Italian general population). The normal distribution of the data was tested using the Kolmogorov-Smirnov test. Depending on the distribution of the data, the t test or non-parametric Mann–Whitney Utest was used for between group comparisons and Pearson's correlation coefficient or the non-parametric Spearman's ρ to evaluate correlations of between group numerical measures. The two-sided chi-squared test was used to compare categorical data and the effect size was calculated using Cohen's d for numerical measures and Cramer's V for categorical data. All statistical analyses were performed with SPSS software v.25 (SPSS Inc., Chicago, IL, USA). The significance level was set at P < 0.05.

Results

Demographic, clinical features and distribution of *CLOCK* and *PER2* genotypes

In the whole cohort, 32 of 68 subjects (47%) were *CLOCK* C carriers (29 TC, 3 CC), whilst 13 of 68 (19%) were *PER2* G carriers (13 CG, 0 GG). The genotypic distribution of the *CLOCK* and *PER2* genes in this sample was in Hardy–Weinberg equilibrium (*CLOCK* T3111C $\chi^2 = 0.91$, P > 0.05; *PER2* C111G $\chi^2 = 0.77$, P > 0.05). There were no differences in the prevalence of *CLOCK* T3111C and *PER2* C111G polymorphisms either in SCD subjects (21/41, 51.21% *CLOCK* T3111C carriers; 8/41, 19% *PER2* C111G

carriers) or in MCI subjects (11/27, 41% *CLOCK* T3111C carriers; 5/27, 19% *PER2* C111G carriers) (Fig. 1 and Table 1).

Concerning *CLOCK* T3111C, at baseline there were no statistically significant differences between C carriers and non-carriers with respect to age at onset of symptoms, age at baseline evaluation, disease duration (time from onset of symptoms and baseline evaluation), follow-up time, familiality, sex, education, TIB, MMSE, HDRS score, sleep quality and ApoE ϵ 4 allele status, not in the whole assay (Table 2) nor in SCD and MCI groups.

On the other hand, PER2 G carriers had lower premorbid intelligence scores on TIB (104.29 \pm 10.74 vs. 109.98 ± 8.06 , P = 0.049), fewer years of education $(7 \pm 3.05 \text{ vs. } 10.73 \pm 4.51, P = 0.007)$ and lower frequency of family history of AD (15.38% vs. 60%, χ^2 8.37, P = 0.004) (Table 2). When the same analysis was performed on the SCD and MCI groups apart, a statistically significant difference was found between subjects carrying the PER2 G allele and non-carriers in years of education only in the SCD group $(7.50 \pm 3.29 \text{ vs. } 12.42 \pm 4.13, P = 0.004)$; in the MCI sample, a family history of AD was more frequent in PER2 G non-carriers as none of the PER2 G carriers presented AD familiality (0% vs. 59.09%, χ^2 5.69, P = 0.017), while in the SCD group this difference was not confirmed (Table 3).

Cognitive functions and neuropsychological assessment

In the SCD group, no significant differences were found at baseline for any neuropsychological tests between *CLOCK* C carriers and non-carriers; similarly, scores on neuropsychological tests did not differ between SCD *PER2* G carriers and G non-carriers. In the MCI

 Table 1 CLOCK and PER2 genotypes in SCD and MCI patients

		SCD (41)	MCI (27)	Total (68)
CLOCK	TT	20	16	36
	TC	20	9	29
	CC	1	2	3
PER2	CC	33	22	55
	CG	8	5	13
	GG	0	0	0

MCI, mild cognitive impairment; SCD, subjective cognitive decline.

sample, *CLOCK* T3111C did not show any statistically significant influence on neuropsychological tests. On the other hand, MCI carrying *PER2* G allele had worse performances at baseline in tests assessing language (token test, P = 0.030), executive function (phonemic fluency test, P = 0.033), memory (five words acquisition, P = 0.042) and visuospatial abilities (Rey–Osterrieth Complex Figure copy, P = 0.049) (Fig. 2). In order to exclude that sleep quality could influence neuropsychological performance, the MCI sample was divided into two groups: good sleepers and poor sleepers. When scores at neuropsychological tests between the two groups were compared between these two groups, no significant differences were found (data not shown).

Effect of *CLOCK* T3111C and *PER2* C111G on progression to AD

During the follow-up, 17 patients converted to AD, 2 of 41 SCD (7.14%) and 15 of 27 MCI (55.55%). Mean conversion time to AD was 12.39 (\pm 2.28) years for SCD and 3.60 (\pm 2.55) years for MCI. A total of 39 SCD subjects did not convert to AD (SCD-nc) and their mean follow-up time was 11.74 (\pm 3.91) years (range 4.33–19.37 years, IQR 5.44 years); 12 MCI patients did not convert to AD (MCI-nc), with a





	PER2				CLOCK			
Features	G non-carriers (55)	G carriers (13)	Р	SE	C non-carriers (36)	C carriers (32)	Р	SE
Age at baseline (±SD) in years	63.72 ± 9.32	65.22 ± 7.26	0.668	-0.180	64.29 ± 9.33	63.69 ± 8.60	0.777	0.067
Age at onset $(\pm SD)$ in years	59.36 ± 10.21	62.38 ± 7.96	0.502	-0.330	60.56 ± 10.87	59.25 ± 8.64	0.694	0.133
Follow-up time (\pm SD) in years	10.79 ± 4.24	12.60 ± 4.68	0.204	-0.405	11.34 ± 4.31	10.90 ± 4.45	0.815	0.100
Disease duration (\pm SD) in years	4.36 ± 3.60	2.83 ± 2.33	0.082	0.505	3.73 ± 3.65	4.44 ± 3.18	0.195	-0.207
Sex (no. females, no. males)	38, 17	10, 3	0.577	0.068	26, 10	22, 10	0.754	0.038
Familiality (%)	60	15.38	0.004	0.351	41.66	62.5	0.086	0.086
Education in years $(\pm SD)$	10.73 ± 4.51	7 ± 3.05	0.007	0.969	9.86 ± 4.80	10.19 ± 4.21	0.737	-0.073
TIB (±SD)	109.98 ± 8.06	104.29 ± 10.74	0.049	0.599	110.30 ± 7.84	107.80 ± 9.45	0.427	0.288
MMSE (±SD)	28.47 ± 1.64	28.31 ± 1.49	0.583	0.102	28.09 ± 1.78	28.81 ± 1.33	0.064	-0.458
HDRS (±SD)	26.47 ± 4.01	27.77 ± 3.03	0.118	-0.366	26.44 ± 3.88	27.03 ± 3.86	0.506	-0.152
Poor sleepers (%)	45.09	63.63	0.264	0.142	48.48	48.27	0.987	0.002
ApoE ε4 (%)	36.36	23.07	0.362	0.110	30.55	37.5	0.546	0.073

Table 2 Demographic data of total samples in relation to PER2 and CLOCK polymorphisms

ApoE ϵ 4, apolipoprotein E; HDRS, Hamilton Depression Rating Scale; MCI, mild cognitive impairment; MMSE, Mini-Mental State Examination; SCD, subjective cognitive decline; SE, size effect; TIB, Test di Intelligenza Breve. Values indicated in the table are mean (\pm SD) or %. *P* indicates the level of significance for comparison between groups (statistical significance at *P* < 0.05, in bold characters). Age at baseline, age at onset, disease duration, follow-up and education are expressed in years. Age at baseline indicates age at the baseline evaluation; age at onset indicates age at the onset of symptoms of SCD or MCI; disease duration indicates time from onset of symptoms for MCI patients and time from the onset of subjective disturbances for SCD subjects at baseline evaluation.

Table 3 Demographic and clinical data of SCD and MCI patients: PER2 G carriers versus non-G carriers

	SCD (41)				MCI (27)			
Features	PER2 G non-carriers (33)	<i>PER2</i> G carriers (8)	Р	SE	PER2 G non-carriers (22)	<i>PER2</i> G carriers (5)	Р	SE
Age at baseline (±SD) in years	61.05 ± 9.46	63.24 ± 6.30	0.409	-0.048	67.73 ± 7.68	68.37 ± 8.27	0.928	-0.080
Age at onset $(\pm SD)$ in years	56.24 ± 10.38	60 ± 6.48	0.373	-0.088	64.05 ± 8.08	66.20 ± 9.33	0.832	-0.246
Follow-up time (\pm SD) in years	11.23 ± 3.58	14.68 ± 4.22	0.058	-0.319	10.12 ± 5.09	9.27 ± 3.45	1	0.12
Disease duration $(\pm SD)$ in years	4.80 ± 3.79	3.24 ± 2.82	0.176	0.514	3.68 ± 3.25	2.17 ± 1.23	0.377	0.856
Sex (no. females, no. males)	23, 10	7, 1	0.308	0.159	15, 7	3, 2	0.726	0.062
Familiality (%)	60.60	25	0.07	0.283	59.09	0	0.017	0.459
Education in years $(\pm SD)$	12.42 ± 4.13	7.50 ± 3.29	0.004	0.85	8.18 ± 3.87	6.20 ± 2.77	0.344	0.412
TIB (±SD)	111.09 ± 7.34	107.82 ± 8.74	0.213	0.043	107.88 ± 9.13	99.87 ± 12.60	0.203	0.112
MMSE (±SD)	28.88 ± 1.43	28.50 ± 1.77	0.640	0.019	27.80 ± 1.79	28.00 ± 1.00	1	-0.010
HDRS (±SD)	26.61 ± 3.77	27.13 ± 3.27	0.528	-0.027	26.25 ± 4.48	28.80 ± 2.58	0.097	-0.124
Poor sleepers (%)	56.66	83.33	0.221	0.204	28.57	40	0.619	0.098
ApoE ε4 (%)	33.33	25	0.650	0.071	40.9	20	0.382	0.168

ApoE ϵ 4, apolipoprotein E; HDRS, Hamilton Depression Rating Scale; MCI, mild cognitive impairment; MMSE, Mini-Mental State Examination; SCD, subjective cognitive decline; SE, size effect; TIB, Test di Intelligenza Breve. Values indicated in the table are mean (\pm SD) or %. *P* indicates level of significance for comparison between groups (statistical significance at *P* < 0.05, in bold characters). Age at baseline, age at onset, disease duration, follow-up and education are expressed in years. Age at baseline indicates age at the baseline evaluation; age at onset indicates age at the onset of symptoms of SCD or MCI; disease duration indicates time from onset of symptoms for MCI patients and time from the onset of subjective disturbances for SCD subjects at baseline evaluation.

mean follow-up time of 12.51 (±4.81) years (range 4.06–20.55 years; IQR 6.51 years). In the SCD group, no significant differences were found between SCD-c and SCD-nc in sex, familiality, disease duration, schooling, MMSE, HDRS and TIB. On the other hand, the MCI-c group was older than the MCI-nc at onset of symptoms (68.20 ± 7.48 vs. 59.75 ± 6.59, P = 0.004) and at baseline visit (71.39 ± 6.30,

P = 0.004). ApoE $\epsilon 4$ was statistically significantly more frequent in MCI-c than MCI-nc (60% vs. 8.33%, P = 0.006) subjects (Table 4).

CLOCK T3111C polymorphism prevalence did not significantly differ between converters and non-converters, both SCD and MCI. Both the SCD-c subjects presented the *PER2* G allele, while none of the SCD *PER2* G non-carriers converted to AD ($\chi^2 = 8.67$,



Figure 2 Neuropsychological assessment in SCD and MCI patients. Neuropsychological test scores are expressed as *z*-scores based on normative values. Negative values indicate worse performances than the age-matched normal population. For TMT-a, TMT-b, TMT-b-a, the higher the score, the worse the performance. DS, digit span; FWA, five words acquisition; PWA, paired words acquisition; BS and BSR, Babcock Short Story immediate and delayed recall; TT, token test; CFT, category fluency task; CT, Corsi tapping test; TMT, trail making test; RFC, Rey–Osterrieth Complex Figure copy; PFT, phonemic fluency test; RBMT, Rivermead Behavioral Memory Test. *Significant differences at P < 0.05. [Colour figure can be viewed at wileyonlinelibrary.com]

P = 0.003). There were no significant differences in the prevalence of *PER2* polymorphism between MCI-c and MCI-nc.

Discussion

This is the first study to investigate the role of *CLOCK* T3111C and *PER2* C111G polymorphisms on cognition and on the progression to AD in SCD and MCI patients.

No association between *CLOCK* T3111C and performances on neuropsychological tests and with the progression to AD was found, in either the SCD or the MCI group.

Differences between MCI PER2 G carriers and G non-carriers were detected in tests assessing language,

memory, executive functions and visuospatial abilities. The challenging question is how PER2 C111G influences cognitive functions, since the underlying mechanisms are largely unknown. Circadian rhythm and sleep-wake cycle are strictly connected with cognitive functions, which have been shown to be impaired by sleep disorders and circadian alterations [39], and this link might be found at molecular level. Clock genes are widely expressed throughout the central nervous system, in particular in the hippocampus [40] and amygdala [41] (involved in learned behaviors) and in the prefrontal and orbitofrontal cortex, in particular in Brodmann areas 11 and 47 (involved in focused attention and executive functions) [42]. During a 24-h period, the expression of clock genes exhibits oscillations which seem to be region-specific and important

	SCD (41)				MCI (27)			
Features	SCD-c (2)	SCD-nc (39)	Р	SE	MCI-c (15)	MCI-nc (12)	Р	SE
Age at baseline (±SD) in years	64.33 ± 3.54	61.33 ± 9.09	0.624	0.435	71.39 ± 6.30	63.42 ± 6.96	0.004	1.201
Age at onset $(\pm SD)$ in years	61 ± 4.24	56.77 ± 9.98	0.585	0.552	68.20 ± 7.48	59.75 ± 6.59	0.004	1.199
Follow-up time (\pm SD) in years	12.39 ± 2.28^{a}	11.74 ± 3.91	0.512	0.203	3.60 ± 2.55^{a}	12.51 ± 4.81	0.000	-2.315
Disease duration $(\pm SD)$ in years	3.33 ± 0.69	4.56 ± 3.73	0.746	-0.459	3.19 ± 3.79	3.67 ± 1.75	0.083	-0.163
Sex (females, males)	2, 0	28, 11	0.380	0.137	10, 5	8, 4	1	0.000
Familiality (%)	50%	53.84%	0.915	0.017	53.33%	41.66%	0.547	0.116
Education in years $(\pm SD)$	6.50 ± 2.12	11.72 ± 4.36	0.102	-1.523	8.47 ± 4.10	7 ± 3.19	0.373	0.400
TIB (±SD)	102.92 ± 14.24	111.09 ± 7.06	0.270	-0.727	107.67 ± 9.68	104.21 ± 10.91	0.5	0.335
MMSE (±SD)	29 ± 1.41	28.79 ± 1.50	1	0.144	28 ± 1.03	27.64 ± 2.24	1	0.206
HDRS (±SD)	26.50 ± 0.70	26.72 ± 3.74	0.790	-0.082	$27~\pm~5.24$	26.45 ± 2.73	0.727	0.132
ΑροΕ ε4 (%)	50%	30.76%	0.569	0.089	60.00%	8.33%	0.006	0.532
Poor sleepers (%)	100%	57.14%	0.230	0.197	28.57%	33.33%	0.793	0.051
PER2 C111G carriers (%)	100% (2/2)	15.38 % (6/39)	0.003	0.459	13.33% (2/15)	25% (3/12)	0.438	0.149
CLOCK T3111C carriers (%)	50% (1/2)	51.28% (20/39)	0.972	0.006	33.33% (5/15)	50% (6/12)	0.381	0.169

Table 4 Demographic, genetic and clinical data: SCD-c versus SCD-nc and MCI-c versus MCI-nc

ApoE ϵ 4, apolipoprotein E; HDRS, Hamilton Depression Rating Scale; MCI, mild cognitive impairment; MCI-c, MCI-converters; MCI-nc, MCI-non-converters; MMSE, Mini-Mental State Examination; SCD, subjective cognitive decline; SCD-c, SCD-converters; SCD-nc, SCD-non-converters; TIB, Test di Intelligenza Breve. Values indicated in the table are mean (\pm SD) or %. *P* indicates level of significance for comparison between groups (statistical significance at *P* < 0.05, in bold characters). Age at baseline, age at onset, disease duration, follow-up and education are expressed in years. Age at baseline indicates age at the baseline evaluation; age at onset indicates age at the onset of symptoms of SCD or MCI; disease duration indicates time from onset of symptoms for MCI patients and time from the onset of subjective disturbances for SCD subjects at baseline evaluation; follow-up time indicates the time from baseline visit to the last evaluation. SE indicates size effect for comparison between SCD-c and SCD-nc and between MCI-c and MCI-nc. ^aIn the SCD-c and MCI-c groups, follow-up time indicates conversion time to Alzheimer's disease.

for cognitive performance [43]: clock gene proteins and other molecules involved in long-term potentiation (LTP) present circadian oscillation in the hippocampus [44]. Studies conducted on mouse models have shown that *PER2*-mutant mice present reduced LTP and a decreased CREB phosphorylation, a critical step in the signaling pathway leading to strong LTP and certain long-term memories [45,46]. The reduced levels of p-CREB but not of total CREB due to *PER2* mutations seem to be regionally specific, since they were detected only in the hippocampus and not in the SCN, leading to the hypothesis that *PER2* has a specific role in the intracellular signaling pathways of LTP and implications for synaptic plasticity and learned behaviors [47].

Considering that clock genes encode for transcriptional factors and epigenetic modulators [48] with a role in memory and learned behaviors, it can be hypothesized that *PER2* may have implications for other cognitive functions using specific pathways not yet explored in other brain regions. In addition, it has been demonstrated that *PER2* expression oscillates in the orbitofrontal cortex, in particular in Brodmann areas 11 and 47, commonly associated with executive functions and focused attention [42].

It was also found that *PER2* G carriers presented lower premorbid intelligence scores and fewer years of education, which represent cognitive reserve proxies.

Cognitive reserve is defined as the capacity to better cope with greater amounts of cerebral damage in brighter individuals [49]. The role of cognitive reserve in the progression from SCD and MCI to AD has recently been studied [50,51]. Many investigations have suggested that both genetic and environmental factors contribute to the amount of cognitive reserve [52]. Nevertheless, no conclusive data are available about the effect of genetic factors on cognitive reserve and most studies on this topic are centered on the influence of genetic factors on synaptic density [53]. In particular, a possible influence of PER2 on cognitive reserve has not been explored yet and should be thoroughly investigated in future investigations. Furthermore, the issue of whether the effect of PER2 on cognitive reserve might be a consequence of the detrimental effect of PER2 polymorphism on cognitive functions or whether cognitive reserve might modulate the effect of PER2 polymorphism on cognitive functions would be an interesting point to explore in future studies.

Concerning sleep disturbances, no differences were detected between good and poor sleepers in the prevalence of *CLOCK* and *PER2* polymorphisms, both in the whole cohort and in the SCD and MCI groups. The role of these two polymorphisms is controversial: Katzenberg *et al.* [20] showed that both homozygous and heterozygous individuals for *CLOCK* T3111C

In conclusion, our preliminary study suggests a role of PER2 C111G in the development of cognitive reserve and cognition, while CLOCK T3111C seems not to exert any influence on this construct. Even if a higher prevalence of PER2 C111G was found in SCD subjects who converted to AD compared to patients who did not progress, the detrimental role of this polymorphism regarding the risk of progression to AD is still unclear. Further studies are needed to better explore and understand the role of genetic factors in cognition and in the development of AD from the earliest phases of the disease, in particular the influence of those genes which drive circadian rhythm and are implicated in cognitive reserve and cognitive functions.

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Disclosure of conflicts of interest

The authors declare no financial or other conflicts of interest.

Data availability statement

Data that support the findings of this study will be shared upon request from any qualified investigator.

References

1. Villemagne VL, Burnham S, Bourgeat P,, et al. Amyloid β deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. Lancet Neurol 2013; 12: 357-367.

allele had greater evening preference in comparison to TT carriers in a Caucasian cohort. Similar results were found in a Japanese population [38], while other studies did not find any association between the C allele and morning or evening preferences [23,54,55]. The role of CLOCK T3111C on circadian rhythm phenotype preference is not clear. Some authors speculated that the same polymorphism could act differently depending on the latitude in which populations live [23]. Similarly, in one report PER2 C111G was associated with alterations of circadian phenotypes, particularly with morningness [22], but another study did not confirm this result and did not support a role of this polymorphism in chronotypes [23].

Considering the conversion to AD, although the two SCD-c subjects were PER2 G carriers and none of the SCD PER2 G non-carriers converted to AD, no differences in the prevalence of PER2 polymorphism were detected in the more likely to convert MCI group. Moreover, MCI patients carrying the PER2 G allele presented a family history of AD less frequently.

PER2 gene and AD seem to be strictly connected since several studies on mouse models have shown that the expression of PER2 is abnormal both in the SCN and hippocampus: circadian oscillation of PER2 mRNA and protein in mouse SCN was disrupted by intrahippocampal injection of AB31-35 [56,57], leading to speculation that the alterations of PER2 induced by $A\beta$ may be relevant to the circadian rhythm disruption which characterizes AD [56]. Moreover, changes in the phase oscillations of clock genes like PER2 have also been observed in the brain tissue of AD patients, and these alterations could lead to a reduced synchronization of clock gene activity across brain regions in AD patients [58].

Despite these interesting earlier results, no clear correlation was found between this polymorphism and conversion to AD. In fact, the absence of a clear prevalence of PER2 G carriers in the MCI-c group and the limited number of SCD-c subjects did not allow to estimate the real contribution of both PER2 gene and PER2 C111G on the risk of progression to AD.

Indeed, the relatively small size of our cohort of patients is the first limitation of our study. For future work, the aim is to expand our sample in order to clarify our current findings and perform multivariant analysis to correct for possible confounding factors. Another limitation is the lack of AD biomarker data. Future studies including cerebrospinal fluid or neuroimaging data could provide interesting and additional information. Finally, as it is a single-center study, there may be biases with regard to assessment and diagnosis procedures.

- 2. Jack CR, Knopman DS, Jagust WJ, *et al.* Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol* 2013; **12**: 207–216.
- Albert MS, DeKosky ST, Dickson D, et al. The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging – Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 2011; 7: 270–279.
- Sperling RA, Aisen PS, Beckett LA, et al. Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging – Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 2011; 7: 280–292.
- Brodaty H, Heffernan M, Kochan NA, et al. Mild cognitive impairment in a community sample: the Sydney Memory and Ageing Study. Alzheimers Dement 2013; 9: 310–317.e1.
- Jessen F, Amariglio RE, van Boxtel M, et al. A conceptual framework for research on subjective cognitive decline in preclinical Alzheimer's disease. *Alzheimers Dement* 2014; 10: 844–852.
- Perrotin A, Mormino EC, Madison CM, Hayenga AO, Jagust WJ. Subjective cognition and amyloid deposition imaging: a Pittsburgh Compound B positron emission tomography study in normal elderly individuals. *Arch Neurol* 2012; 69: 223–229.
- Amariglio RE, Becker JA, Carmasin J, *et al.* Subjective cognitive complaints and amyloid burden in cognitively normal older individuals. *Neuropsychologia* 2012; 50: 2880–2886.
- Stewart R, Godin O, Crivello F, *et al.* Longitudinal neuroimaging correlates of subjective memory impairment: 4-year prospective community study. *Br J Psychiatry* 2011; 198: 199–205.
- Mitchell AJ, Beaumont H, Ferguson D, Yadegarfar M, Stubbs B. Risk of dementia and mild cognitive impairment in older people with subjective memory complaints: meta-analysis. *Acta Psychiatr Scand* 2014; 130: 439–451.
- Reisberg B, Gauthier S. Current evidence for subjective cognitive impairment (SCI) as the pre-mild cognitive impairment (MCI) stage of subsequently manifest Alzheimer's disease. *Int Psychogeriatr* 2008; 20: 1–16.
- Tranah GJ, Blackwell T, Stone KL, *et al.* Circadian activity rhythms and risk of incident dementia and mild cognitive impairment in older women. *Ann Neurol* 2011; **70:** 722–732.
- Lim AS, Kowgier M, Yu L, Buchman AS, Bennett DA. Sleep fragmentation and the risk of incident Alzheimer's disease and cognitive decline in older persons. *Sleep* 2013; 36: 1027–1032.
- 14. Bubbico G, Di Iorio A, Lauriola M, *et al.* Subjective cognitive decline and nighttime sleep alterations, a longitudinal analysis. *Front Aging Neurosci* 2019; **2:** 142.
- 15. Beaulieu-Bonneau S, Hudon C. Sleep disturbances in older adults with mild cognitive impairment. *Int Psychogeriatr* 2009; **21:** 654–666.
- Ju YE, McLeland JS, Toedebusch CD, et al. Sleep quality and preclinical Alzheimer disease. JAMA Neurol 2013; 70: 587–593.

- 17. Xie L, Kang H, Xu Q, *et al.* Sleep drives metabolite clearance from the adult brain. *Science* 2013; **18**: 373–377.
- Mohawk JA, Green CB, Takahashi JS. Central and peripheral circadian clocks in mammals. *Annu Rev Neurosci* 2012; 35: 445–462.
- Pagliai G, Sofi F, Dinu M, *et al.* CLOCK gene polymorphisms and quality of aging in a cohort of nonagenarians the MUGELLO study. *Sci Rep* 2019; 9: 1472.
- Katzenberg D, Young T, Finn L, et al. A CLOCK polymorphism associated with human diurnal preference. *Sleep* 1998; 15: 569–576.
- Carpen JD, Archer SN, Skene DJ, Smits M, von Schantz M. A single-nucleotide polymorphism in the 5'untranslated region of the hPER2 gene is associated with diurnal preference. J Sleep Res 2005; 14: 293–297.
- Choub A, Mancuso M, Coppedè F, et al. Clock T3111C and Per2 C111G SNPs do not influence circadian rhythmicity in healthy Italian population. *Neurol Sci* 2011; **32**: 89–93.
- Lawton MP, Brody EM. Assessment of older people: self-maintaining and instrumental activities of daily living. *Gerontologist* 1969; 9: 179–186.
- Winblad B, Palmer K, Kivipelto M, et al. Mild cognitive impairment – beyond controversies, towards a consensus: report of the International Working Group on Mild Cognitive Impairment. J Intern Med 2004; 256: 240–246.
- 25. McKhann GM, Knopman DS, Chertkow H, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement 2011; 7: 263–269.
- Neary D, Snowden JS, Gustafson L, et al. Frontotemporal lobar degeneration: a consensus on clinical diagnostic criteria. *Neurology* 1998; **51**: 1546–1554.
- Román GC, Tatemichi TK, Erkinjuntti T, *et al.* Vascular dementia: diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. *Neurology* 1993; 43: 250–260.
- Bracco L, Amaducci L, Pedone D, et al. Italian Multicentre Study on Dementia (SMID): a neuropsychological test battery for assessing Alzheimer's disease. J Psychiatr Res 1990; 24: 213–226.
- Caffarra P, Vezzadini G, Dieci F, Zonato F, Venneri A. Rey–Osterrieth complex figure: normative values in an Italian population sample. *Neurol Sci* 2002; 22: 443–447.
- Baddeley A, Della Sala S, Papagno C, Spinnler H. Dual-task performance in dysexecutive and nondysexecutive patients with a frontal lesion. *Neuropsychology* 1997; 11: 187–194.
- Spinnler H, Tognoni G. Standardizzazione e taratura italiana di test neuropsicologici: Gruppo italiano per lo studio neuropsicologico dell'invecchiamento. Milano: Masson Italia Periodici, 1987.
- Giovagnoli AR, Del Pesce M, Mascheroni S, Simoncelli M, Laiacona M, Capitani E. Trail making test: normative values from 287 normal adult controls. *Ital J Neurol Sci* 1996; 17: 305–309.
- Brazzelli M, Della Sala S, Laiacona M. Calibration of the Italian version of the Rivermead Behavioural Memory Test: a test for the ecological evaluation of memory. *Boll Psicol Appl* 1993; 206: 33–42.

- Colombo L, Sartori G, Brivio C. Stima del quoziente intellettivo tramite l'applicazione del TIB (Test Breve di Intelligenza). *G Ital Psicol.* 2002; 3: 613–637.
- 35. Nelson H. National Adult Reading Test (NART): For the Assessment of Premorbid Intelligence in Patients with Dementia: Test Manual. Windsor: NFER-Nelson, 1982.
- Hamilton M. A rating scale for depression. J Neurol Neurosurg Psychiatry. 1960; 23: 56–62.
- Sorbi S, Nacmias B, Forleo P, *et al.* ApoE allele frequencies in Italian sporadic and familial Alzheimer's disease. *Neurosci Lett* 1994; 177: 100–102.
- Mishima K, Tozawa T, Satoh K, Saitoh H, Mishima Y. The 3111T/C polymorphism of hClock is associated with evening preference and delayed sleep timing in a Japanese population sample. *Am J Med Genet B Neuropsychiatr Genet* 2005; 5: 101–104.
- Heyde I, Kiehn JT, Oster H. Mutual influence of sleep and circadian clocks on physiology and cognition. *Free Radic Biol Med* 2018; 1: 8–16.
- 40. Wakamatsu H, Yoshinobu Y, Aida R, Moriya T, Akiyama M, Shibata S. Restricted-feeding-induced anticipatory activity rhythm is associated with a phaseshift of the expression of mPer1 and mPer2 mRNA in the cerebral cortex and hippocampus but not in the suprachiasmatic nucleus of mice. *Eur J Neurosci* 2001; 13: 1190–1196.
- Lamont EW, Robinson B, Stewart J, Amir S. The central and basolateral nuclei of the amygdala exhibit opposite diurnal rhythms of expression of the clock protein Period2. *Proc Natl Acad Sci USA* 2005; **102:** 4180– 4184.
- 42. Chen CY, Logan RW, Ma T, *et al.* Effects of aging on circadian patterns of gene expression in the human prefrontal cortex. *Proc Natl Acad Sci USA* 2016; **5:** 206–211.
- Eckel-Mahan KL, Phan T, Han S, et al. Circadian oscillation of hippocampal MAPK activity and cAmp: implications for memory persistence. Nat Neurosci 2008; 11: 1074–1082.
- Chaudhury D, Wang LM, Colwell CS. Circadian regulation of hippocampal long-term potentiation. J Biol Rhythms 2005; 20: 225–236.
- Lonze BE, Ginty DD. Function and regulation of CREB family transcription factors in the nervous system. *Neuron* 2002; 35: 605–623.
- Bozon B, Kelly A, Josselyn SA, Silva AJ, Davis S, Laroche S. MAPK, CREB and zif268 are all required for

the consolidation of recognition memory. *Philos Trans* R Soc Lond B Biol Sci 2003; **358:** 805–814.

- 47. Wang LM, Dragich JM, Kudo T, *et al.* Expression of the circadian clock gene Period2 in the hippocampus: possible implications for synaptic plasticity and learned behaviour. *ASN Neuro.* 2009; **10:** e00012.
- Doi M, Hirayama J, Sassone-Corsi P. Circadian regulator CLOCK is a histone acetyltransferase. *Cell* 2006; 125: 497–508.
- Stern Y. What is cognitive reserve? Theory and research application of the reserve concept. J Int Neuropsychol Soc JINS 2002; 8: 448–460.
- Bessi V, Mazzeo S, Padiglioni S, et al. From subjective cognitive decline to Alzheimer's disease: the predictive role of neuropsychological assessment, personality traits, and cognitive reserve. A 7-year follow-up study. J Alzheimers Dis 2018; 63: 1523–1535.
- Mazzeo S, Padiglioni S, Bagnoli S, et al. The dual role of cognitive reserve in subjective cognitive decline and mild cognitive impairment: a 7-year follow-up study. J Neurol 2019; 266: 487–497.
- 52. Lee JH. Genetic evidence for cognitive reserve: variations in memory and related cognitive functions. *J Clin Exp Neuropsychol* 2003; **25:** 594–613.
- 53. Goyal MS, Raichle ME. Gene expression-based modeling of human cortical synaptic density. *Proc Natl Acad Sci USA* 2013; **110:** 6571–6576.
- 54. Robilliard DL, Archer SN, Arendt J, et al. The 3111 Clock gene polymorphism is not associated with sleep and circadian rhythmicity in phenotypically characterized human subjects. J Sleep Res 2002; 11: 305–312.
- Pedrazzoli M, Louzada FM, Pereira DS, *et al.* Clock polymorphisms and circadian rhythms phenotypes in a sample of the Brazilian population. *Chronobiol Int* 2007; 24: 1–8.
- 56. Wang X, Wang L, Yu Q, *et al.* Alterations in the expression of Per1 and Per2 induced by Aβ31-35 in the suprachiasmatic nucleus, hippocampus, and heart of C57BL/6 mouse. *Brain Res* 2016; **1642:** 51–58.
- 57. Wu M, Zhou F, Cao X, *et al.* Abnormal circadian locomotor rhythms and Per gene expression in six-monthold triple transgenic mice model of Alzheimer's disease. *Neurosci Lett* 2018; **676:** 13–18.
- Cermakian N, Lamont EW, Boudreau P, Boivin DB. Circadian clock gene expression in brain regions of Alzheimer 's disease patients and control subjects. *J Biol Rhythms* 2011; 26: 160–170.