



UNIVERSITÀ
DEGLI STUDI
FIRENZE

FLORE

Repository istituzionale dell'Università degli Studi di Firenze

In vitro production of interleukin 1 by normal and malignant human B lymphocytes.

Questa è la Versione finale referata (Post print/Accepted manuscript) della seguente pubblicazione:

Original Citation:

In vitro production of interleukin 1 by normal and malignant human B lymphocytes / PISTOIA V; F. COZZOLINO; RUBARTELLI A; TORCIA M; RONCELLA S; FERRARINI M. - In: JOURNAL OF IMMUNOLOGY. - ISSN 0022-1767. - STAMPA. - 136:(1986), pp. 1688-1692.

Availability:

This version is available at: 2158/206757 since:

Publisher:

American Association of Immunologists:9650 Rockville Pike:Bethesda, MD 20814:(301)634-7024, EMAIL:

Terms of use:

Open Access

La pubblicazione è resa disponibile sotto le norme e i termini della licenza di deposito, secondo quanto stabilito dalla Policy per l'accesso aperto dell'Università degli Studi di Firenze (<https://www.sba.unifi.it/upload/policy-oa-2016-1.pdf>)

Publisher copyright claim:

(Article begins on next page)

IN VITRO PRODUCTION OF INTERLEUKIN 1 BY NORMAL AND MALIGNANT HUMAN B LYMPHOCYTES¹

VITO PISTOIA,*[†] FEDERICO COZZOLINO,** ANNA RUBARTELLI,* MARIA TORCIA,*
SILVIO RONCELLA,* AND MANLIO FERRARINI*^{†2}

From the *Servizio di Immunologia Clinica, Istituto Nazionale per la Ricerca sul Cancro, Viale Benedetto XV, 10 - 16132 Genova; and [†]Istituto di Oncologia Clinica e Sperimentale, Università di Genova, Viale Benedetto XV, 10 - 16132 Genova; and [‡]Istituto di Clinica Medica II, Università di Firenze, Viale Pieraccini, 18 - 50139 Firenze

In this study, the capacity of normal and neoplastic B lymphocytes to release interleukin 1 (IL 1) has been investigated. Peripheral blood B cells from normal donors were isolated by depletion of E rosetting cells and by positive selection of cells expressing surface immunoglobulin (sIg) or the B1 marker. Peripheral blood B cells from patients with B cell chronic lymphocytic leukemia (B-CLL) were purified by removal of E rosetting cells followed by complement-mediated cytotoxicity with selected monoclonal antibodies. All of the normal B cell suspensions and the large majority of the B-CLL cells produced in culture high amounts of IL 1 in the absence of any apparent stimulus. Control experiments ruled out that small numbers of monocytes in the B cell suspensions could represent the source of IL 1. These data support the contention that B cells participate to the immune response as accessory cells for T cell activation not only by physically presenting antigen, but also by releasing IL 1.

In recent years, numerous studies have shown that lymphocytes produce soluble mediators generally called lymphokines (1-5). Some of these, for example interleukin 2 (IL 2), deliver signals to lymphocytes and sustain their proliferation (6). Others, such as interferon- γ , activate and potentiate effector mechanisms of immunity (7). Subpopulation studies have demonstrated that both T cells (1-6) and natural killer (NK) cells (8-14) produce lymphokines whereas little is known about the capacity of B cells to release these factors. Lack of information in this regard may be attributed to the widespread concept that the sole B cell function is that of producing specific antibodies. Consequently, most studies have been addressed to the process of B cell maturation, both at the cellular (15) and at the molecular (16) level. More recently, however, it has been shown that B lymphocytes can function as antigen-presenting cells for T lymphocytes (17-25) and consequently subserve an accessory role for

the induction of immune response.

In this study, we have investigated whether or not B cells release interleukin 1 (IL 1), a cytokine that appears to play an important role in the early steps of T cell activation and in the control of the inflammatory response (as reviewed in Reference 26). We demonstrate that peripheral blood B cells from normal individuals and from patients with B cell chronic lymphocytic leukemia (B-CLL)³ are effective producers of IL 1.

MATERIALS AND METHODS

Cell fractionation procedures. Peripheral blood mononuclear cells (MNC) from normal donors were separated on Ficoll-Hypaque (F-H) density gradients. Monocytes were partially removed by adherence. T cells were removed from the suspensions by rosetting with neuraminidase-treated sheep erythrocytes (9). Cells expressing surface immunoglobulin (sIg) were isolated by rosetting non-T cells with ox erythrocyte coated with rabbit F(ab')₂ anti-human Ig serum (27). Cells expressing the B1 marker were purified by an indirect rosetting technique. Briefly, non-T cell fractions were treated with OKM1 monoclonal antibody (Ortho Diagnostic Systems, Milano, Italy) and complement as reported (13). These non-T cells (3×10^7) were treated with 100 μ l of the B1 monoclonal (Coulter-Kontron, Milano, Italy) and were rosetted with 2 ml of 5% ox erythrocytes coated with a purified rabbit anti-mouse Ig antibody (27). After a 30-min incubation at 4°C, rosetting cells (B1⁺ cells) were separated from nonrosetting cells (B1⁻ cells) on F-H density gradients. In some experiments, monocytes were recovered from plastic after adherence by a rubber policeman.

Cells from 12 B-CLL patients were also studied. The diagnosis of B-CLL was established by clinic, morphologic, and immunologic criteria. All of the patients had 25,000 white blood cells/mm³ or more (90% lymphocytes). B-CLL MNC suspensions were depleted of E rosettes as above and treated with the OKM1 and Leu-7 monoclonal antibodies and complement (13). In some experiments, B-CLL B cells were purified as B1⁺ cells by using the technique described above.

Cell surface marker analysis. Cells with receptors for sheep erythrocytes or sIg were detected as reported (13). The murine monoclonal antibodies used in this study were the pan-T reagent OKT3 (Ortho Diagnostic System); the B cell-specific antibody B1; the large granular lymphocytes (LGL)-specific reagents Leu-7 and Leu-11b (Becton Dickinson Laboratory System, Milano, Italy); the anti-monocyte-macrophage monoclonal, which also stains LGL, OKM1; and the anti-HLA-DR monoclonal PTF 29/12 (28) that was kindly donated by Dr. G. Damiani. All of the above reagents were used in indirect immunofluorescence with a fluorescein isothiocyanate (FITC)-conjugated rabbit F(ab')₂ anti-mouse Ig. Preparations were observed with a Leitz Orthoplan fluorescence microscope. The percent of positive cells was calculated on at least 200 cells per preparation. In some experiments, cell suspensions were also stained for the cytochemical localization of α -naphthyl acetate esterase (ANAE) as reported (9).

Preparation of culture supernatants. B1⁺ cells or recovered adherent cells were cultured (1×10^6 /ml) for 48 hr in RPMI 1640 supplemented with 10% fetal calf serum (FCS) (Flow Laboratories, Milano, Italy) in 24 macro-well plates (Flow). Supernatants were harvested after centrifugation and were stored at -20°C.

³ Abbreviations used in this paper: sIg, surface immunoglobulin; B-CLL, B cell chronic lymphocytic leukemia; MNC, mononuclear cells.

Received for publication July 11, 1985.

Accepted for publication November 19, 1985.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

¹ This work was supported in part by grants from the Italian Consiglio Nazionale delle Ricerche (P. F. Oncologia) and from the Ministero della Pubblica Istruzione.

² Address reprint requests to Vito Pistoia, M.D., Servizio di Immunologia Clinica, Istituto Nazionale per la Ricerca sul Cancro, Viale Benedetto XV, 10 16132 Genova.

IL 1 assay. IL 1 activity was assayed by the conventional thymocyte assay (29). In brief, thymocytes from C3H/HeJ mice were cultured for 72 hr in 96-well flat bottomed plates (Flow) at the concentration of 1.5×10^6 /well in the presence of phytohemagglutinin (PHA; Difco, Detroit, MI) (1 μ l/ml) alone, or of PHA plus serial dilutions of the supernatant to be tested. At the end of this time, cells were pulsed with 1 μ Ci of [3 H]thymidine for 8 hr, were harvested, and were counted. One unit of IL 1 was defined as the amount of supernatant capable of doubling the [3 H]thymidine uptake by 1.5×10^6 PHA-stimulated thymocytes.

Because the presence of IL 2 in test supernatant could interfere with the results of the IL 1 assay, IL 2 activity was measured in all of the conditioned media as reported (14). Briefly, the capacity of the following cells to be stimulated was determined: a PHA-induced normal T cell line that had been expanded and kept in culture for 2 mo by adding exogenous IL 2; this line was unresponsive to PHA in the absence of any irradiated accessory cell; and the murine cell line CTLL-2. Both assays consistently failed to detect any IL 2 activity in B cell or monocyte supernatants.

Molecular weight determination of IL 1. To 80 ml of normal B cell or monocyte culture supernatants $(\text{NH}_4)_2\text{SO}_4$ was added to a final concentration of 75%. The precipitate was centrifuged, was resuspended in phosphate-buffered saline containing 50 μ g/ml gentamicin (PBS), and was dialyzed extensively against the same buffer. Aliquots of 3 ml were then gel filtered on a Sephadex G-75 Superfine (Pharmacia) column equilibrated with PBS and were calibrated with the following markers: aldolase (158,000 m.w.), human serum albumin (69,000 m.w.), isolated Ig light chains (23,000 m.w.), and cytochrome c (12,300 m.w.). A flow rate of 10 ml/hr was used, and 2-ml fractions were collected. Aliquots of the fractions were 0.22 μ m filtered and were assayed for IL 1 activity as described above.

RESULTS

In initial studies, normal B cells were isolated from the non-T cell fraction of peripheral blood by rosetting with ox erythrocytes coated with anti-Ig antibodies (Table I). slg^+ cells produced large amounts of IL 1 (Table II), but monocyte contamination of these cell populations was

TABLE I
Surface phenotype of normal and B-CLL-enriched B cell suspensions

Surface Marker	Percent of Positive Cells ^a		
	slg^+ cells ^b	B1^+ cells ^c	B-CLL B cells ^d
slg	92 \pm 5 ^e	95 \pm 4	N.D. ^f
B1	80 \pm 6	96 \pm 3 ^e	95 \pm 3
HLA-DR	95 \pm 3	98 \pm 2	98 \pm 5
OKT3	<1	<1	<1
OKM1	15 \pm 3	<2	<1
Leu-11	4 \pm 1	<1	<2
Leu-7	2 \pm 0.3	<1	<1

^a Mean \pm SE from three different experiments. The SE is calculated as the square root of the pooled variance divided by the overall number of samples. The homogeneity of variance between experiments was demonstrated, and no significant difference between the means was documented.

^b B cells were isolated by direct rosetting according to the expression of slg (see *Materials and Methods*).

^c B cells were isolated by indirect rosetting according to the expression of B1 antigen (see *Materials and Methods*).

^d B-CLL B cells were obtained by depletion of E rosetting cells followed by treatment with OKM1 and Leu-7 monoclonal antibodies plus complement.

^e Detected by rosetting during the separation procedures.

^f Not done because of the low-density expression of slg by B-CLL B cells.

TABLE II
IL 1 production by cell suspensions enriched for B cells and monocytes

Supernatants from	IL 1 Activity ^a (U/ml)		
	Expt. 1	Expt. 2	Expt. 3
slg^+ cells	130	116	110
B1^+ cells	129	118	98
Autologous monocytes ^b	78	84	79

^a Supernatants were tested at different dilutions, and the reported values are relative to those yielding the highest activity. IL 1 produced by 1×10^6 cells cultured for 48 hr.

^b Monocytes were recovered from plastic after adherence.

too high to indicate conclusively that B cells could be the source for the detectable cytokine (Table I).

To overcome these difficulties, the following purification procedure was used: peripheral blood non-T cells partially depleted of monocytes by adherence to plastic were treated with OKM1 antibody and complement. The remaining cells were subsequently fractionated into B1^+ and B1^- cells by an indirect rosetting technique. The B1^+ cell suspensions contained 95% or more B cells (as detected by staining for slg), thus being considerably less contaminated than the slg^+ cell preparations (Table I). The reasons for the improved B cell purification are depletion of OKM1⁺ monocytes by complement-mediated cytotoxicity and failure of the B1 monoclonal antibody to bind to monocytes, as opposed to the anti-Ig reagent, which consistently reacted with slg molecules passively absorbed on a proportion of monocytes. The capacity of B1^+ cells to produce IL 1 was tested and was compared with that of autologous monocytes recovered after adherence to plastic surfaces (Table II). These adherent cells contained 90% \pm 5% monocytes, as determined by reactivity with the OKM1 monoclonal antibody and by the characteristic pattern of ANAE staining. Minor contaminants were represented by B cells (4%), T cells (2%), and LGL (2%). B1^+ cells released IL 1 in quantities that, on a per cell basis, were comparable with, or higher than, those produced by monocytes (Table II). This finding makes it unlikely that the few monocytes present in the B1^+ cell suspensions were responsible for IL 1 production. The possibility that B cells released IL 1 as a consequence of endotoxin stimulation was ruled out by the observation that the batch of FCS used had a very low endotoxin content, as assessed by the Limulus assay. Furthermore, B cells cultured in medium containing 1% human serum albumin in the place of FCS retained the ability to produce IL 1.

B cells from 12 B-CLL patients were also tested for IL 1 release (Table III). B cells were isolated from peripheral blood MNC by removing E rosettes and subsequently treating the suspensions with the OKM1 and Leu-7 monoclonal antibodies and complement. The last treatment was necessary, because these suspensions still contained 4% \pm 1 OKM1⁺ and 3% \pm 1 Leu-7⁺ cells. After this procedure, the suspensions were enriched for B cells up to 95% or greater (Table I). In some cases, B cells were purified by enrichment for B1^+ cells by using rosette methods, and results were virtually identical. Nine of the 12 cases studied (75%) produced IL 1 in amounts that

TABLE III
IL 1 production by B cells from B-CLL patients

Patient	IL 1 ^a (U/ml)
A	160
B	0
C	0
D	135
E	42
F	0
G	93
H	52
I	82
L	32
M	21
N	12

^a Supernatants were tested at different dilutions, and the reported values are relative to those yielding the highest activity. IL 1 produced by 1×10^6 cells cultured for 48 hr.

were comparable with those of normal B cells (Table III). Interestingly, neither normal nor malignant B cells released IL 2, measured by the support of IL 2-dependent T cell growth. Thus, the IL 1 activity detected by the thymocyte assay could not be due to the presence of IL 2 (data not shown). Normal B cell supernatants were ammonium sulphate precipitated and were subsequently gel filtered over a Sephadex G75 column. Figure 1 shows the elution pattern of the B cell-derived factor with IL 1 activity, which is superimposable to that released by normal monocytes. As evident, the apparent m.w. of both molecules is in the range between 15,000 and 20,000.

DISCUSSION

Our study demonstrates that peripheral blood B cells from both normal donors and B-CLL patients produce IL 1. Normal and malignant B cells apparently released IL 1 constitutively in the absence of any stimulation. On the basis of m.w. determination (15,000 to 20,000), the available data suggest that this factor could be similar to that released by monocytes. The possibility exists that cells were induced to release IL 1 by contact with FCS or even plastic tissue culture ware. Alternatively, IL 1 may have been produced by B cells preactivated *in vivo* (30). While this manuscript was in preparation, Matsushima *et al.* reported similar data (31).

The results reported here were obtained by using monoclonal antibodies and rosetting techniques for cell fractionation. Normal B cells were isolated from peripheral blood non-T cells according to the expression of sIg or of the B1 marker. B cell enrichment within the B1⁺ fraction proved by far superior to that detected in sIg⁺ cells, which contained relevant proportions of monocytes. In addition, B cell suspensions could be additionally purified by treatment with anti-monocyte monoclonal antibodies and complement. In theory, it cannot be excluded that the

separation procedures used in these experiments select a small subset of slightly adherent, OKM1⁻, Fc receptor positive cells, which are highly efficient IL 1 producers and co-purify with B cells through nonspecific binding of the B1 monoclonal antibody or of antibody coated ox erythrocytes. Furthermore, Fc receptor interaction with ox erythrocytes could deliver an effective signal for cytokine release. However, it is unlikely that IL 1 production was attributable to contaminant monocytes for the following reasons: the high degree of purity for the normal B cell fractions used; the finding that on a per cell basis, B cell-enriched suspensions were as efficient IL 1 producers as autologous monocytes, and the observation that CLL B cells purified by negative selection methods (*i.e.*, removal of E rosetting cells followed by killing of residual non-B cells with monoclonal antibodies and complement) produced IL 1 in amounts comparable with those of normal B cell suspensions. Nonetheless, because a small contaminant of monocytes was still present in the B cell suspensions, the possibility that monocytes cooperate with B cells in inducing IL 1 release by the latter cells cannot be dismissed.

We have not yet observed a relationship between IL 1 production and other characteristics of the single B-CLL cases, such as lymphocyte counts, treatment, duration, or stage of disease. Previously we have shown that individual B-CLL clones reach different maturational stages *in vivo*, which can be assessed on the basis of the capacity of the cells to produce and secrete Ig molecules and of their responsiveness to growth and differentiation factors (32, 33). When these parameters were used to determine the maturational stages of the B-CLL cases studied, no correlation was found between the degree of maturation of the malignant B cells and IL 1 production. Whether or not B-CLL cells that release IL 1 *in vitro* also produce the same cytokine *in vivo* is presently unknown; addi-

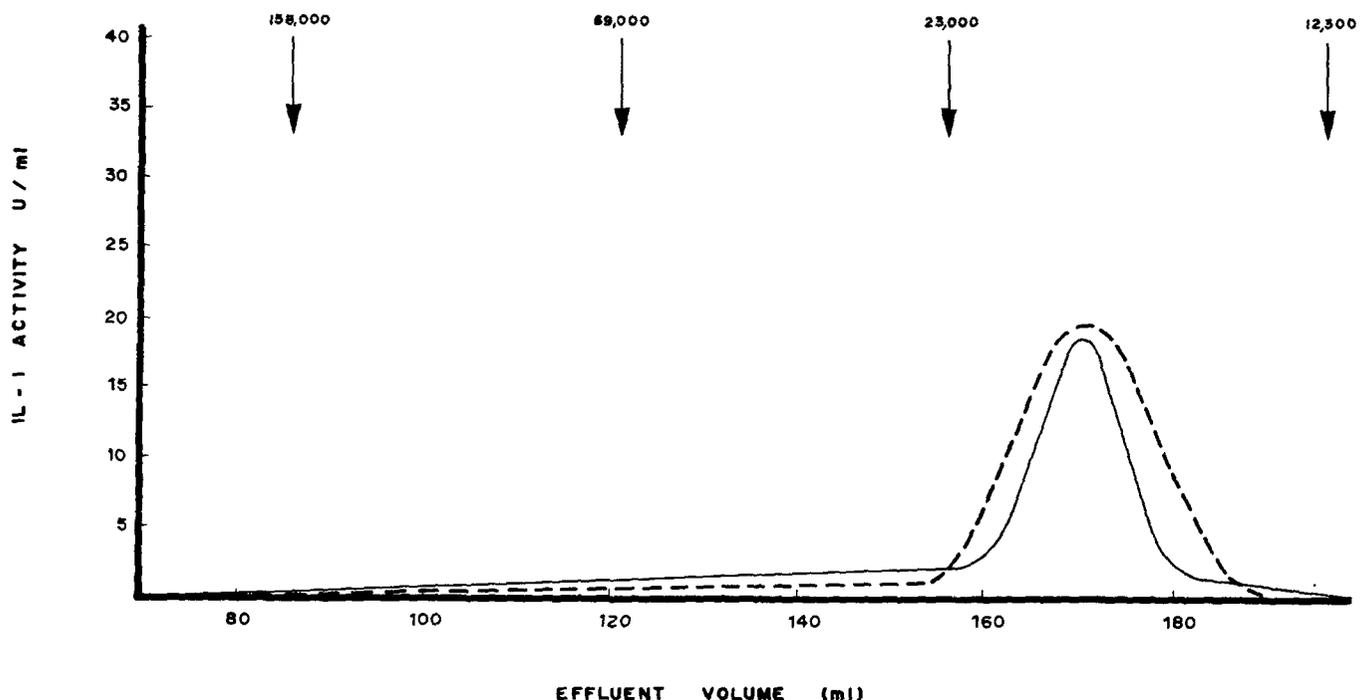


Figure 1. Elution patterns of normal B cell-derived (—) or normal monocyte-derived (---) factors with IL 1 activity. Culture supernatants were ammonium sulphate precipitated, and precipitates were applied to a Sephadex G-75 column; fractions were collected, and aliquots of each fraction were assayed for IL 1 activity.

tional studies on serum and urine (34) from these patients are now in progress.

Several explanations for the heterogeneity in IL 1 production by B-CLL cells can be considered. First, IL 1 production could be restricted to a B cell subset of which the lymphokine producing B-CLL cases represent the counterpart. Second, IL 1 may be released at a particular B cell maturational stage that cannot yet be defined with the available methods. Finally, all B-CLL cells could produce IL 1, but in some cases, this lymphokine could be absorbed on the malignant cells. This possibility is suggested by the following observations: IL 1 can affect both proliferation and differentiation of B cells (35-40), and some Epstein Barr virus-infected B cell lines produce IL 1, which can, at least in part, maintain their proliferative capacity (41).

The demonstration that B cells release IL 1 is in accordance with their capacity to present antigens (17-25) and reinforces the view that B cells function as accessory cells for T cell proliferation. Finally, in view of the role that IL 1 plays for the induction and/or maintenance of inflammatory reaction (26), our data suggest a novel B cell function in this particular feature of the response to pathogens. In this connection, it is of note that we have preliminarily shown that both normal and malignant B cells are also capable of producing considerable amounts of granulocyte-macrophage colony stimulating factor (Pistoia et al., manuscript in preparation).

Acknowledgments. We thank Drs. R. Adami and P. Strada for the supply of normal blood samples; Dr. A. Marmont and his group for providing blood samples from B-CLL patients; Dr. G. Damiani for the PTF 29/12 monoclonal antibody; Drs. Loran Clement, Carlo Grossi, Arabella Tilden and Kenneth Zuckerman for critical reading of the manuscript; Ms. Jane Hamner for secretarial help. Peripheral blood samples from B-CLL patients were obtained after informed consent. This study has been approved by the Authorities of our Universities.

REFERENCES

- Morgan, D. A., F. W. Ruscetti, and R. C. Gallo. 1976. Selective in vitro growth of T lymphocytes from normal human bone marrow. *Science* 193:1007.
- Howard, M., J. Farrar, M. Hilfiker, B. Johnson, K. Takatsu, T. Hamaoka, and W. E. Paul. 1982. Identification of a T-cell derived B-cell growth factor distinct from interleukin 2. *J. Exp. Med.* 155:914.
- Nathan, D. G., L. Chess, D. G. Hillman, B. Clark, J. Breard, E. Merler, and D. E. Housman. 1978. Human erythroid burst-forming unit: T cell requirement for proliferation in vitro. *J. Exp. Med.* 147:324.
- Cline, M. J., and D. W. Golde. 1979. Cellular interactions in hematopoiesis. *Nature* 277:177.
- Green, J. A., S. R. Cooperband, and S. Kibrick. 1969. Immune specific induction of interferon production in cultures of human blood lymphocytes. *Science* 164:1415.
- Watson, J., and D. Mochizuki. 1980. Interleukin 2: a class of T cell growth factors. *Immunol. Rev.* 51:257.
- Preble, O. T., and R. M. Friedman. 1983. Biology of disease. Interferon-induced alterations in cells: relevance to viral and non-viral diseases. *Lab. Invest.* 49:1.
- Trinchieri, G., and D. Santoli. 1978. Anti-viral activity induced by culturing lymphocytes with tumor-derived or virus-transformed cells. Enhancement of human natural killer activity by interferon and antagonistic inhibition of susceptibility of target cells to lysis. *J. Exp. Med.* 147:1314.
- Pistoia, V., A. Nocera, R. Ghio, A. Leprini, A. Perata, M. Pistone, and M. Ferrarini. 1983. PHA-induced human T cell colony formation: enhancing effect of large granular lymphocytes. *Exp. Hematol.* 11:251.
- Timonen, T., J. R. Ortaldo, and R. B. Herberman. 1981. Characterization of human large granular lymphocytes and relationship to natural killer and K cells. *J. Exp. Med.* 153:569.
- Kasahara, T., J. Y. Djeu, S. F. Dougherty, and J. J. Oppenheim. 1983. Capacity of human large granular lymphocytes (LGL) to produce multiple lymphokines: interleukin 2, interferon, and colony-stimulating factor. *J. Immunol.* 131:2379.
- Scala, G., P. Allavena, J. Y. Djeu, T. Kasahara, J. R. Ortaldo, R. B. Herberman, and J. J. Oppenheim. 1984. Human large granular lymphocytes (LGL) are potent producers of interleukin 1. *Nature* 309:56.
- Pistoia, V., R. Ghio, A. Nocera, A. Leprini, A. Perata, and M. Ferrarini. 1985. Large granular lymphocytes have a promoting activity on human peripheral blood erythroid burst-forming units. *Blood* 65:464.
- Pistoia, V., F. Cozzolino, M. Torcia, E. Castigli, and M. Ferrarini. 1985. Production of B cell growth factor by a Leu-7⁺, OKM1⁺ non-T cell with the features of large granular lymphocytes (LGL). *J. Immunol.* 134:3179.
- Calvert, J. E., S. Maruyama, T. F. Tedder, C. F. Webb, and M. D. Cooper. 1984. Cellular events in the differentiation of antibody-secreting cells. *Sem. Hematol.* 21:226.
- Honjo, T. 1983. Immunoglobulin genes. *Ann. Rev. Immunol.* 1:499.
- Chesnut, R. W., and H. M. Grey. 1981. Studies on the capacity of B cells to serve as antigen-presenting cells. *J. Immunol.* 126:1075.
- Glimcher, L. H., K. J. Kim, I. Green, and W. E. Paul. 1982. Ia-antigen bearing B cell tumor lines can present protein antigen and alloantigen in a major histocompatibility complex-restricted fashion to antigen-reactive T cells. *J. Exp. Med.* 155:455.
- Issekutz, T., E. Chu, and R. S. Geha. 1982. Antigen presentation by human B cells: T cell proliferation induced by Epstein Barr virus B lymphoblastoid cells. *J. Immunol.* 129:1446.
- Chesnut, R. W., S. M. Colon, and H. M. Grey. 1982. Antigen presentation by normal B cells, B cell tumors and macrophages: functional and biochemical comparison. *J. Immunol.* 128:1764.
- Chesnut, R. W., S. M. Colon, and H. M. Grey. 1982. Requirements for the processing of antigens by antigen-presenting B cells. I. Functional comparison of B cell tumors and macrophages. *J. Immunol.* 129:2382.
- Grey, H. M., S. M. Colon, and R. W. Chesnut. 1982. Requirement for the processing of antigen-presenting B cells. II. Biochemical comparison of the fate of antigen in B cell tumor and macrophages. *J. Immunol.* 129:2389.
- Walker, E. B., L. L. Lanier, and N. L. Warner. 1982. Characterization and functional properties of tumor cell lines in accessory cell replacement assays. *J. Immunol.* 128:802.
- Kakiuchi, T., R. W. Chesnut, and H. M. Grey. 1983. B cells and antigen presenting cells: the requirements for B cell activation. *J. Immunol.* 131:109.
- Lanzavecchia, A. 1985. Antigen-specific interaction between T and B cells. *Nature* 3134:537.
- Durum, S. K., J. A. Schmidt, and J. J. Oppenheim. 1985. Interleukin 1: an immunological perspective. *Ann. Rev. Immunol.* 3:263.
- Ling, N. R., S. Bishop, and R. Jefferis. 1977. Use of antibody-coated red cells for the sensitive detection of antigen and in rosette tests for cells bearing surface immunoglobulin. *J. Immunol. Methods* 15:279.
- Corte, G., G. Damiani, F. Calabi, M. Fabbi, and A. Bargellesi. 1981. Analysis of HLA-DR polymorphism by two dimensional peptide mapping. *Proc. Natl. Acad. Sci. USA* 78:534.
- Mizel, S. B., B. L. Rosenstreich, and J. J. Oppenheim. 1978. Phorbol myristic acetate stimulates LAF production by the macrophage cell line P 388D1. *Cell Immunol.* 40:230.
- Kehrl, J., and A. S. Fauci. 1983. Identification, purification and characterization of antigen-activated and antigen-specific human B lymphocytes. *J. Exp. Med.* 157:1692.
- Matsushima, K., A. Procopio, H. Abe, G. Scala, J. R. Ortaldo, and J. J. Oppenheim. 1985. Production of interleukin 1 activity by normal peripheral blood lymphocytes. *J. Immunol.* 135:1132.
- Rubartelli, A., R. Sitia, A. Zicca, C. E. Grossi, and M. Ferrarini. 1983. Differentiation of chronic lymphocytic leukemia cells: correlation between the synthesis and secretion of immunoglobulins and the ultrastructure of the malignant cells. *Blood* 62:495.
- Rubartelli, A., R. Sitia, C. E. Grossi, and M. Ferrarini. 1985. Maturation of chronic lymphocytic leukemia B cells: correlation between the capacity of responding to T cell factors in vitro and the stage of maturation reached in vivo. *Clin. Immunol. Immunopathol.* 34:296.
- Kimball, E. S., S. F. Pickeral, J. J. Oppenheim, and J. L. Rossio. 1984. Interleukin 1 activity in normal human urine. *J. Immunol.* 133:256.
- Howard, M., S. B. Mizel, L. Lachman, J. Ansel, B. Johnson, and W. E. Paul. 1983. Role of interleukin 1 in anti immunoglobulin induced B cell proliferation. *J. Exp. Med.* 157:1529.
- Falkoff, R. J., A. Muraguchi, J. X. Hong, J. L. Butler, C. A. Dinarello, and A. S. Fauci. 1983. The effects of interleukin 1 on human B cell activation and proliferation. *J. Immunol.* 131:801.
- Booth, R. J., and J. D. Watson. 1984. Interleukin 1 induces proliferation in two distinct B cell subpopulations responsive to two different murine B cell growth factors. *J. Immunol.* 133:1346.
- Giri, J. G., and P. W. Kincaid. 1984. Interleukin 1-mediated induction of κ -light chain synthesis and surface immunoglobulin expression on pre-B cells. *J. Immunol.* 133:223.

39. Lipsky, P. E., P. A. Thompson, L. J. Rosenwasser, and C. A. Dinarello. 1983. The role of interleukin 1 in human B cell activation: inhibition of B cell proliferation and the generation of immunoglobulin-secreting cells by an antibody against human leukocytic pyrogen. *J. Immunol.* 130:2708.
40. Falkoff, R. J., J. L. Butler, C. A. Dinarello, and A. S. Fauci. 1984. Direct effects of a monoclonal B cell differentiation factor and of purified interleukin 1 on B cell differentiation. *J. Immunol.* 133:692.
41. Scala, G., Y. D. Kuang, R. E. Hall, A. B. Muchmore, and J. J. Oppenheim. 1984. Accessory cell function of human B cells. I. Production of both interleukin 1-like activity and an interleukin 1 inhibitory factor by an EBV-transformed human B cell line. *J. Exp. Med.* 159:1637.